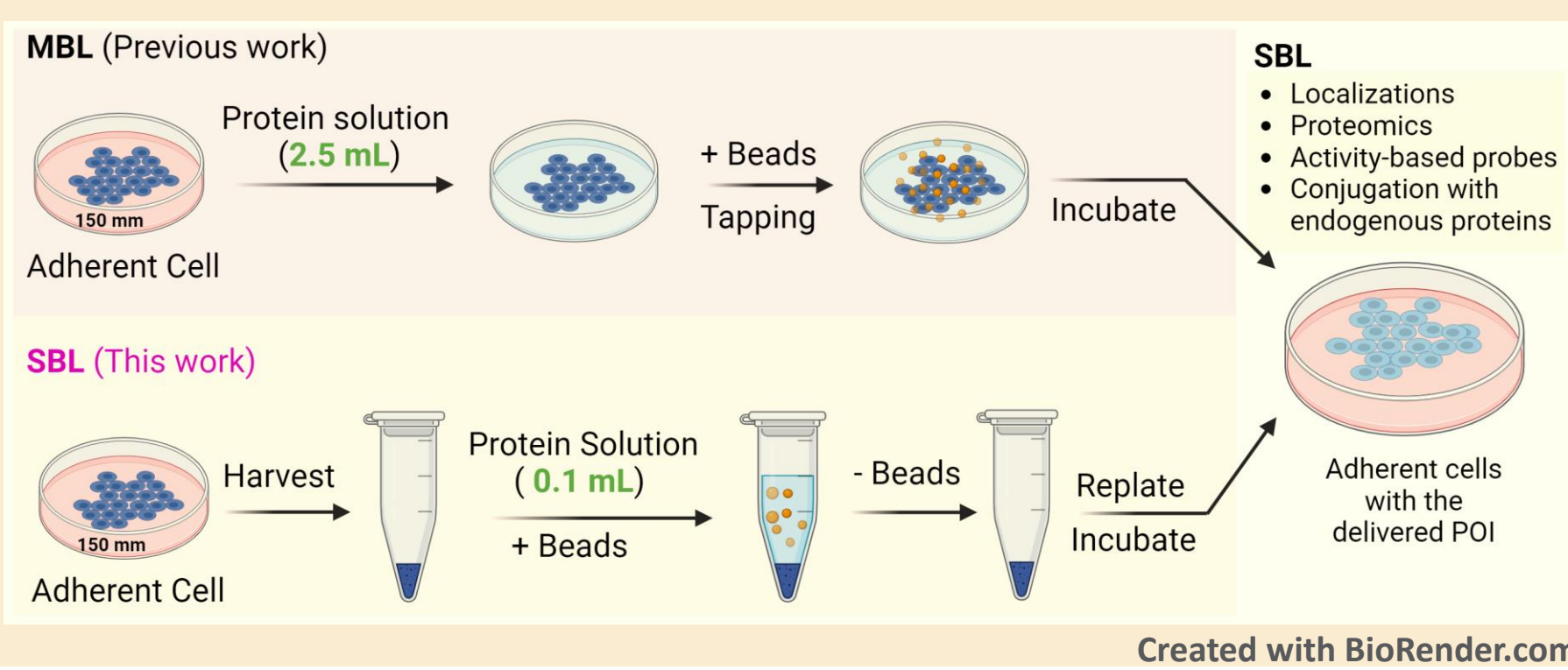


Suspension Bead Loading (SBL): An Economical Protein Delivery Platform to Study URM1's Behavior in Live Cells

Reem Mousa[‡], Abhishek Saha[‡], Yam Alalouf, Pradeep Sadhu, Mahdi Hasan, Shaswati Mandal, Guy Mann, and Ashraf Brik*
[‡] Schulich Faculty of Chemistry, Technion-Israel Institute of Technology, Haifa, 3200008, Israel.
 Reem.mousa@campus.technion.ac.il

