

Development and Antifungal Efficacy of Hydrocarbon-Stapled Peptide Analogs Based on Human Antimicrobial Peptides for Candidiasis Therapy



Hyosuk Yun, Ganbaatar Byambasuren, and Chul Won Lee*

Department of Chemistry, Chonnam National University, Gwangju 61186, South Korea

*Corresponding author : cwlee@jnu.ac.kr

Abstract

KR-12, derived from the human antimicrobial peptide LL-37, maintains the antimicrobial properties of LL-37 while minimizing human cell toxicity. The KR-12 analog, named KR-12-pa, is characterized by its amphipathic α -helical structure, contributing to its strong antimicrobial and anti-inflammatory effects. In this study, we designed hydrocarbon-stapled peptide analogs based on KR-12-pa incorporating chiral non-natural amino acids at positions *i* and *i*+4. All stapled peptides were synthesized by solid-phase peptide synthesis method, verified by LC-MS. The stapled peptides showed broad-spectrum antifungal activities, with minimum inhibitory concentrations (MIC) ranging from 2 to 32 $\mu\text{g/mL}$, against pathogenic *Candida* strains including the fluconazole-resistant *Candida krusei* strain. Notably, a subset of these analogs, specifically S3, S5, and S7, demonstrated low human cell toxicity comparable to the original KR-12 peptide. Among these, S3 stood out for its balance of potent antifungal activity and minimal toxicity, making it a promising candidate for further development.

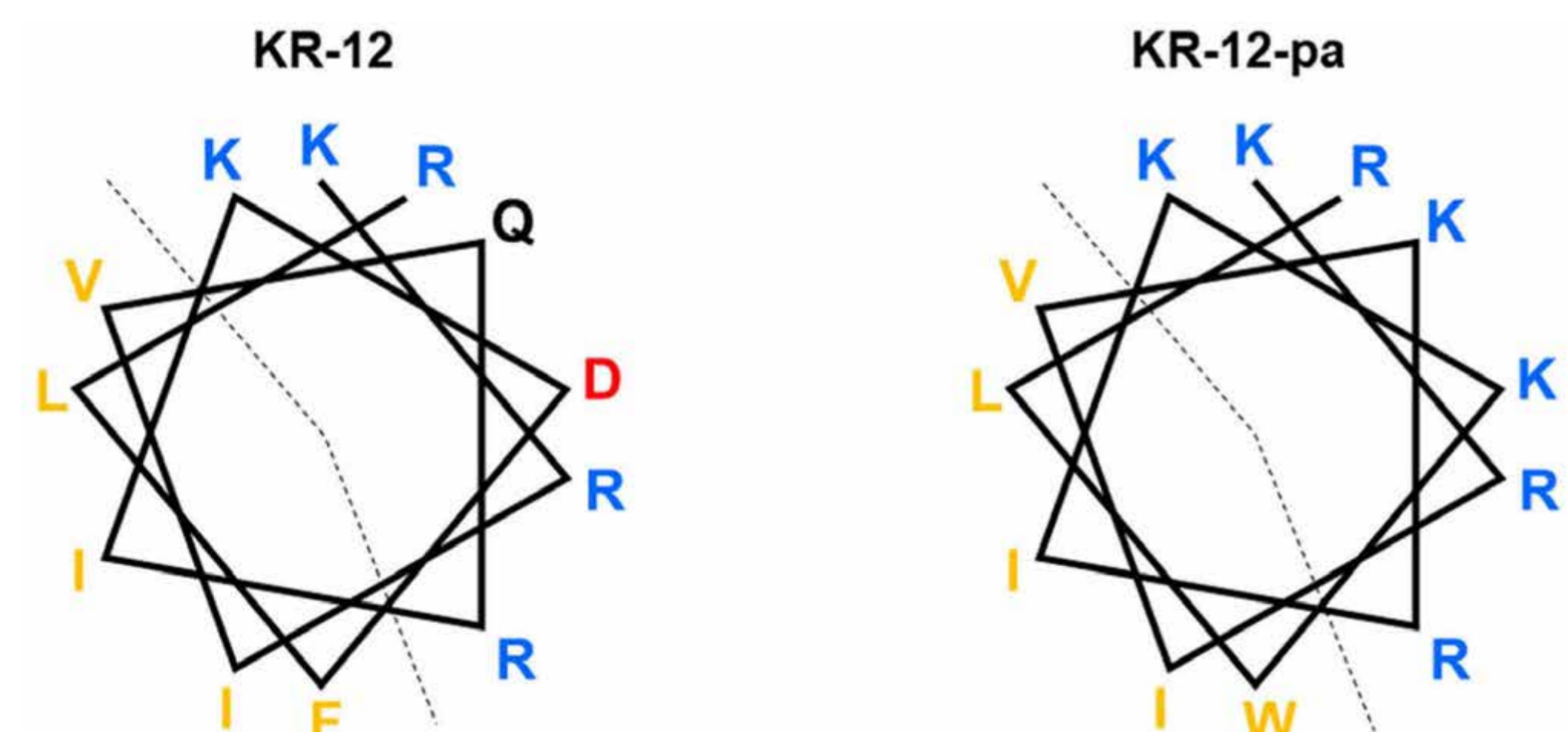
Further structural analysis through circular dichroism indicated that S3 maintains an α -helical conformation in buffer solutions, differing from the wild-type peptide. Preliminary *in vivo* assessments using the *Galleria mellonella* model underscored S3's reduced toxicity and heightened antifungal efficacy. Altogether, these results position the S3 stapled analog of KR-12-pa as a viable and effective option for treating candidiasis, highlighting its potential as a therapeutic innovation.

