Planar aggregation of the influenza viral fusion peptide alters membrane structure and promotes poration

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Background

The **fusion peptide** (FP) domain of influenza A hemagglutinin is the only portion of the virus that interacts directly with the target membrane of the host organism; its primary function is to facilitate fusion between the viral membrane and the host.



Side and top views of the helical hairpin structure^[1] of Influenza A fusion peptide domain (sequence: GLFGAIAGFIEGGWTGMIDGWYG). Side chain coloring: red – anionic; green – polar; grey – nonpolar.

Results

Dimers are favored regardless of bilayer constituents

Potentials of mean force (PMF), for antiparallel dimer formation in four membrane compositions. The minima at a peptide COM distance of 12.2 Å corresponds to the antiparallel dimer structure.



peptide COM distance



Antiparallel dimer stabilized, with $\Delta G \approx -5$ kcal/mol in the four membranes simulated, indicating population at equilibrium should primarily be antiparallel dimers



Simulation snapshots, depicting an antiparallel dimer (circled) that forms at $t = 4 \mu s$ and remains stable through the remainder of the 21 µs simulation. Top down view of bilayer; lipids and water omitted. Taken from Ref [2]

Previous experiments and simulations have shown the ability of the isolated FP domain to aggregate when membrane-bound, and form pores in model membrane systems^[2,3]; poration was demonstrated to be inhibited by cholesterol. It was hypothesized that FP clustering promotes poration by decreasing the hydrophobic thickness a pore must span in order to form.



FP-induced dye leakage in GUVs as a function of cholesterol content. The inset depicts a typical dye leakage experiment, where fluorescent dye leaks into the GUV after FP is added. Adapted from ref. [2].

Cholesterol mol %

subsequent simulations cholesterol However, in appeared to enhance FP clustering, seemingly in conflict with the original hypothesis. Here, we aim to resolve this apparent discrepancy by characterizing the energetic landscape of FP–FP interactions and elucidating the role that membrane-specific properties play in FP clustering and poration.

Tetramer formation is largely membrane-dependent



Potentials of mean force (PMF), for tetramer formation from two antiparallel dimers. Both have a minima at a dimer COM distance ~17.7 Å, which corresponds to dimers with no lipid separating them. The PMF in **POPC has a second minimum at** ~24 Å, corresponding to separation by a single layer of lipids, a feature which is absent from the chol:POPC PMF.

> Dimer-dimer interfaces in POPC often include lipids, whereas in chol:POPC direct contact

Spontaneous curvature drives lipid enrichment in the bottom leaflet

Lipid radial distribution functions, demonstrating lipiddependent enrichment underneath an FP dimer. Lipids that generate negative spontaneous curvature (DOG and cholesterol) are enriched while lysolipid which generates positive spontaneous curvature is depleted. The gray shaded regions represent the approximate extent of one FP, along its shorter and longer axes.



Simulation Details

- CHARMM 36 force field
- Unbiased long timescale MD simulations
 - 10 FP in POPC, 1:1 chol:POPC, 7:3 OPC:POPC • 21 µs each
 - 21 FP in POPC and 1:1 chol:POPC
 - 3x10 µs each
 - 10 FP dimers in POPC and 1:1 chol:POPC • 3x10 µs each
- Free energy calculations Umbrella Sampling • POPC, 1:1 chol:POPC, 7:3 OPC:POPC, & 24:76 DOG:POPC
 - Dimerization PMFs
 - 35 windows, 1 µs per window
 - Tetramerization PMFs
 - 26 windows, 1 µs per window
- Simulation packages
 - OpenMM^[4] and Anton 2^[5]
- Analysis

between dimers is more likely



Simulation snapshots, depicting arrangement of ten antiparallel FP dimers in POPC (left) and chol:POPC (right) after 10 µs of equilibration. Top down view of bilayer, lipids and water omitted.

COM distance (radial)

Lipids that generate negative spontaneous curvature are enriched under FP aggregates, while lipids that generate positive spontaneous curvature are depleted, consistent with predictions from theory ^[8]



In pure **POPC** membranes, dimer-dimer interfaces typically include several lipids. These lipids are highly perturbed by the neighboring FP, destabilize the

In **chol:POPC** membranes, cholesterol is excluded from

dimer-dimer interfaces and many interfaces contain no lipids. These direct peptide-peptide contacts prevent water from penetrating into the bilayer, potentially stabilizing the bilayer and inhibiting pore formation.

Hypothesis

• PMFs – WHAM ^{[6]1} • RDFs – LOOS ^[7]

bilayer, and represent a weak point in the bilayer, thus acting as sites of pore nucleation.

References

[1] Lorieau, Louis, Bax, PNAS **2010**, *107*, 11341–11346 [2] Rice et al. Nat. Comm. 2022, 13(1), 7336 [3] Rice, Zimmerberg, Pastor, *Biophys. J.* **2023**, *122*(6), 1018–1032 [4] Eastman et al. PLOS Comp. Biol. 2017, 13, e1005659 [5] Shaw et al. *IEEE* **2014**, 41–53 [6] Grossfield, "WHAM : the weighted histogram analysis method" v. 2.0.9 [7] Romo, Grossfield, Conf. Proc. IEEE Eng. Med. Biol. Soc. 2009 [8] Barnoy, Parton, Kozlov, *bioRxiv*, doi: 10.1101/2024.07.09.602763 [9] Humphrey, Dalke, Schulten, J. Molec. Graphics 1996, 14, 33–38

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Conclusions

- Formation of antiparallel FP dimers is highly favorable, driven primarily by peptide-peptide contacts, and not strongly influenced by membrane composition
- Higher order clustering of FP is membrane dependent:
 - Cholesterol presence favors large aggregates with little lipid separating dimers
 - In POPC, interfaces separated by a single lipid layer are also favorable
- In all systems, POPC is enriched around FP
- Lipid composition under peptide aggregates is influenced by spontaneous curvature generation of lipids:
 - Lipids that generate negative spontaneous curvature are enriched
 - OPC (generates positive spontaneous curvature) is depleted
 - Enrichment of chol or DOG under FP may generate negative curvature, stabilizing tetramer formation
- Hypothesize that a single layer of lipids between adjacent peptides destabilizes the bilayer, facilitating poration – potentially explaining why cholesterol inhibits poration