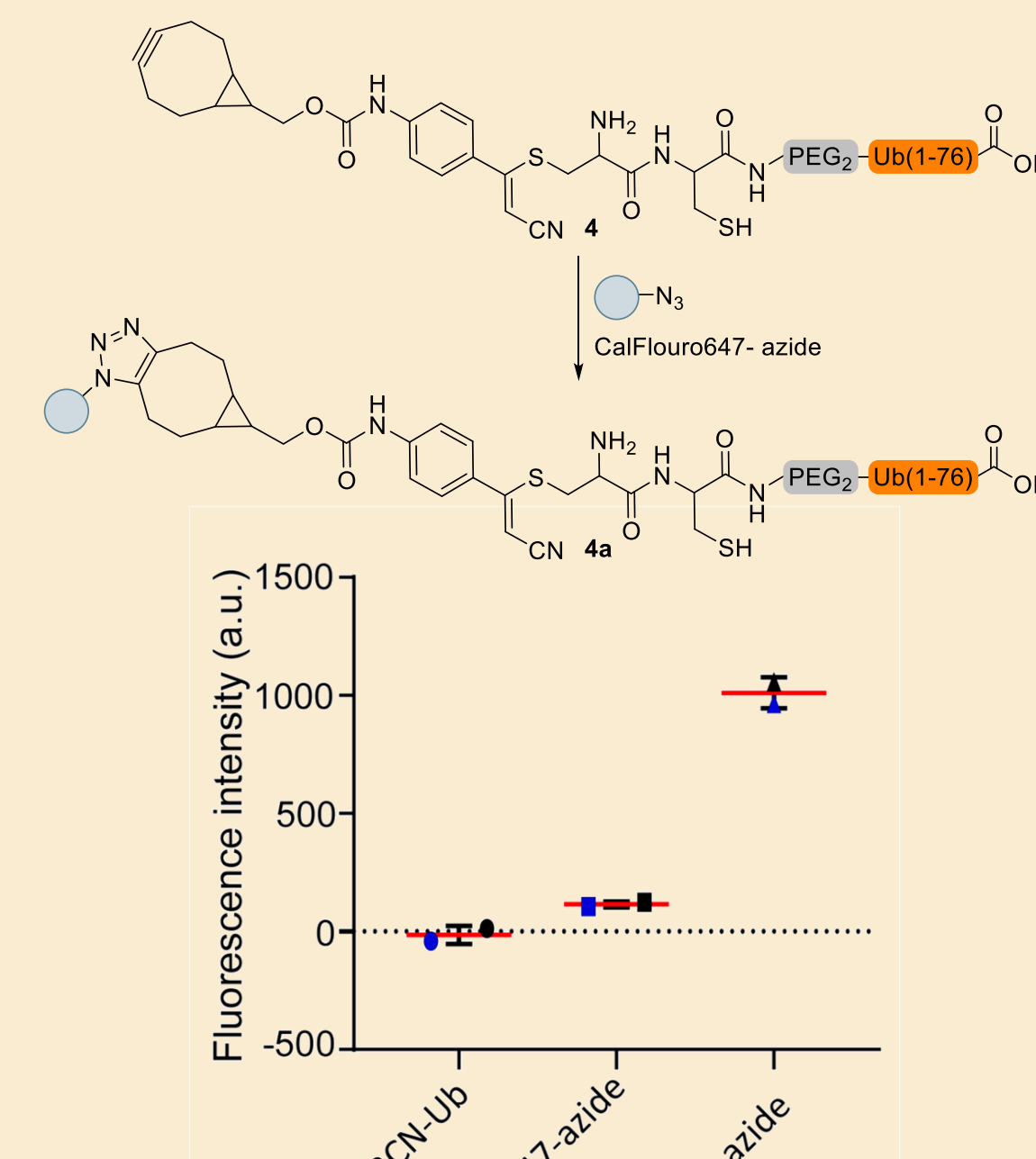
Abstract

Uniquely modified synthetic proteins are difficult to produce in large quantities, which could limit their use in various *in vitro* settings and in cellular studies. In this study, we developed a method named "suspension bead loading" (SBL), to deliver protein molecules into suspended living cells using glass beads, which significantly reduces the amount of protein required for effective delivery. We investigated the delivery efficiency of functionally different proteins and evaluated the cytotoxic effect of our method and the chemical and functional integrity of the delivered protein. We utilized SBL to address questions related to ubiquitin-related modifier 1 (URM1). Employing minimal protein quantities, SBL has enabled us to study its behavior within live cells under different redox conditions, including subcellular localization and conjugation patterns. We demonstrate that oxidative stress alters both the localization and conjugation pattern of URM1 in cells, highlighting its possible role in cellular response to such extreme conditions.