Introduction

Antimicrobial peptides (AMPs) are an important component of the innate immune system and are produced by almost all living organisms, from plants and insects to mammals, including humans. The ability to physically destroy bacterial cell membranes and induce lysis allows AMPs to target bacteria, fungi, and enveloped viruses and renders them a potential alternative to conventional antibiotics. LL-37 is a natural antimicrobial peptide expressed in the human body.

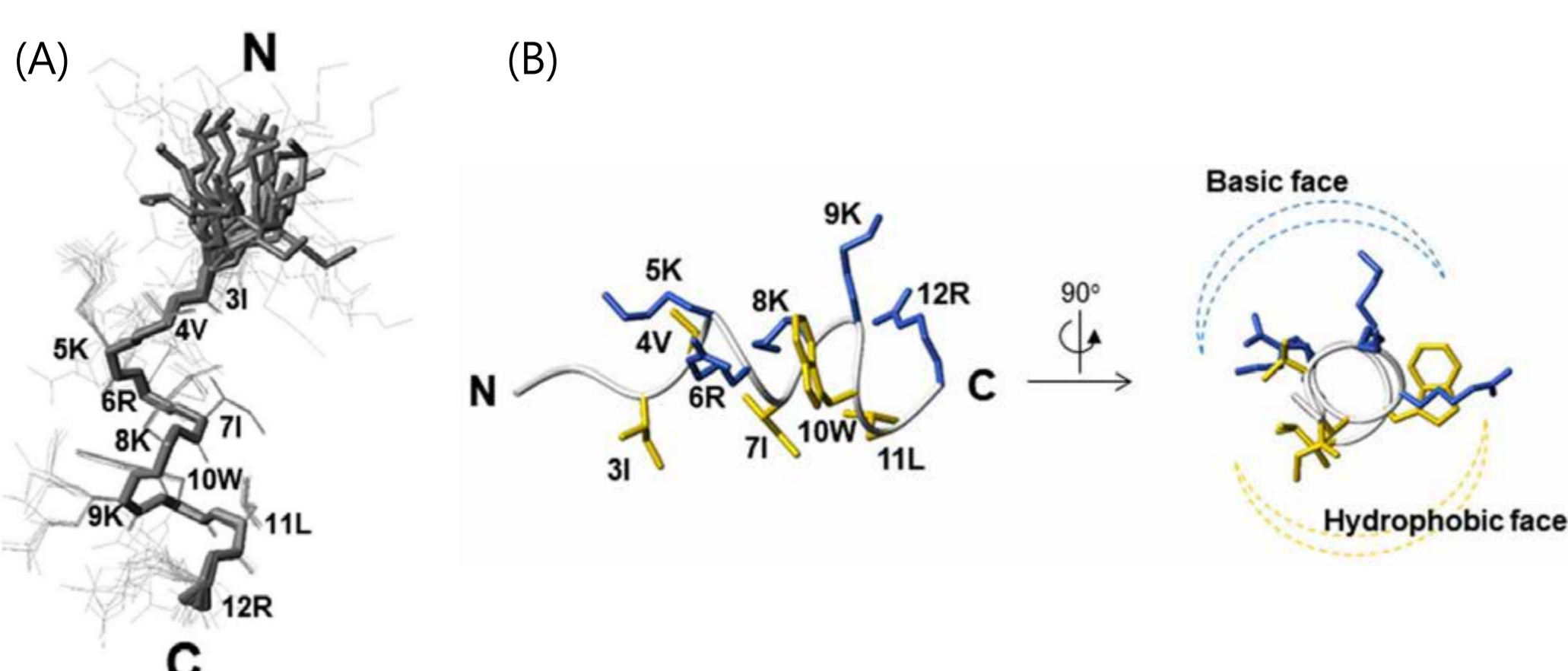
KR-12-pa

LL-37 LLGDFFRKSKEKIGKEFKRIVQRIKDFLRNQLVPTES
 KR-12 LLGDFFRKSKEKIGKEFKRIVQRIKDFLRNQLVPTES
 KR-12-pa LLGDFFRKSKEKIGKEFKRIVKRKKFLRNQLVPTES



