

TOWARDS PEPTIDE-BASED THERAPEUTICS AGAINST CARDIAC DISEASE Prediction and simulation of the S100A1ct peptide with a membrane environment

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On the left: S100A1ct sequence. In bold the S100A1-derived sequence, with red and yellow colours representing the polar and apolar portions. Below, the peptide is highlighted with the same colour scheme on the parent protein^[5]. On the right: dose-dependent augmentation of SERCA2a activity by different concentrations of S100A1ct in cardiac SR vesicles, compared to control^[5].

Peptide drugs can achieve a high degree of selectivity and specificity, however they come with drawbacks such as difficulties in oral and nasal administration in the absence of modifications or carriers ^[6]. S100A1ct is not immune to such weaknesses, therefore further modifications are needed to improve its potency and pharmacokinetics. In order to do so, a deeper characterization of S100A1ct interactions is needed.

Introduction

Cardiovascular diseases are the leading cause of death worldwide according to WHO estimates^[1]. Heart failure accounted for over 60 million deaths in 2017[2] and can be classified into Heart Failure with reduced (HFrEF), preserved (HFpEF) and mid-range (HFmrEF) Ejection Fraction, with HFrEF accounting for many of these hospitalizations ^[3]. S100A1ct is a peptide derived from S100A1, a member of the Ca²⁺ binding EF-hand protein family, that has been observed to exert an inotropic effect on cardiomyocytes, both *in vivo and in vitro*, influencing with RyR2 and SERCA2a similarly to the parent protein^[4,5].

S100A1ct conformations in a membrane bilayer model

ϵ = POPC ϵ = S100A1ct ϵ = Ions ϵ = Water

cMD – Self Assembly

Confirmation of helical propensity in different force fields

- 9 runs of 2 μs at 300K for S100A1ct from each starting point (tot. = 18)
- 9 runs of 2 μs at 310K for S100A1ct from each starting point (tot. = 18)
- 9 runs of 2 μs at 310K for the flexible linker mutant from each starting point (tot. $= 18$)

Representative of 3 successful assemblies

7 0.95 0.61 0.5 0.46 0.14

94 0.61 0.32 0.23 0.07

73 0.71 0.41 0.13 0.0 0.0

0.83 0.51 0.32 0.23 0.07

 0.61 0.5 0.46 0.1

14 00 00

A mixture of POPC lipids, 0.15mM NaCl, water and S100A1ct was used as starting point for conventional Molecular Dynamics at 300K until a membrane bilayer is formed. 10 Replicates were run with the initial peptide conformation obtained after a short implicit solvent simulation and 9 with the α helical conformation obtained from *Glaser et al.*

In each force field, conventional Molecular Dynamics simulations were run for 600 ns from 8 starting points, each with two replicates. Four starting points with "M1"-like and "M2"-like conformations inserted in the membrane leaflet and transmembrane and other four with an helical conformation inserted in the same positions.

80 different runs were obtained from Monte Carlo (MC) simulations, 20 for each starting point (S100A1ct in the leaflet, transmembrane, in water and in the middle of the implicit membrane). The simulations were run at 310K using SLIM implicit membrane model^[6] and Amber99sb*-ILDN force field using SIMONA software^[8]. The obtained conformations were clustered by energy after high energy outliers (over 3 times the standard deviation) were removed.

The process of insertion in the membrane follows a pathway that relies on the appearance of gaps between the polar heads, as recently observed for another peptide^[9]. Importantly:

S100A1ct peptide has been observed to interact with SERCA2a and predicted to do so with a transmembrane (TM) helical conformation. Here we show that:

- A TM peptide arrangement can be found using an unbiased "assembly" approach and it is stable across the GaMD trajectories
- S100A1ct appears, however, to be found more frequently at the interface between a single membrane leaflet and the aqueous environment
- These findings were confirmed with short MD simulations in two different force fields
- Monte Carlo simulations employing an implicit membrane model also show agreement with these results, providing an efficient means to quickly explore how mutations and different chemical modifications can impact the identified states

- Evaluation of the impact of amino acid mutations on the peptide's preferred conformations in membrane with Monte Carlo simulations
- Simulation of both insertion and passage through the phospholipid bilayer in a more realistic cardiomyocyte model
- Clarify the role of the different kinked states in the peptide pharmacological properties
- Design and evaluation of peptidomimetics derived from such peptide.
- Investigating the possible role of cooperative effects in the switch between stable conformations

Outlook/Ongoing

Conclusions

References

Starting from the representative systems shown, the following MD simulations were performed:

 $=$ Peptide in the leaflet = Transmembrane

e energy here reported comes the force field and can be used ompare different mutants, it does represent the real one

MC-sampled, TOP_100_PIL, secondary structure propensities MC-sampled, TOP_100_TM, secondary structure propensities

0.0 0.0 0.0 0.0 0.0 0.0 0.18 0.64 0.67 0.67 0.27 0.0 0.0 0.0 0

Monte Carlo simulations in implicit membrane

• WT: • FULL: 3/32 • PARTIAL: 12/32

- MUT:
	- FULL: 1/18 • PARTIAL: 3/18

• WT: • FULL: 1/32 • PARTIAL: 0/32 • MUT: • FULL: 0/18

• PARTIAL: 0/18

• WT: • FULL: 0/32 • PARTIAL: 0/32 • MUT: • FULL: 1/18 • PARTIAL: 0/18

• WT: • FULL: 0/32 • PARTIAL: 0/32 • MUT: • FULL: 2/18 • PARTIAL: 0/18

cMD

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- Aromatic residues play a major role in the process, especially when a rigid linker is present at the N-terminus
- When such linker is more flexible, pathways relying on hydrophobic stretches appear, highlighting the importance of carefully designing the 'non-active' parts of the peptide

S100A1ct insertion in a membrane bilayer model

