

Synthesis and antimicrobial activity of hybrid peptides as dual inhibitor of intracellular targets in ESKAPE pathogens



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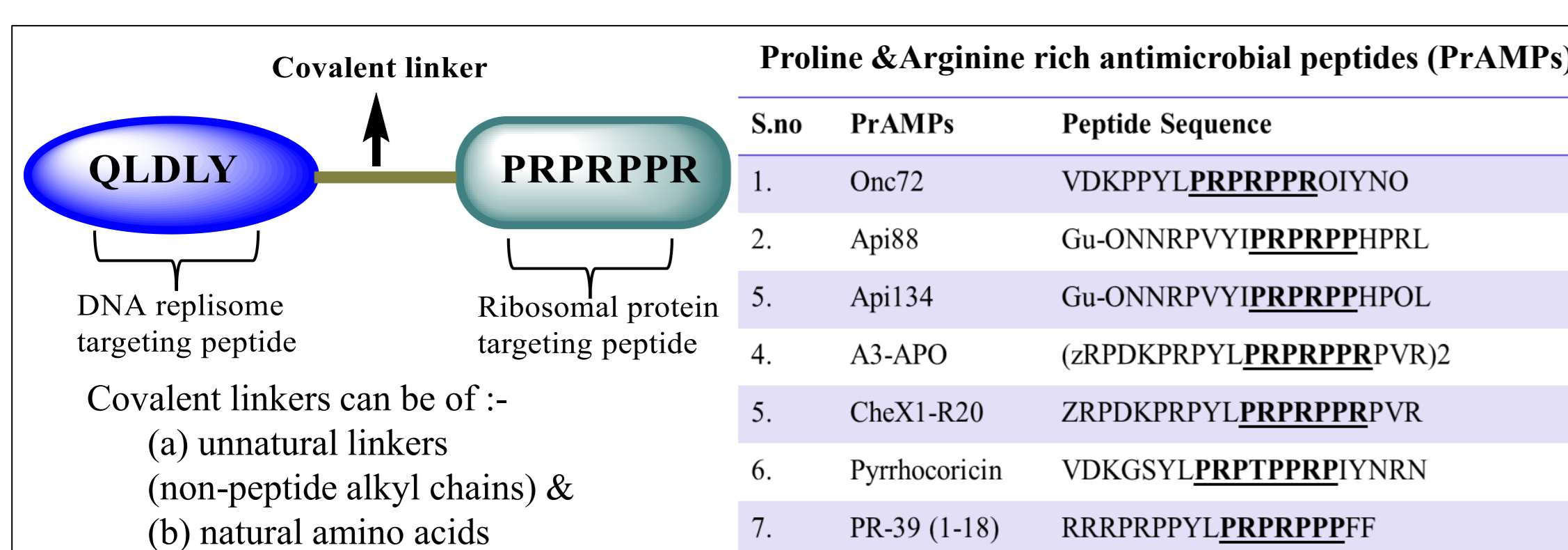
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Background

- The infections caused by ESKAPE pathogens are a global challenge in clinical settings due to their multidrug resistance and even last-resort antibiotics ineffective for treatment.
- Advances in science have shown that bacteria adapt and mutate to resist antibiotics. To combat this, new antibiotics with novel targets and mechanisms are needed.
- Antimicrobial peptides (AMPs), which have evolved over millennia as natural antimicrobial agents and immune response mediators, are one of the promising candidates in this fight.

Rationale & Objectives



S.no	PrAMPs	Peptide Sequence
1.	Onc72	VDKPPYLPRRPPROIYNO
2.	Api88	Gu-ONNRPVYIPRRPPHPRL
5.	Api134	Gu-ONNRPVYIPRRPPHPOL
4.	A3-APO	(zRPDKPRPYLPRRPPRPVR) ₂
5.	CheX1-R20	ZRPDKPRPYLPRRPPRPVR
6.	Pythocorcin	VDKGSYLPRRPPPIYNRN
7.	PR-39 (1-18)	RRRPPPYLPRRPPPF

Objectives:

- Synthesis of the designed hybrid analogues of proline rich antimicrobial peptides.
- Purification & Characterization of the peptide through HPLC and LC-MS
- Determination of anti-bacterial potency of the peptides against ESKAPE pathogens.
- Evaluation of *in vitro* cell cytotoxicity and hemolytic activity of the peptides.