- ▶ Primary sequences and helical wheel diagrams of LL-37, KR-12, and KR-12-pa. Positively charged residues are in blue
- ▶ Negatively charged residues are in red. Hydrophobic residues are in yellow

NMR structure of KR-12-pa



- (A) Stereo view of twenty converged line structures of KR-12-pa for backbone heavy atoms
- (B) The lowest target function structure of KR-12-pa

Results

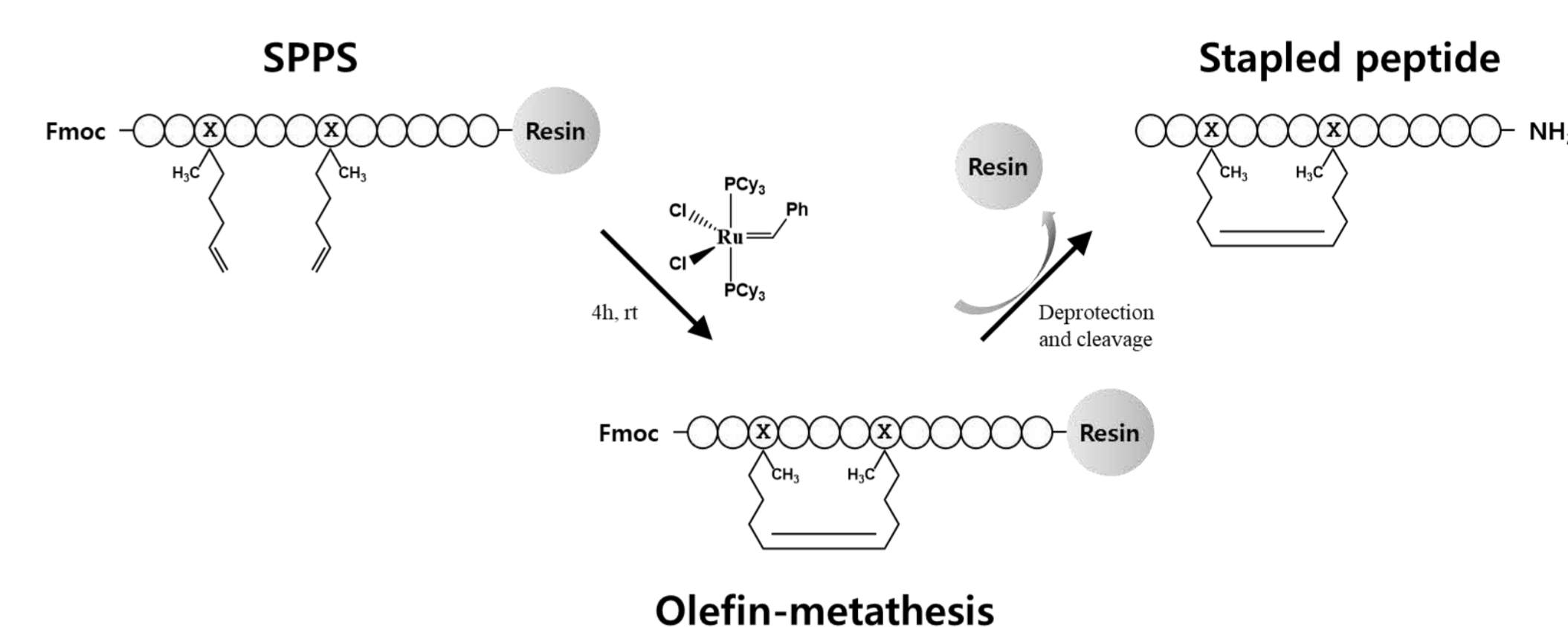
Design and synthesis of stapled peptide analogs based on KR-12-pa

Design of stapled peptides

	Sequence	MW	
		Linear	Stapled
No stapled	WT KRIVKRIKKWLR-NH ₂	1623.10	-
Single stapled (i, i+4)	S1 XRIVXRIKKWLR-NH ₂	1645.14	1617.08
	S2 KXIVKXIKKWLR-NH ₂	1589.11	1561.01
	S3 KRXXRXXKWLR-NH ₂	1675.17	1647.11
	S4 KRXXRXXKWLR-NH ₂	1674.18	1646.12
	S5 KRIVXRIKKWLR-NH ₂	1645.14	1617.08
	S6 KRIVKXIKKWLR-NH ₂	1559.08	1531.03
	S7 KRIVKXXKXWLR-NH ₂	1675.17	1647.11
	S8 KRIVKRIKKWLR-NH ₂	1617.12	1589.07