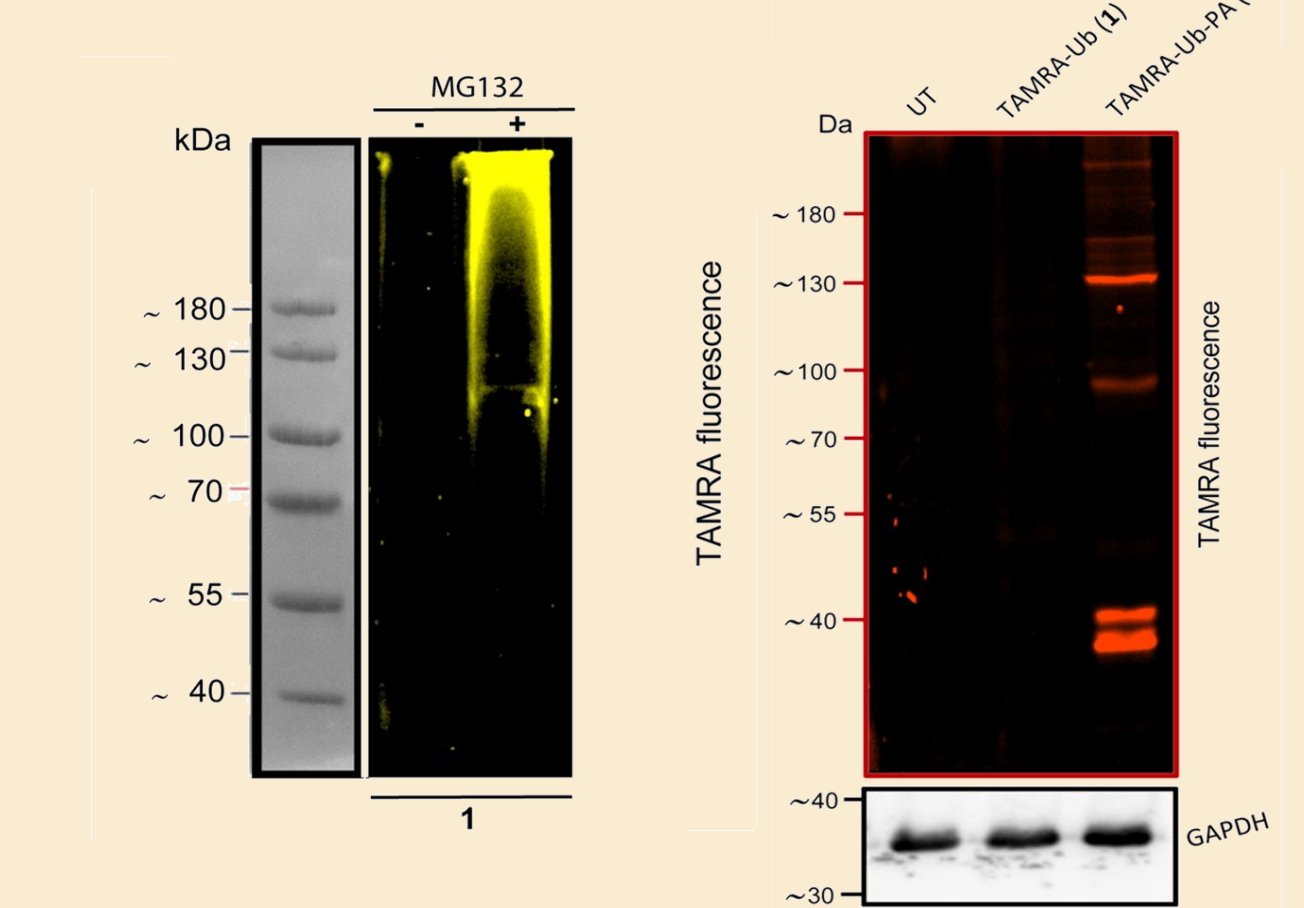
Chemical and functional characterization of Ub cargos

Fluorescent intensity analysis of cargo (4) using SPAAC reaction confirms the integrity of our synthetic Ub in live cells

