

Enzymatic Peptide and Protein Bromination: The BromoTrp Tag

Nicolai Montau¹ and Norbert Sewald¹

Organic and Bioorganic Chemistry, Faculty of Chemistry, Bielefeld University, Germany

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Introduction: Flavin-Dependent Halogenases

- Flavin-dependent tryptophan halogenases (FDHs) catalyze the chlorination, bromination or iodination of the Trp-sidechain indole at C5, C6 or C7.^[1,2]
 - Environmentally-friendly conditions: aqueous buffer, neutral pH, halide-salt, ambient temp.
 - Remarkable regioselectivity, no oversubstitution
- Poor catalytic parameters, narrow substrate scope, stability and process scalability^[3-11]
- FDHs require FADH₂ in stoichiometric amounts
 - Cofactor regeneration strategy required!
- Halotryptophan is part of many bioactive natural products & peptides.^[4,5]
- Br-Trp: useful chemical handle for transition metal catalyzed cross-coupling reactions

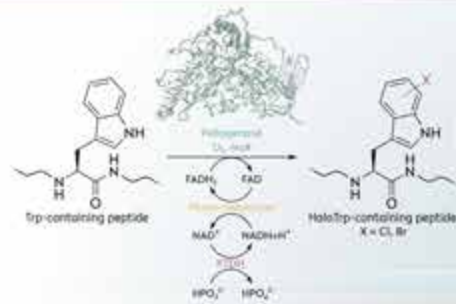


Fig. 1. Mechanism of FDH catalyzed halogenation. FADH₂ reacts with molecular oxygen, forming a C5a-hydroperoxyflavin intermediate. This is attacked by a halide ion (X⁻ = Cl⁻, Br⁻, I⁻) to generate hypohalous acid as an activated halogen transfer reagent. After deprotonation through an aspartate, the halogenating species is activated by a lysine and glutamine residues to form an active halogenating species. Aromatic halogenation proceeds via electrophilic aromatic substitution.

Summary & Outlook

- Enzyme engineering campaign over three rounds of mutagenesis improved conversion of peptides with C-terminal Trp by up to 25-fold.
 - Regioselectivity for indole-C6 fully retained
 - conversion of N-protected Trp-derivatives also greatly improved
 - triple mutant Thal-AAC → strong bromide preference vs. chloride
 - YNIW as a viable protein tag → "BromoTrp Tag"
- Post-translational protein bromination
 - Successful for three model proteins (66-93% conversion)
- Suzuki coupling of brominated DARPin-E01 with phenyl boronic acid
- Potential applications:
 - Novel bioconjugation technique (e.g. for ADCs) → SMC yields stable C-C-Bond
 - Fluorogenic protein labelling^[10]
 - Protein macrocyclization^[12] Palladium-catalyzed protein-protein cross-coupling^[13]
 - Radiobromine labelling^[14] → positron emitters (⁷⁶Br, ⁷⁸Br), Auger emitters (⁷⁷Br, ^{80m}Br)

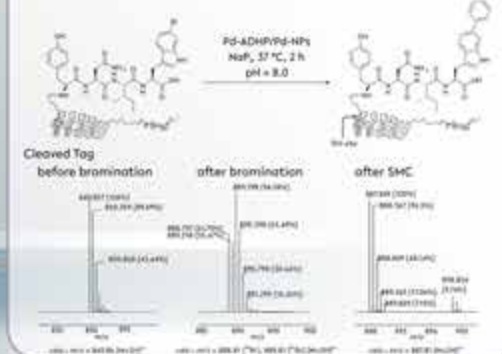
Late-Stage Enzymatic Halogenation



- We previously described effective peptide halogenation of N-terminal Trp-residues.^[11]
- Halogenation of internal or C-terminal Trp → poor conversion

Bioconjugation by Suzuki-coupling

- Pd-catalyzed Suzuki-Miyaura coupling of brominated DARPin-E01 → successful with both Pd-ADHP and Pd-nanoparticles.^[8,9]



Protein Engineering of Thal:
 Peptide Bromination?
 Protected Trp?
 Tagged Proteins?