Methodology

Design and synthesis of peptides

- Natural host defense cationic antimicrobial peptides (AMPs) are effective against multidrug-resistant (MDR) pathogens, but their membrane-disruptive action can be toxic to mammalian cells, limiting systemic use.
- However, intracellular-targeting AMPs, which can inhibit DNA or protein synthesis, bypass these restrictions. This study proposes the design of such AMPs to target enzymes involved in DNA replication and protein synthesis.
- The peptides were synthesized using standard solid-phase Fmoc chemistry and characterized by HPLC and ESI-MS.

Antimicrobial potency

- Modified serial broth dilution method was used.
- The peptides were tested against a range of gram-negative and gram-positive pathogens, revealing lead sequences with potential against drug-resistant bacteria.

Cell cytotoxicity assay

- Hemolytic activity of the designed peptides were determined on fresh human red blood cells (hRBC) and were found non-hemolytic even at the concentration of 0.5 mg/mL.
- We conducted cytotoxicity study of the lead peptides by MTT assay method against HEK-293T cell lines at higher concentrations of the calculated MIC value obtained from antibacterial assay (as depicted as in Figure 3.2).

Drug resistance studies

- Resistance development studies of the peptides till 17 passages of bacteria. a) for *E. coli* ATCC 11775 and b) for *S. aureus* ATCC 33591.

Mode of action studies

- Peptide internalization studies (using confocal microscopy).
- Bacterial membrane depolarization assay.