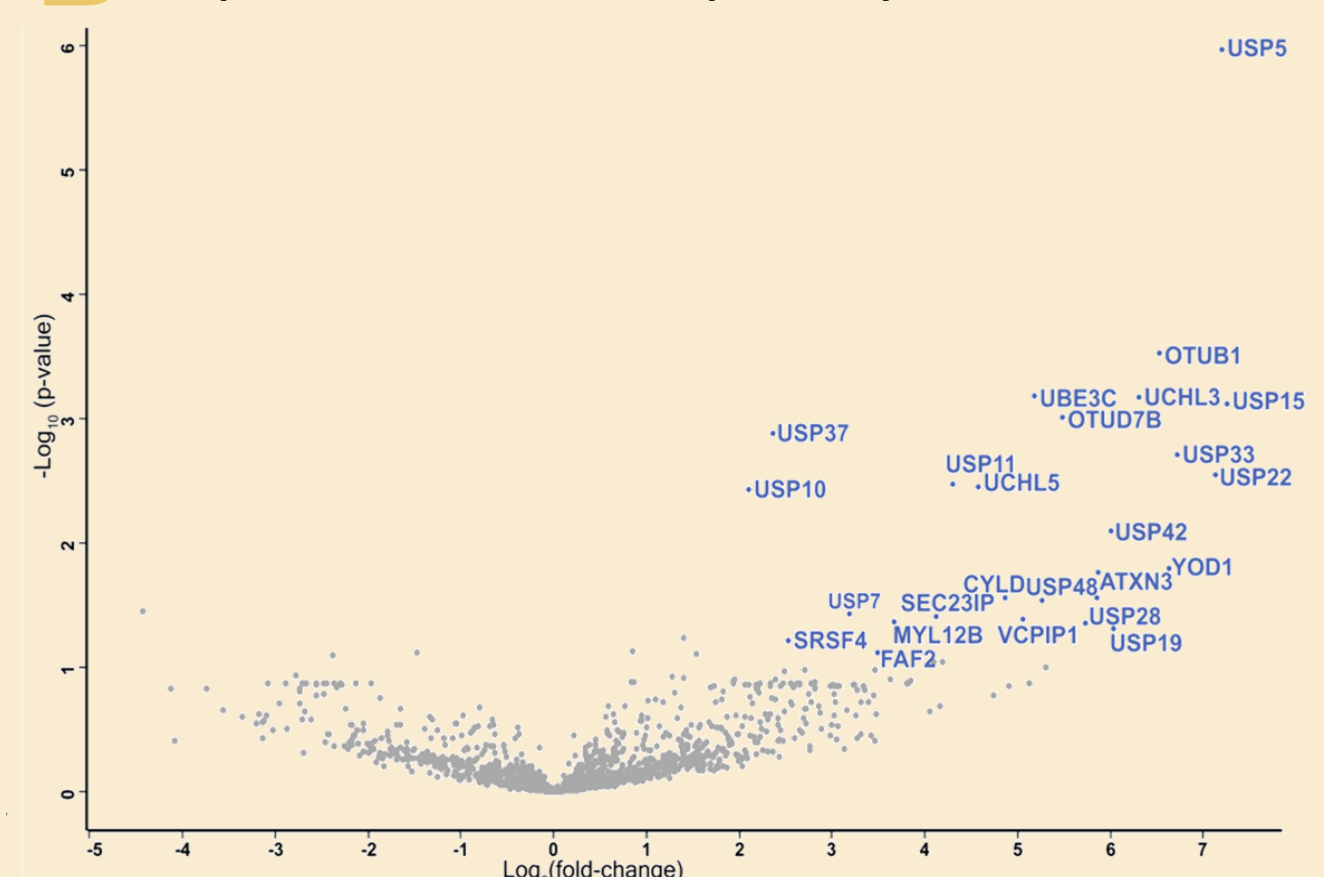


All functional analyses confirm that SBL didn't affect the integrity and functionality of our delivered proteins

Fluorescent gel analysis of the conjugation pattern of synthetic Ub (1), and for Ub-PA (5) capturing DUBs in live cells