X : (S)-2-(4-pentenyl)Ala-OH

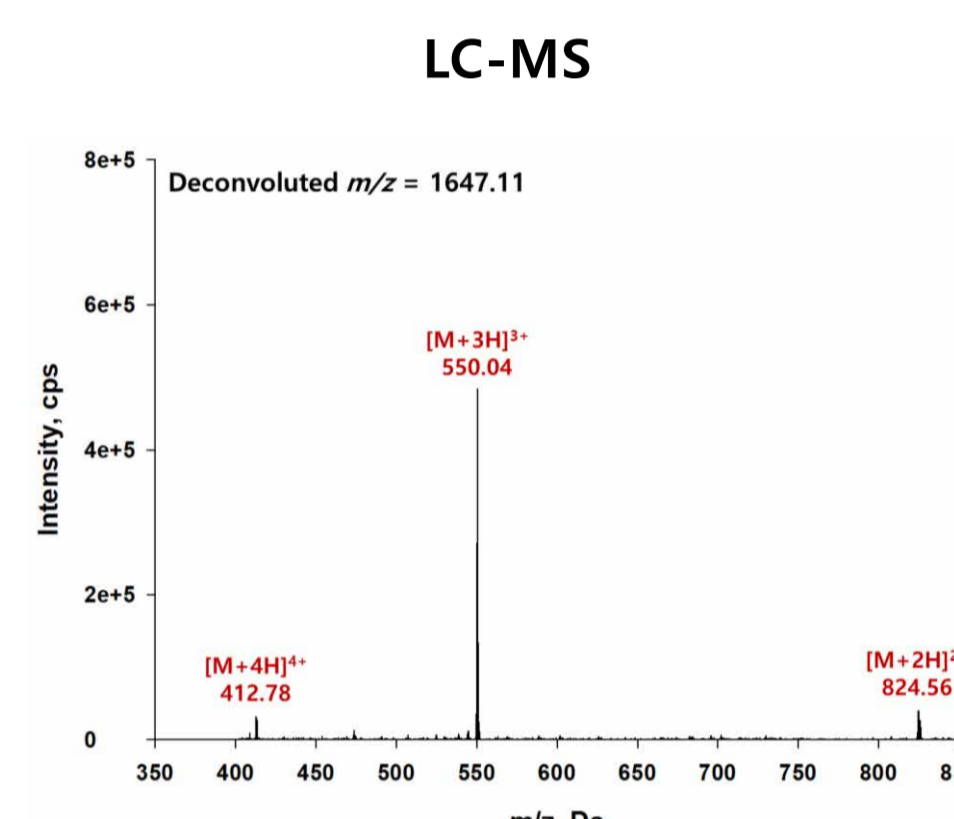
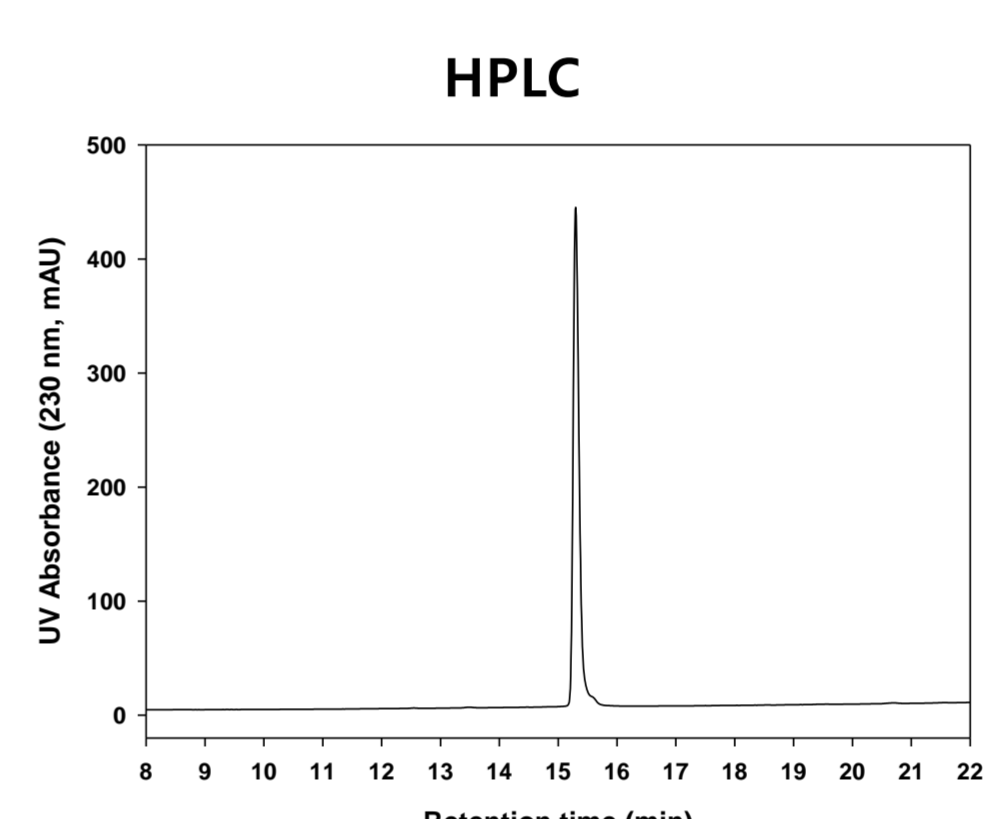