Enzyme Engineering Campaign

- Enzyme engineering extended the substrate scope of Thal to peptides with Trp at the C-terminus and N^ε-protected Trp derivatives; Conversions improved up to 25x using a triple mutant variant of Thal.
- Thal-AAC exhibits a pronounced halide preference for bromide over chloride.

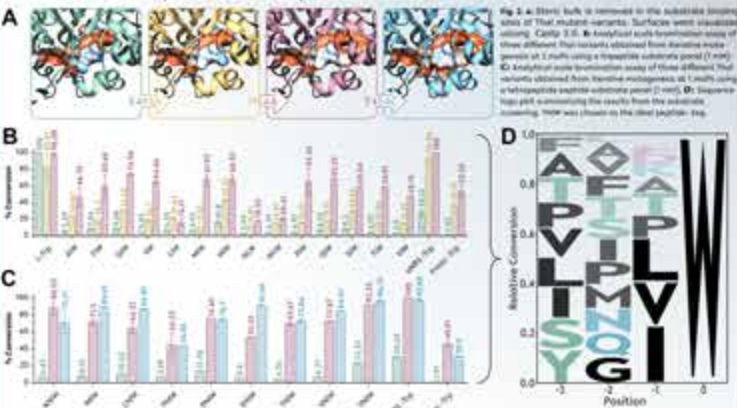


Fig. 2. A) Thal's bulk is removed to the substrate binding sites of Thal mutant variants. Surfaces were visualized using Chimera 3.6. B) Analytical scale bromination assays of three different Trp variants obtained from derivative mutagenesis at 1 mM using a tripeptide substrate panel (1 mM). C) Analytical scale bromination assays of three different Thal variants obtained from derivative mutagenesis at 1 mM using a tetrapeptide substrate panel (1 mM). D) Sequence logo plot summarizing the results from the analytical scale bromination assays (see also in the main paper). Trp

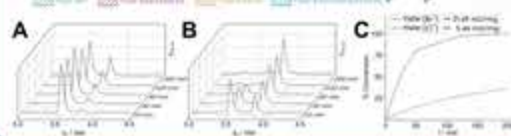


Fig. 3. Thal-AAC exhibits a pronounced halide preference for bromide over chloride. A) HPLC chromatograms monitoring bromination of the peptide YNIW by Thal-AAC. B) HPLC chromatograms monitoring bromination of the peptide YNIW by Thal-AAC. C) Conversion plotted against time. Specific activity is markedly higher for bromination compared to chlorination.

The BromoTrp Tag: Selective Protein Bromination

- The BromoTrp tag YNIW was appended to model proteins via a flexible linker peptide → Coexpression with halogenase cascade in one *E. coli* cell → selective monobromination

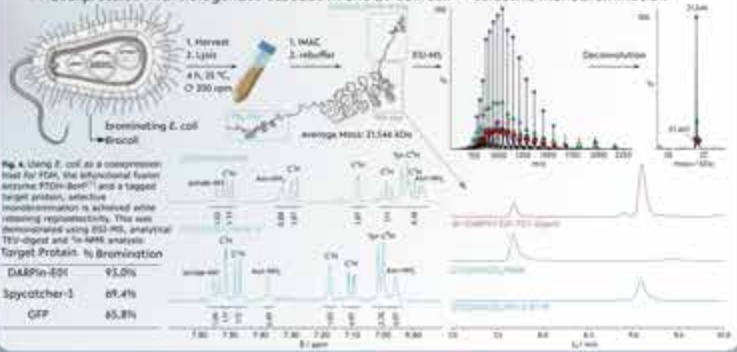


Fig. 4. Using *E. coli* as a coexpression host for FADH₂ the bifunctional fusion enzyme YNIW-Brc¹ and a tagged target protein, selective monobromination is achieved while retaining regiospecificity. This was demonstrated using HPLC analysis. Target Protein % Bromination: DARPin-E01 93.0%, SpyCatcher-1 65.4%, OFF 65.8%.

References & Acknowledgements

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