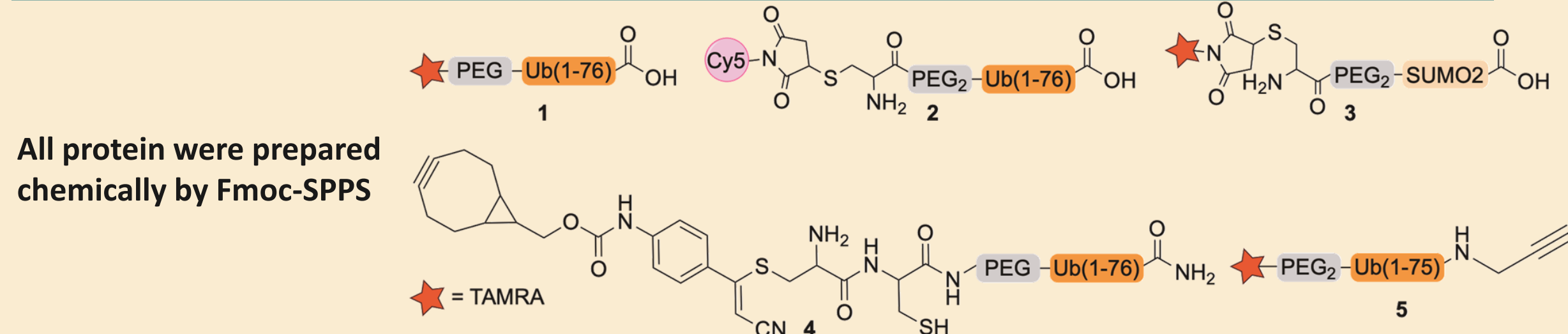


Volcano plot of enriched DUBs captured by Biotin-Ub-PA

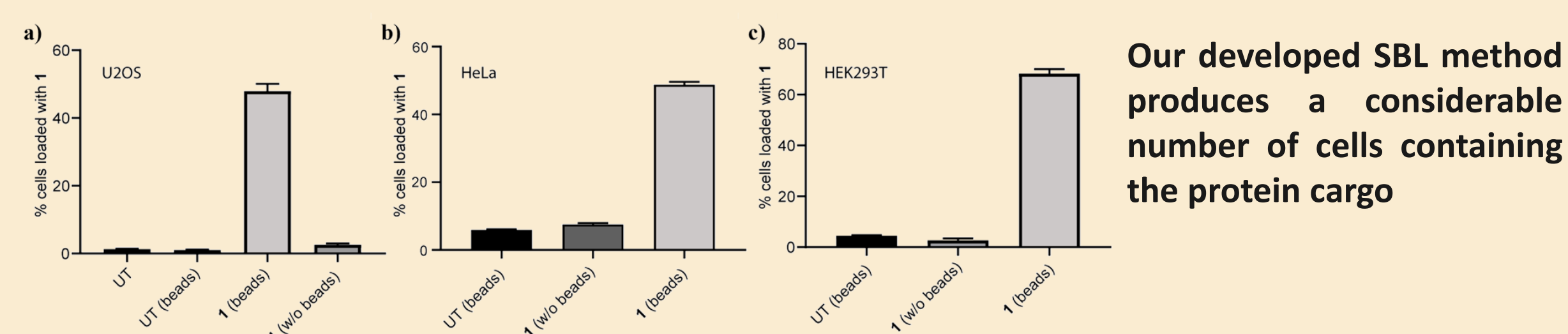


Results