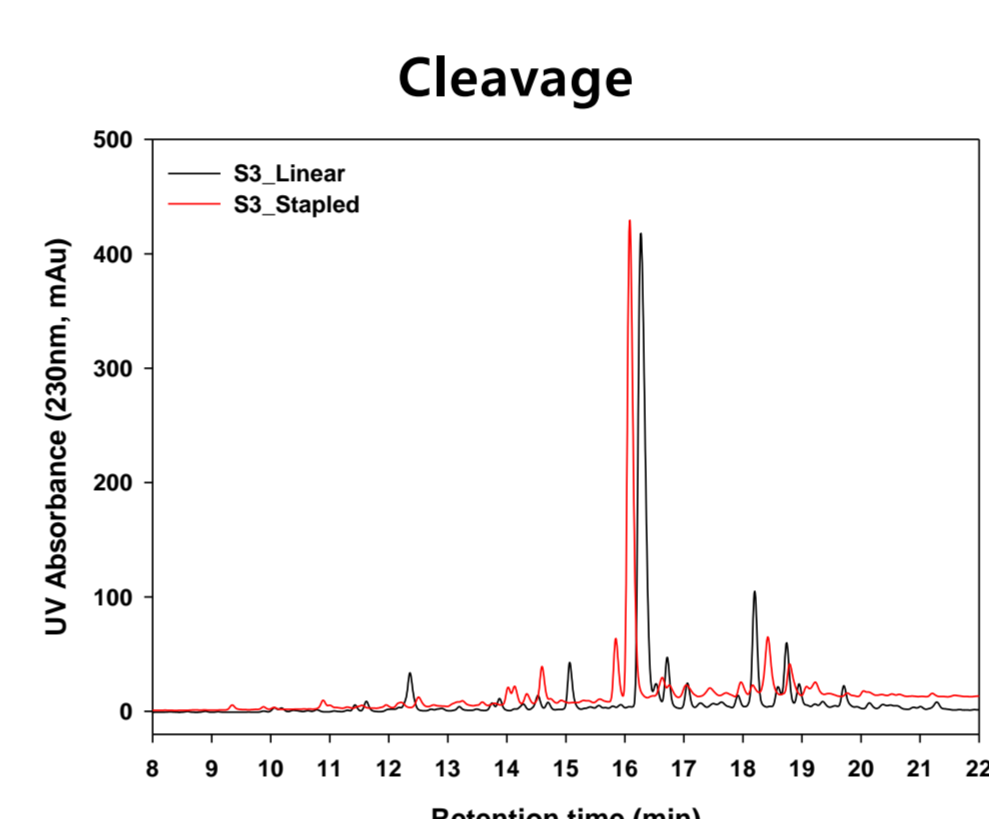
Synthetic scheme of stapled peptides