Results

1. Characterisation: HPLC, UPLC & ESI-MS

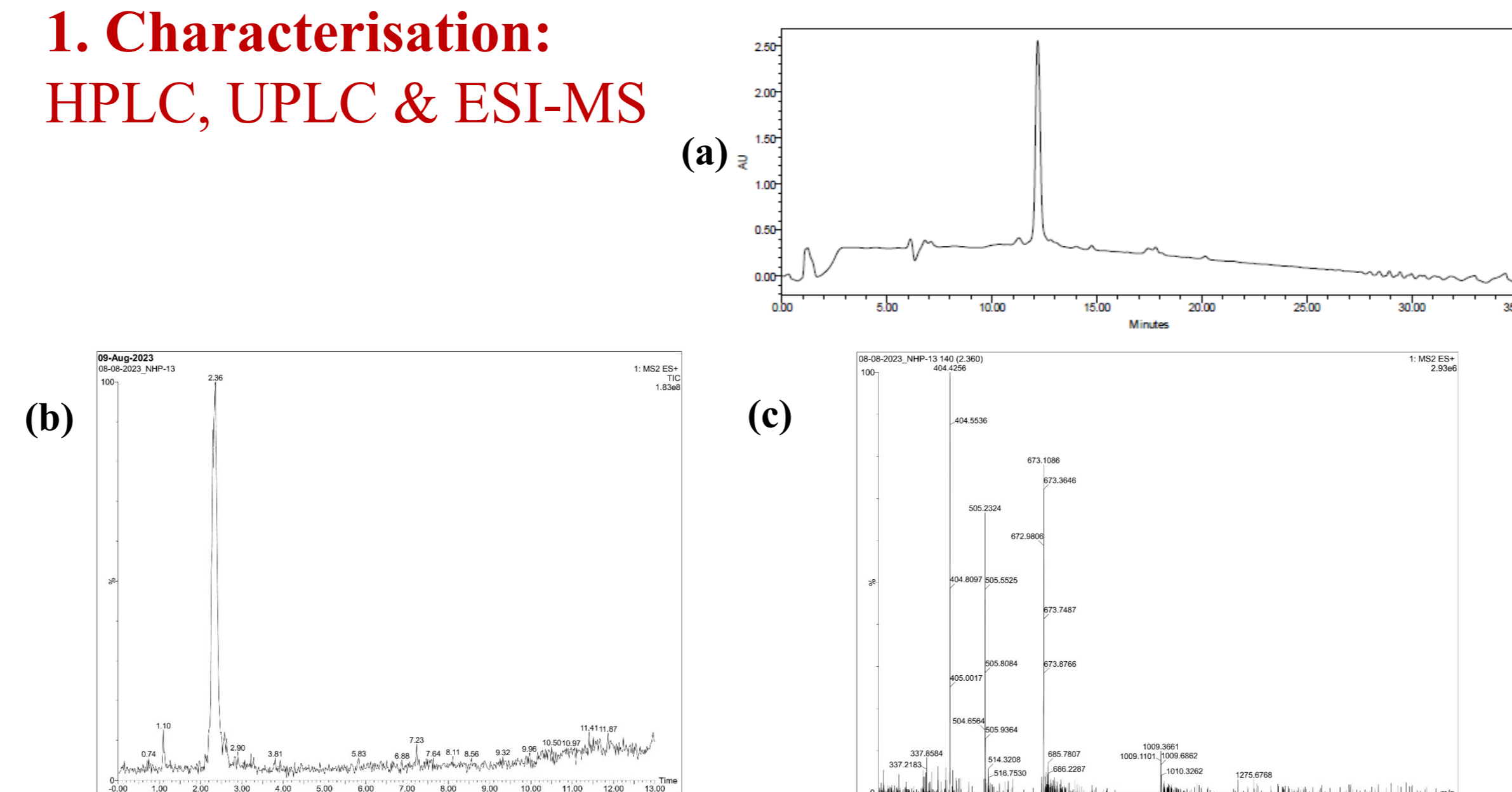


Fig 1: (a) HPLC, (b) UPLC chromatogram, and (c) ESI-Mass spectra of peptides.

2. Antibacterial Assay

Table 1: Peptide name, sequence, and MIC of the synthesized peptides.

S.No	Sequence of peptides	Sample Code	S. aureus ATCC 33591	S. aureus ATCC BAA-44	M. luteus ATCC 56625	B. subtilis ATCC 6693	S. epidermidis ATCC 51625	E. coli ATCC 11775	E. coli ATCC BAA-47	E. faecium ATCC 35667	A. baumannii ATCC 19606	K. pneumoniae ATCC BAA-1705	E. faecalis ATCC 7080
1	Ac-PRRPPR- CONH ₂	Prn	>500	>500	>500	>500	>500	>500	>500	>500	>500	>500	>500
2	Ac-QLDLY-R-PRRPPR- CONH ₂	NHP-11	500	500	500	500	500	>500	500	500	500	500	500
3	Ac-QLDLY-RR-PRRPPR- CONH ₂	NHP-12	250	500	500	>500	500	250	500	250	250	500	250
4	Ac-QLDLY-RRR-PRRPPR- CONH ₂	NHP-13	31.2	31.2	62.5	62.5	31.2	15.1	62.5	31.2	31.2	125	125
5	Ac-QLDLY-Spermine	Spn-1	62.5	31.2	62.5	62.5	31.2	250	250	250	250	250	250
6	Lauric acid-QLDLY-Spermine	Spn-4	7.8	7.8	15.1	7.8	7.8	31.2	62.5	31.2	31.2	125	62.5
7	Vancomycin	Van	0.4	1.9	1	ND	1	62.5	125	62.5	125	ND	125
8	Ciprofloxacin	Cipro	0.2	62.5	0.03	ND	0.03	0.02	0.8	0.5	0.8	ND	0.8