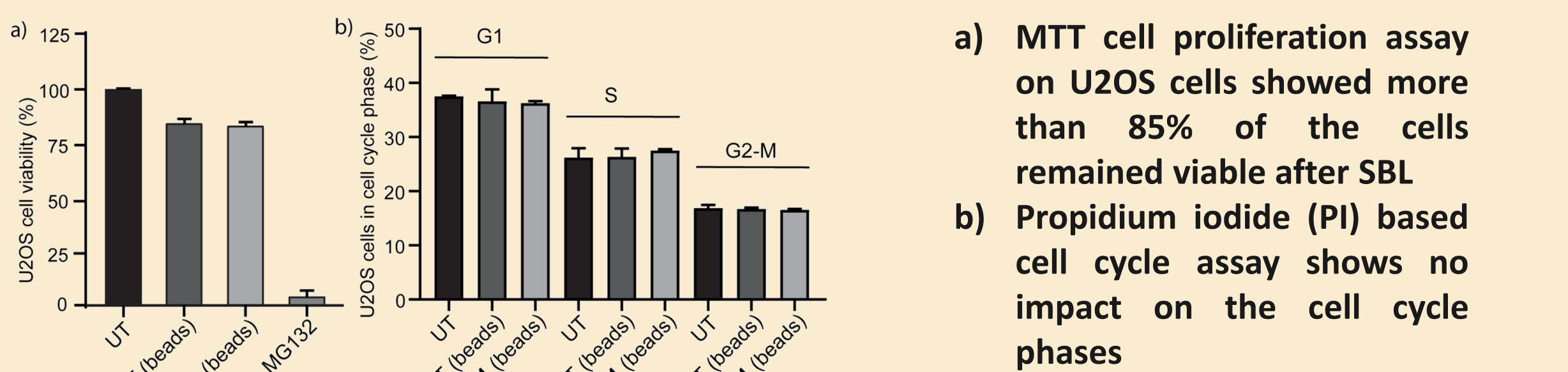
The synthetic proteins employed in SBL development



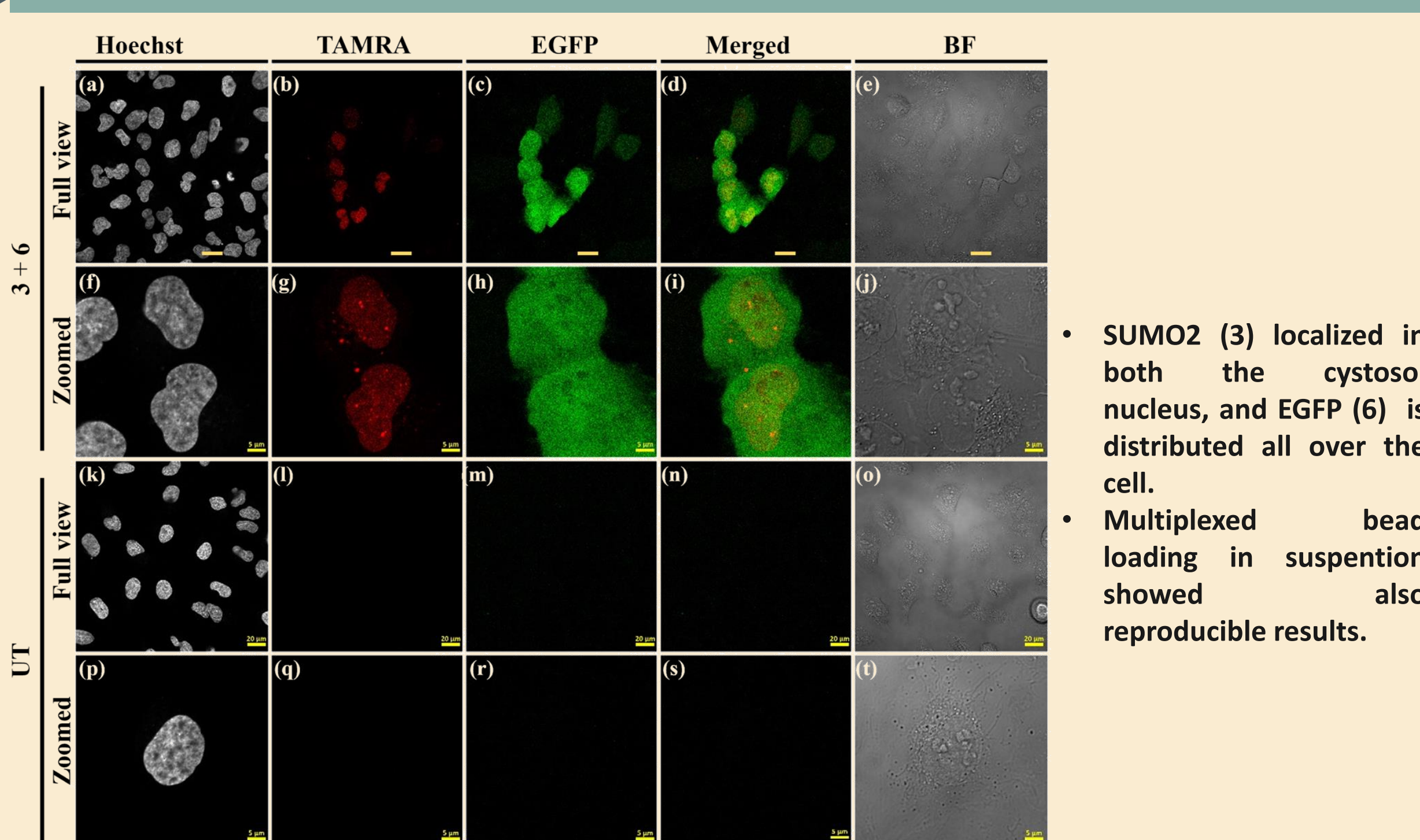
Cellular uptake of TAMRA-Ub (1)