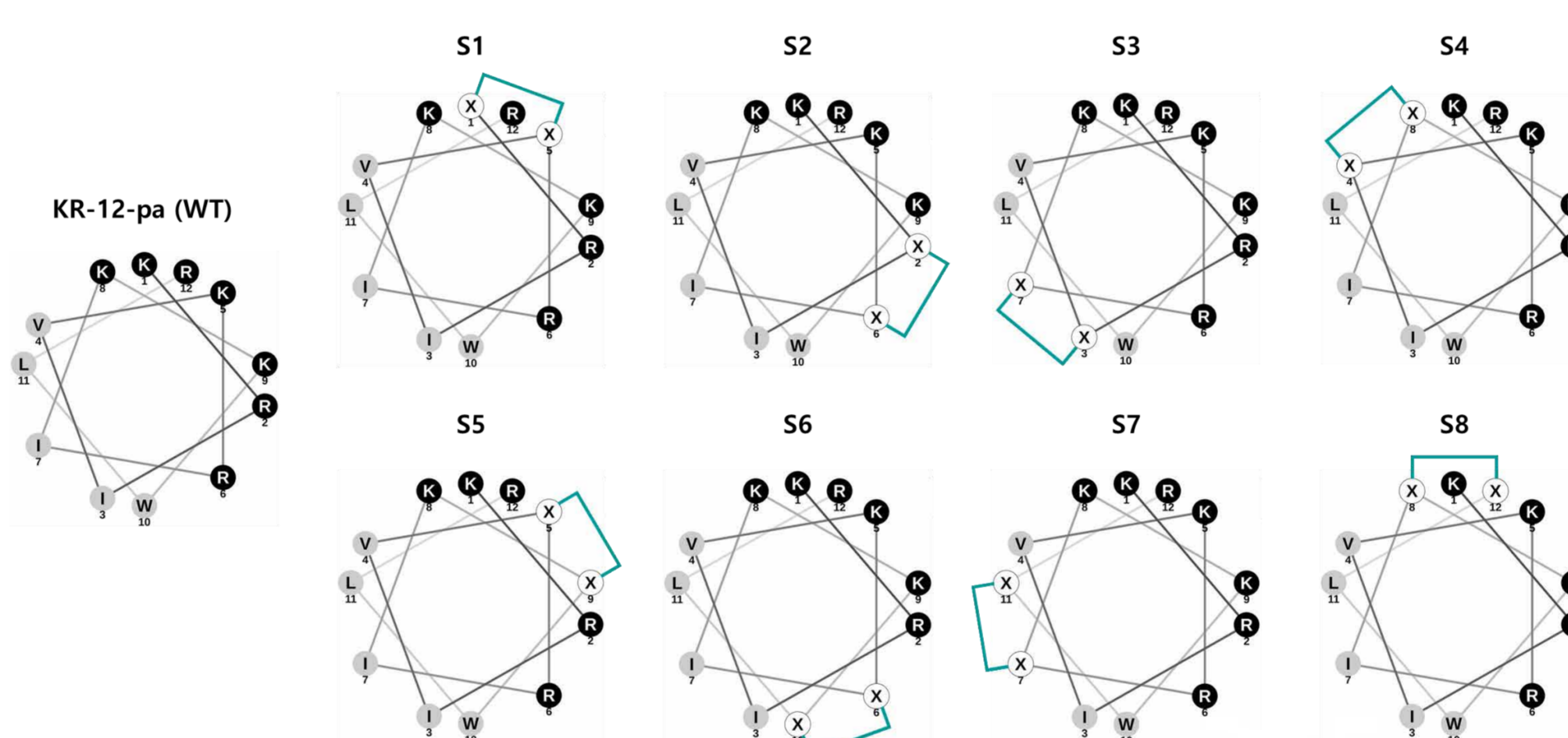
Preparation of KR-12-S3 stapled peptides