3. Cell cytotoxicity and Hemolytic assay

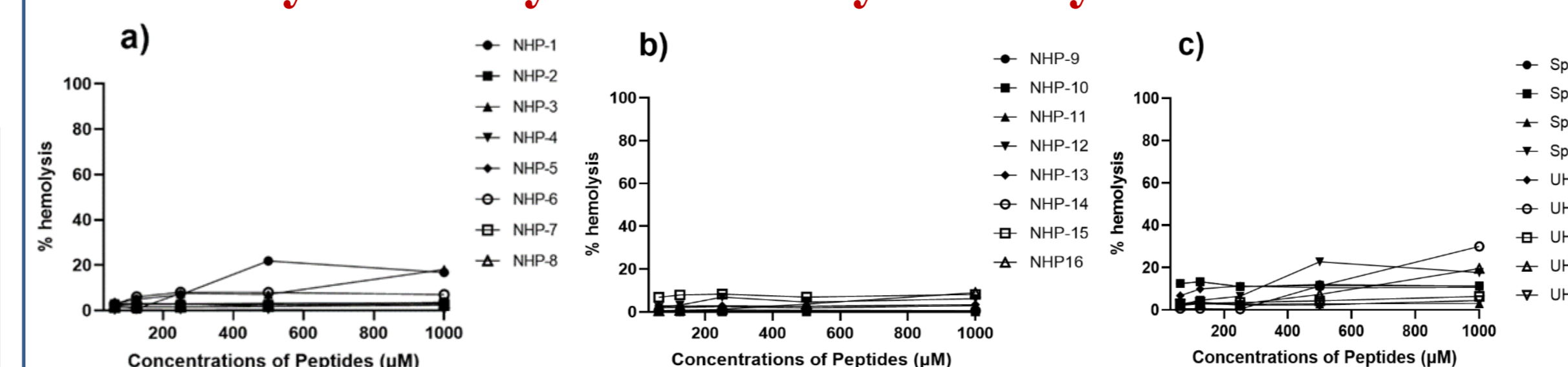


Fig 3.1: Cytotoxicity assay of the peptides against human RBCs.

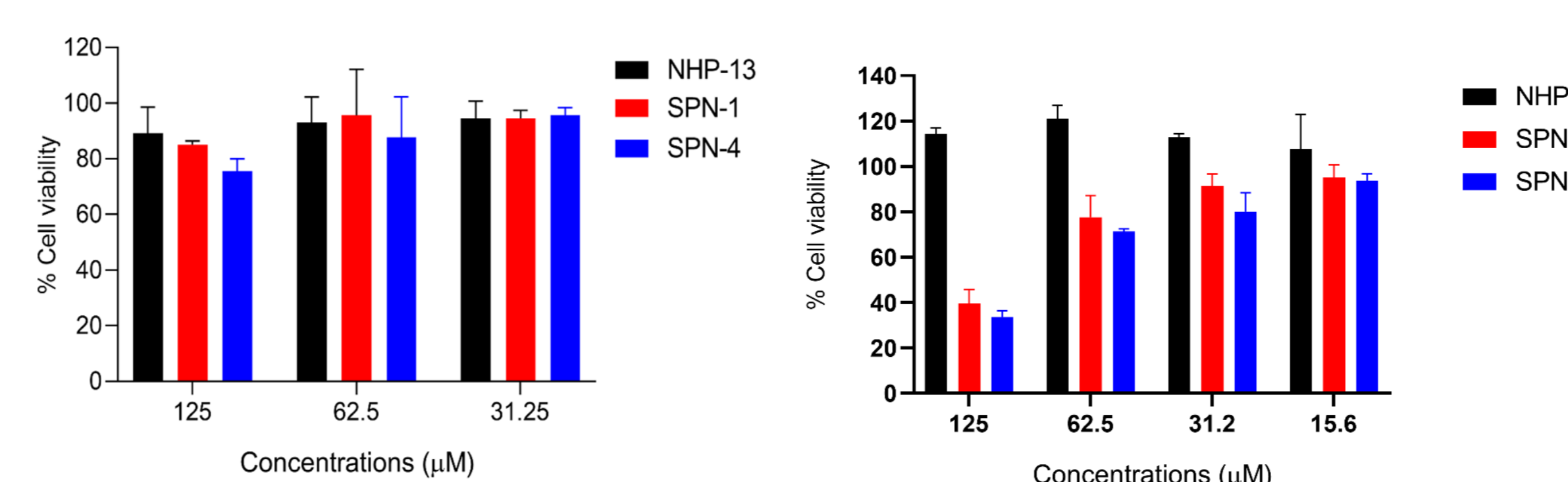


Fig 3.2: (a) Cytotoxicity assay of the peptides against HEK-293T cell lines. (b) Cytotoxicity assay of the peptides against peripheral blood mononuclear cells (Primary cells). The assay was performed based on reduction of alamar blue dye.