Cytotoxicity studies of SBL

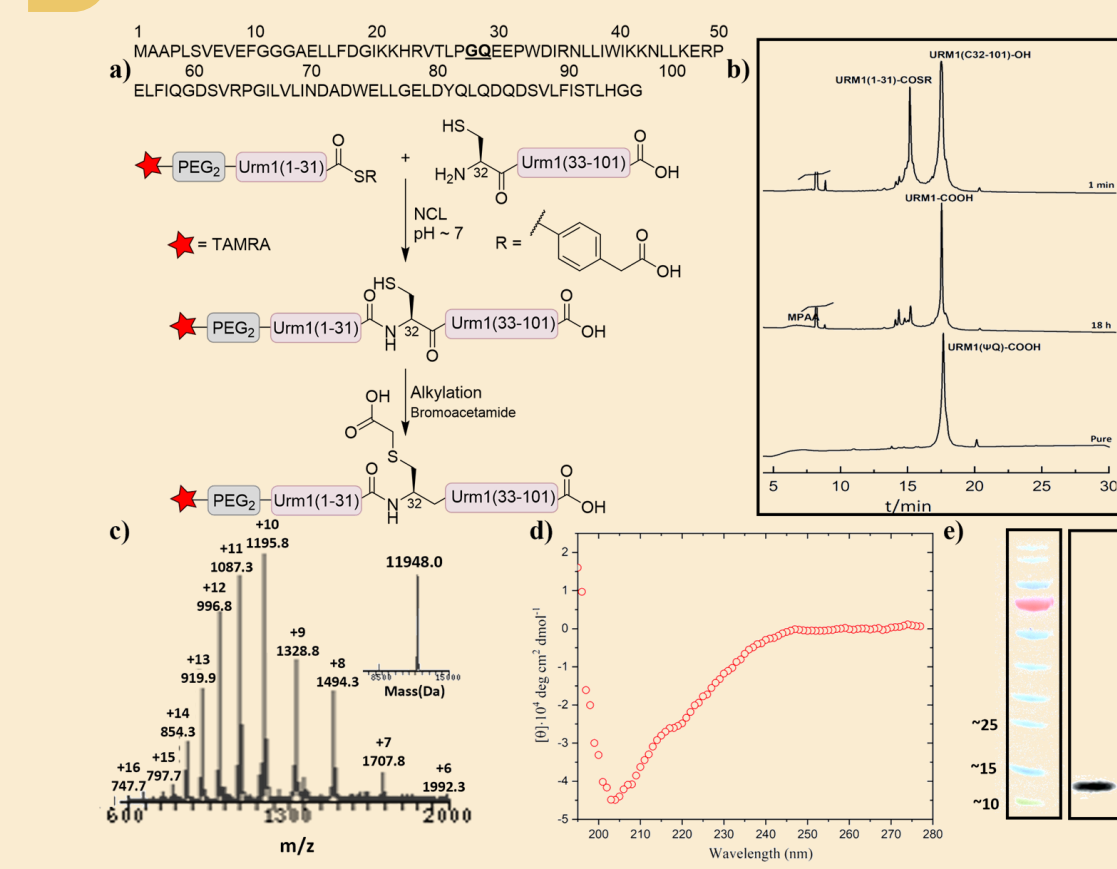


Live cells LSCM images of U2OS cells loaded with SUMO2 and EGFP

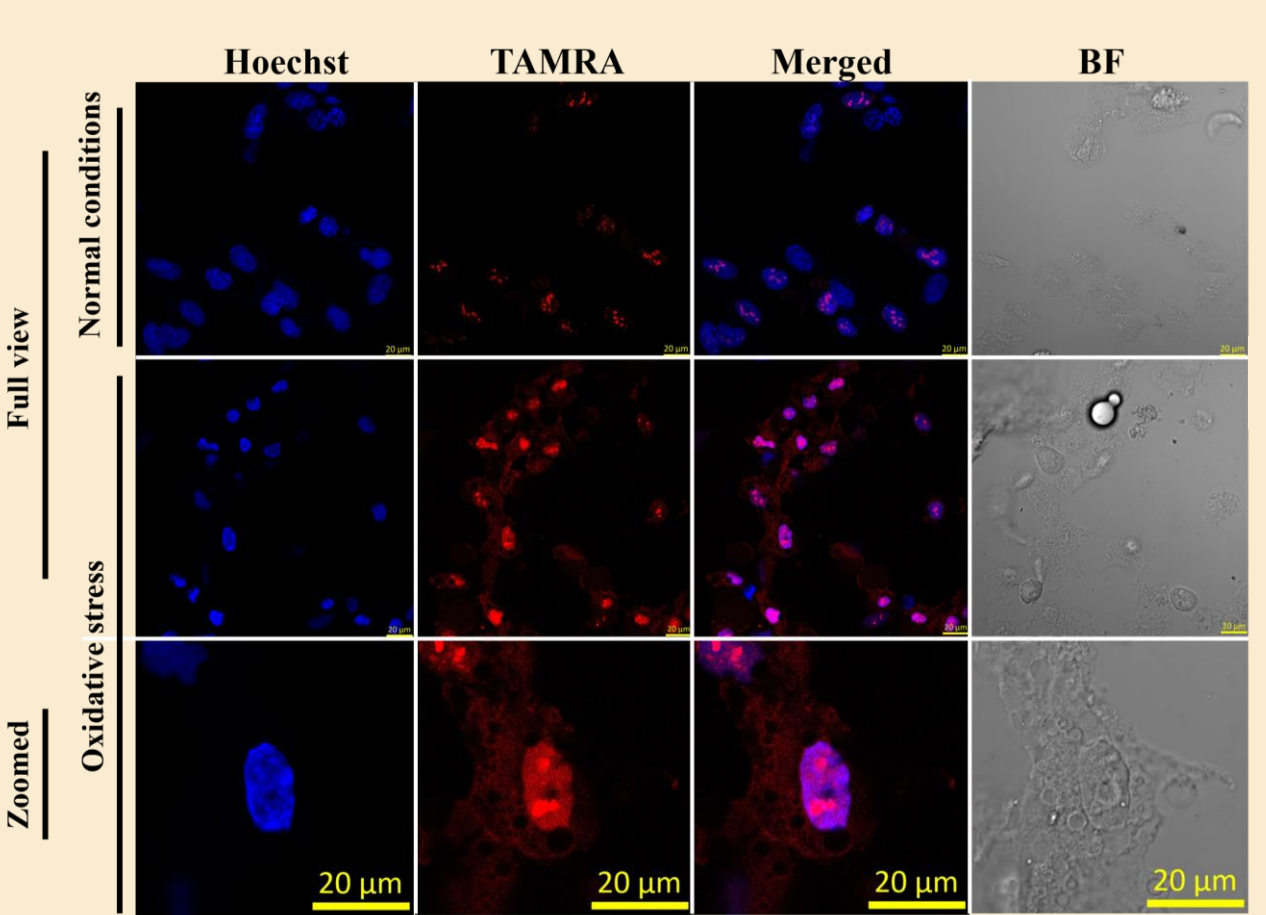


Applying SBL to study ubiquitin like modifier-1 (URM1)