Instrumental conditions of peptide with LC-MS

LC-MS system Shimadzu-10AD/AB Sciex API2000
 Column Sunfire C18 (4.5 x 250 mm)
 Buffer A: 100% H₂O containing 0.05% TFA
 B: 100% ACN containing 0.05% TFA
 Method 5-65% for 30 min
 Detector 230 nm
 Flow rate 1 mL/min



Helical wheel diagrams



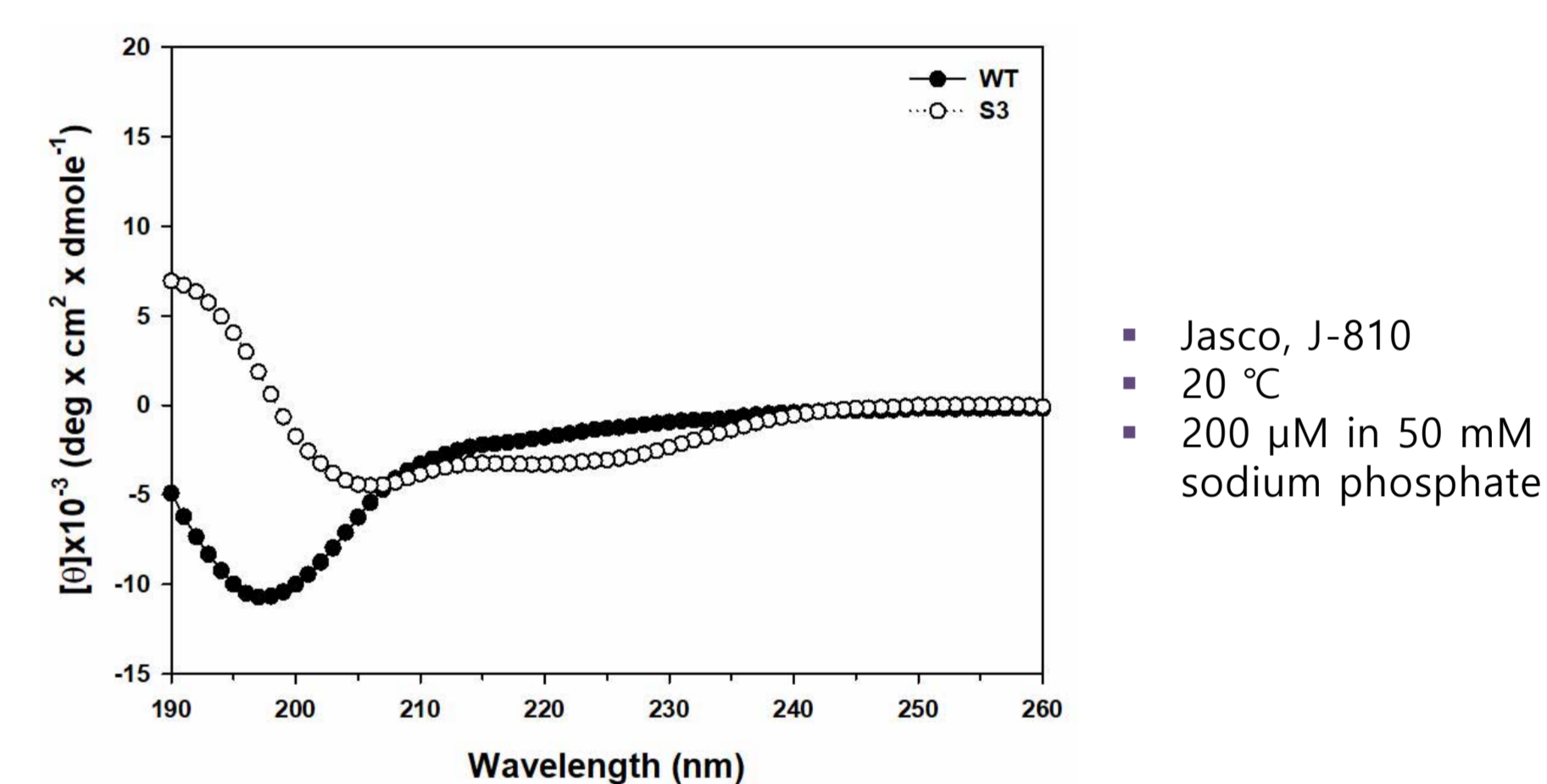
Single stapled (i, i+4), Polar amino acids, Nonpolar amino acids

Antimicrobial activity

Strain	ATCC	MIC ($\mu\text{g/mL}$)								
		WT	S1	S2	S3	S4	S5	S6	S7	S8
<i>C. parapsilosis</i>	ATCC 22019	16	16	4	4	2	16	4	4	2
<i>C. albicans</i>	ATCC MYA-2876	64	16	4	8	4	32	16	16	4
<i>C. glabrata</i>	ATCC 90030	8	4	2	2	2	4	2	2	2
<i>C. glabrata</i>	ATCC 2001	>64	64	4	8	8	>64	8	16	8
<i>C. krusei</i>	ATCC 6258	64	16	4	4	4	16	8	16	4
<i>C. tropicalis</i>	ATCC 750	4	8	2	2	2	4	2	4	2