4. In-vitro antiendotoxin neutralization assay

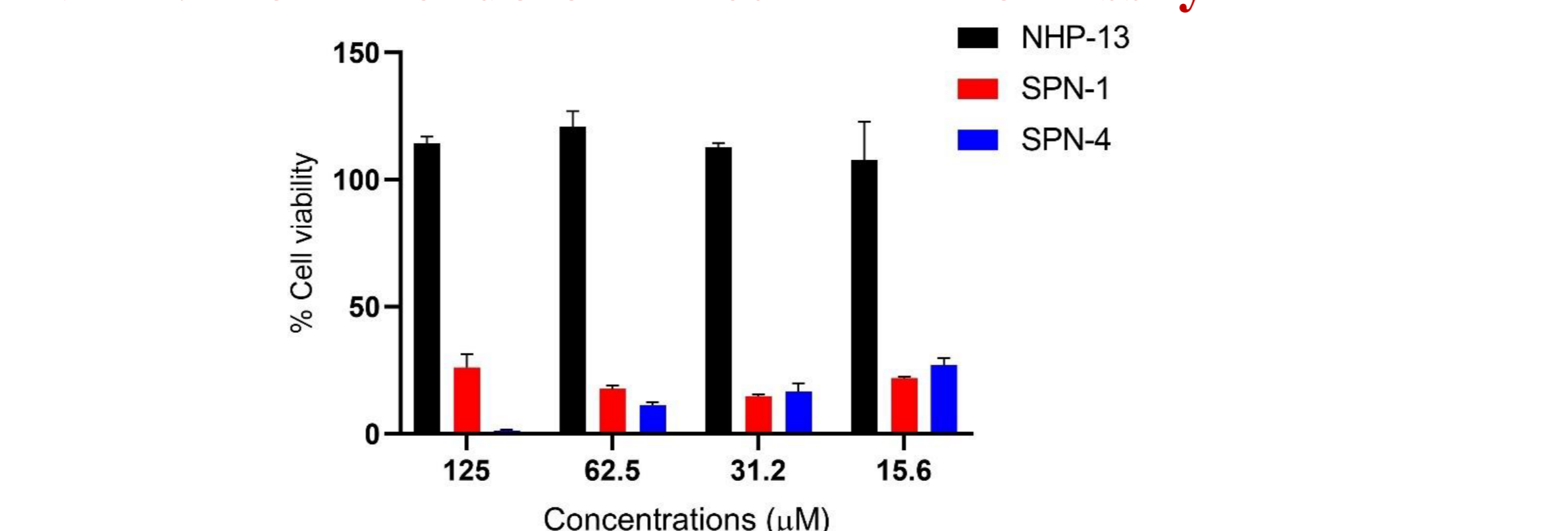


Fig 4.1: (b) Cytotoxicity assay of peptides against THP-1 cells by alamar blue assay.