Chemical protein synthesis of URM1

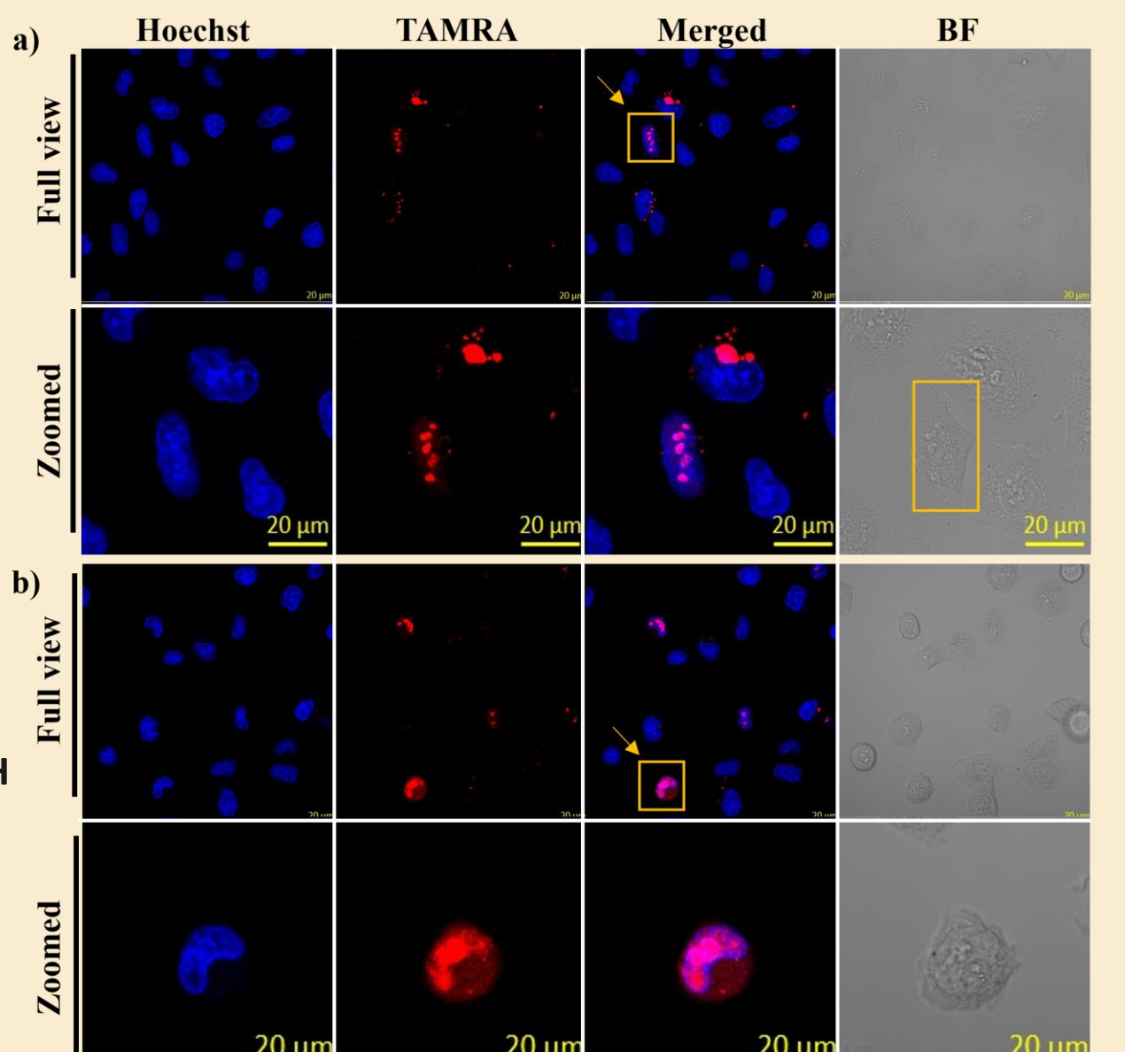


Live cell LSCM images of U2OS cells loaded with URM1-COOH using bead loading

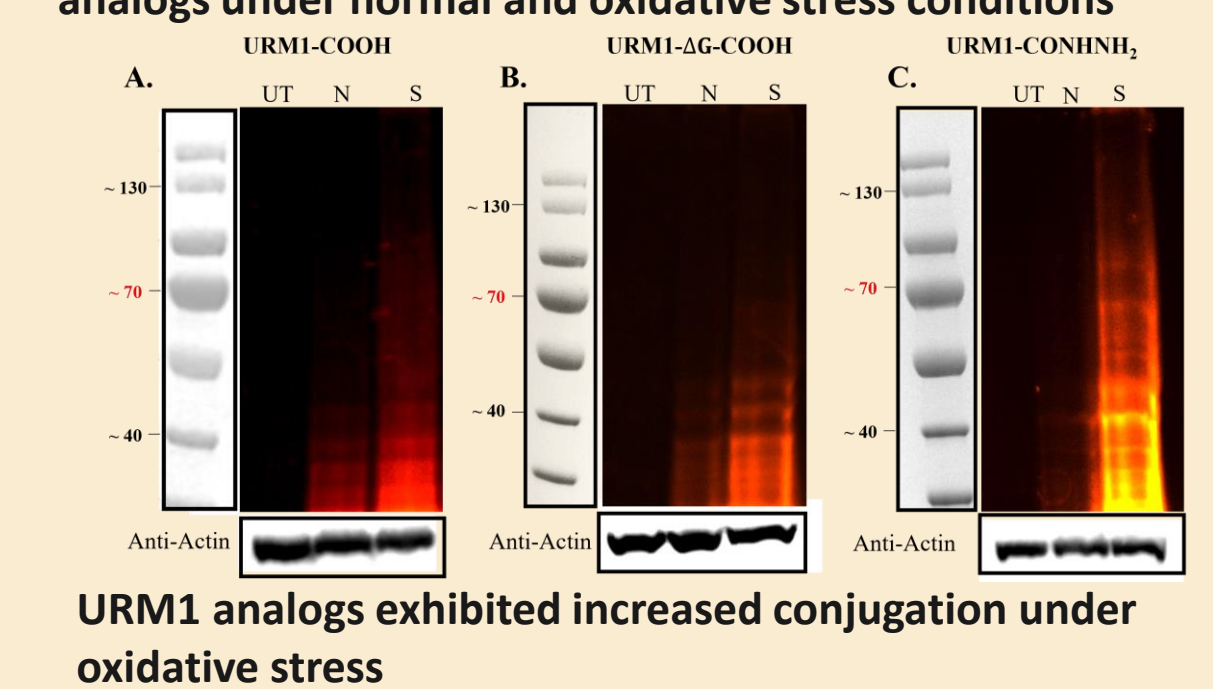


All URM1 analogs localize in the nucleolus under normal condition and diffuse out under oxidative stress.

Live cell LSCM images of U2OS cells loaded with URM1-COOH using the SBL



Fluorescent gel analysis of U2OS cells loaded with URM1 analogs under normal and oxidative stress conditions



Conclusions

- We developed a new general approach for delivering different functional proteins, in suspension phase, applying small amounts of protein(s).
- SBL is also suitable for multiplexed loading of proteins without requiring any special machinery or equipment.
- SBL allowed accessing for the first time URM1, and study some of its cellular properties. Despite challenges in URM1 synthesis, SBL enabled applying minimal quantities (0.1 ml vs. 1ml of protein solution).
- URM1 localizes in the nucleoli under normal conditions and diffuses out under oxidative stress, emphasizing its role and involvement in stress damage response.
- Regardless of its C-terminal functionality, URM1 forms fewer conjugates under normal conditions compared to oxidative stress.

1. Abhishek Saha[‡], Reem Mousa[‡], et al. Suspension Bead Loading (SBL): An Economical Protein Delivery Platform to Study URM1's Behavior in Live Cells, *Angew. Chem. Int. Ed.* Under revision .
 2. Mann, G.; Sadhu, P.; Brik, A. Multiplexed Delivery of Synthetic (Un) Conjugatable Ubiquitin and SUMO2 Enables Simultaneous Monitoring of Their Localization and Function in Live Cells. *ChemBioChem* 2022, 23, e202200122.
 3. Mann, G.; Sadhu, P.; Brik, A. Synthetic Proteins behind the Plasma Barrier: Molecular Spies. *Acc. Chem. Res.* 2022, 55, 2055–2067.