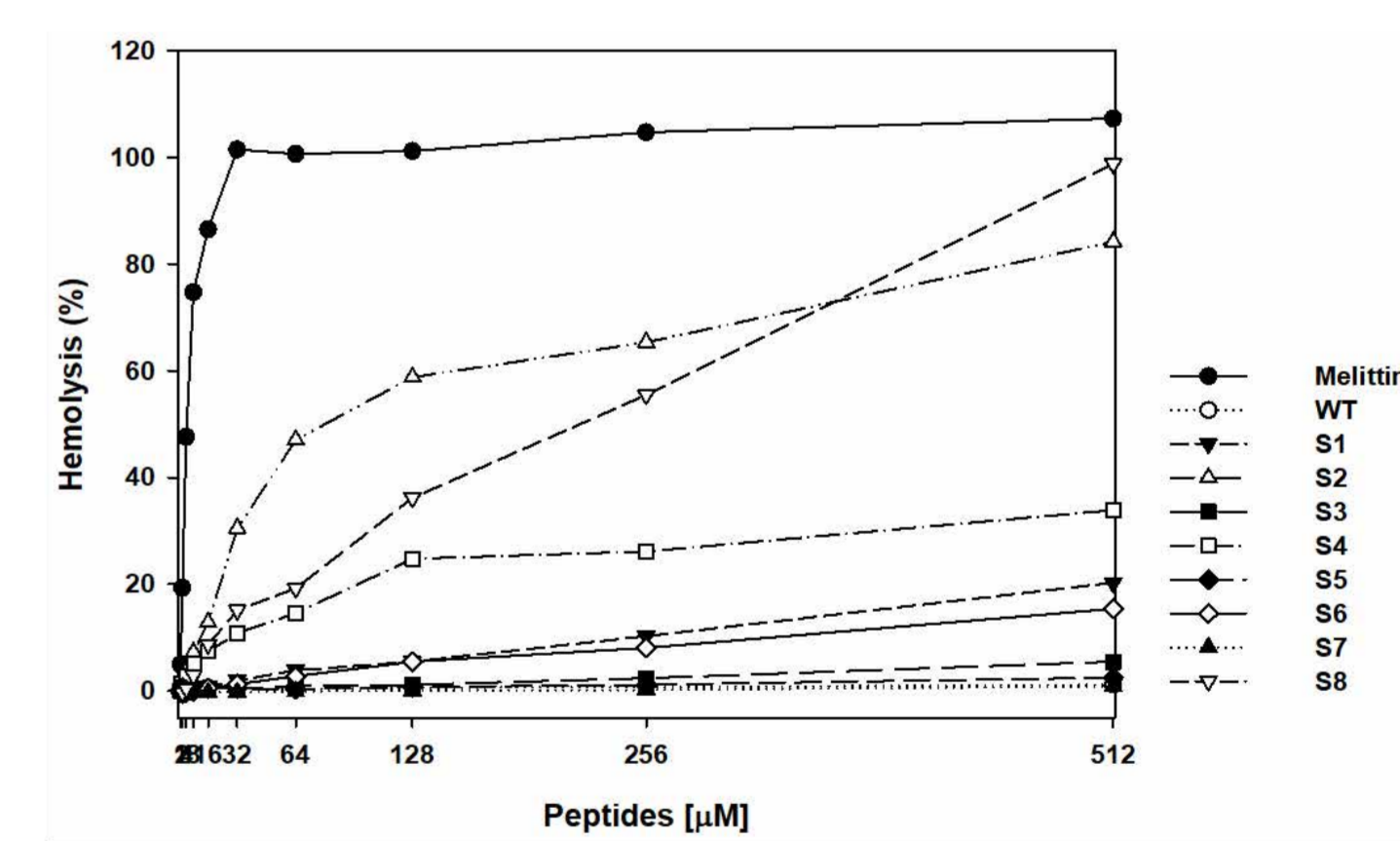
- Antifungal susceptibility strains : ATCC 22019, MYA-2876, 750
- Antifungal susceptible-dose dependent(SDD) strains : ATCC 90030, 2001
- Antifungal resistant strains : ATCC 6258

CD spectra of WT and S3



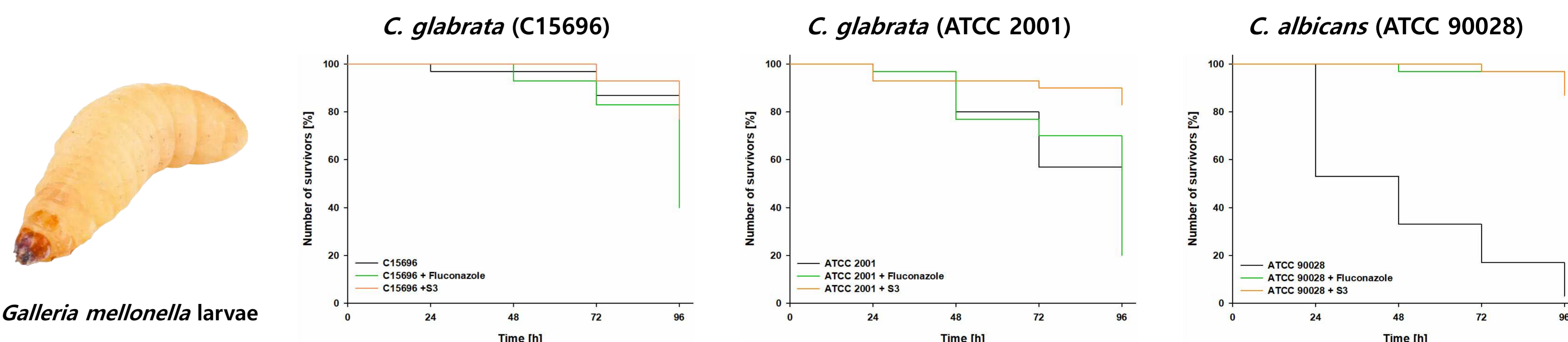
- Jasco, J-810
- 20 °C
- 200 μM in 50 mM sodium phosphate

Hemolytic activity



S5, S7 < WT < S3 < S6 < S1 < S4 < S2 < S8

Effect of S3 stapled peptide from *Galleria mellonella*



Conclusion

Eight stapled peptides based on KR-12-pa, derived from the human antimicrobial peptide LL-37, were synthesized using solid-phase synthesis and purified by RP-HPLC. Among the eight stapled peptides, S3 demonstrated high antifungal activity and low hemolytic activity. The CD spectrum showed that S3 maintains an α -helical conformation in buffer solutions. Preliminary *in vivo* assessments of the S3 peptide, confirmed using *Galleria mellonella* larvae, showed low toxicity and heightened antifungal efficacy.