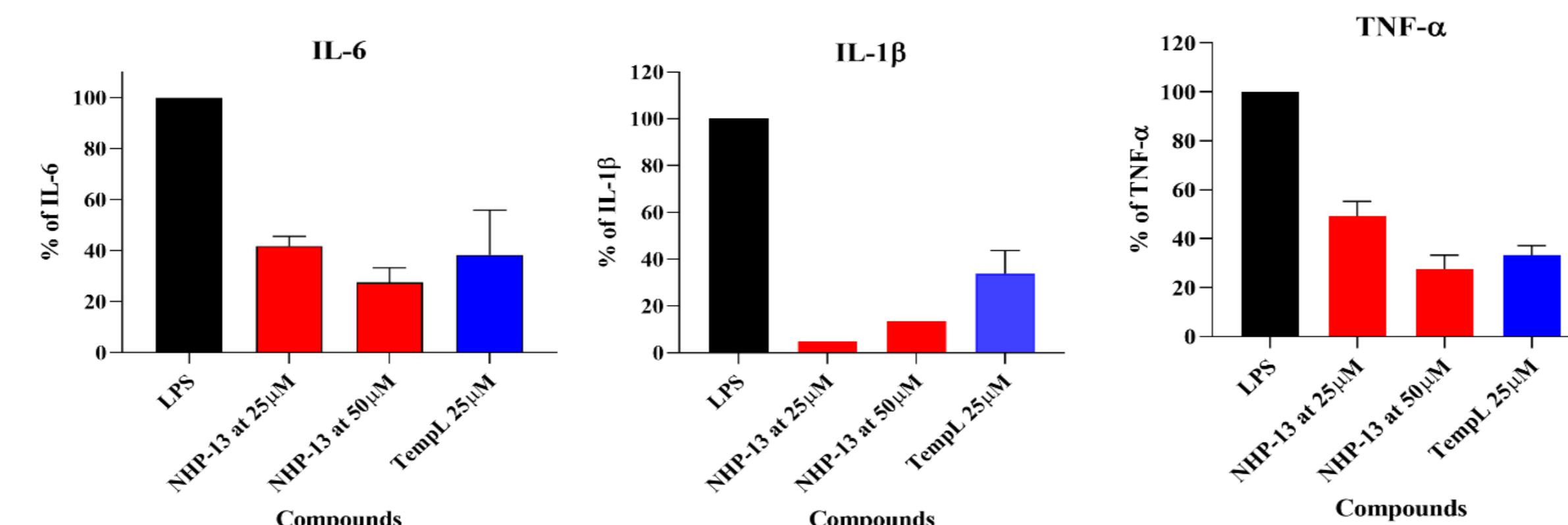


Fig 4.2: Effect of peptide treatment on the production levels of pro-inflammatory cytokines in THP-1 cells by ELISA experiments.

Results

5. Drug resistance studies

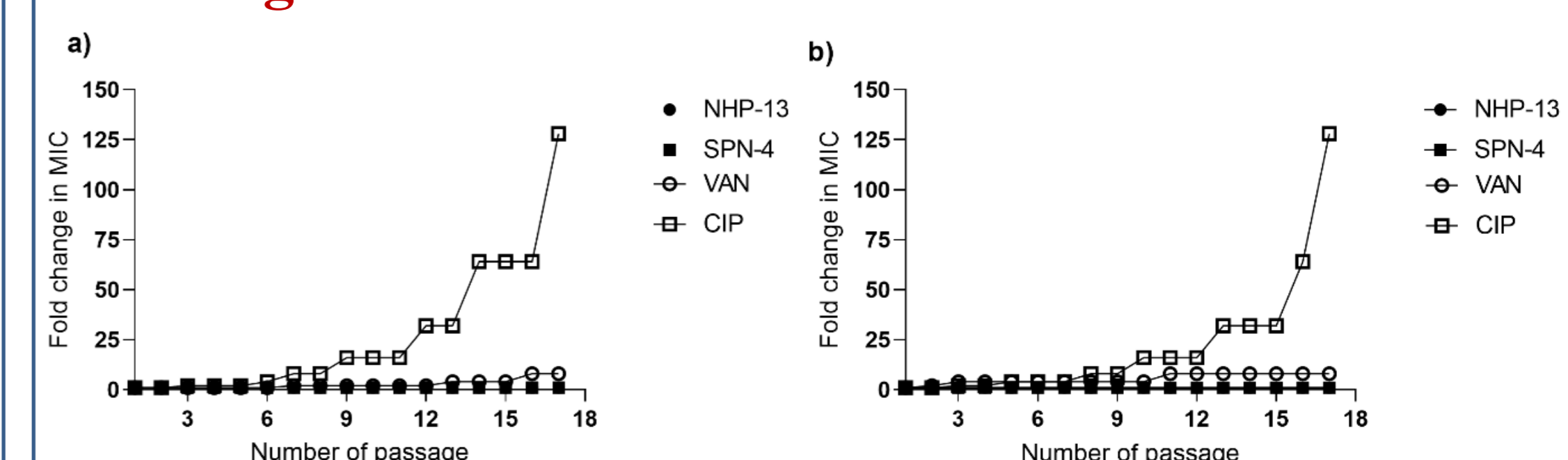


Fig 5: Resistance development studies of the peptides till 17 passages of bacteria. a) for *E. coli* ATCC 11775 and b) for *S. aureus* ATCC 33591.

6. Mode of action studies

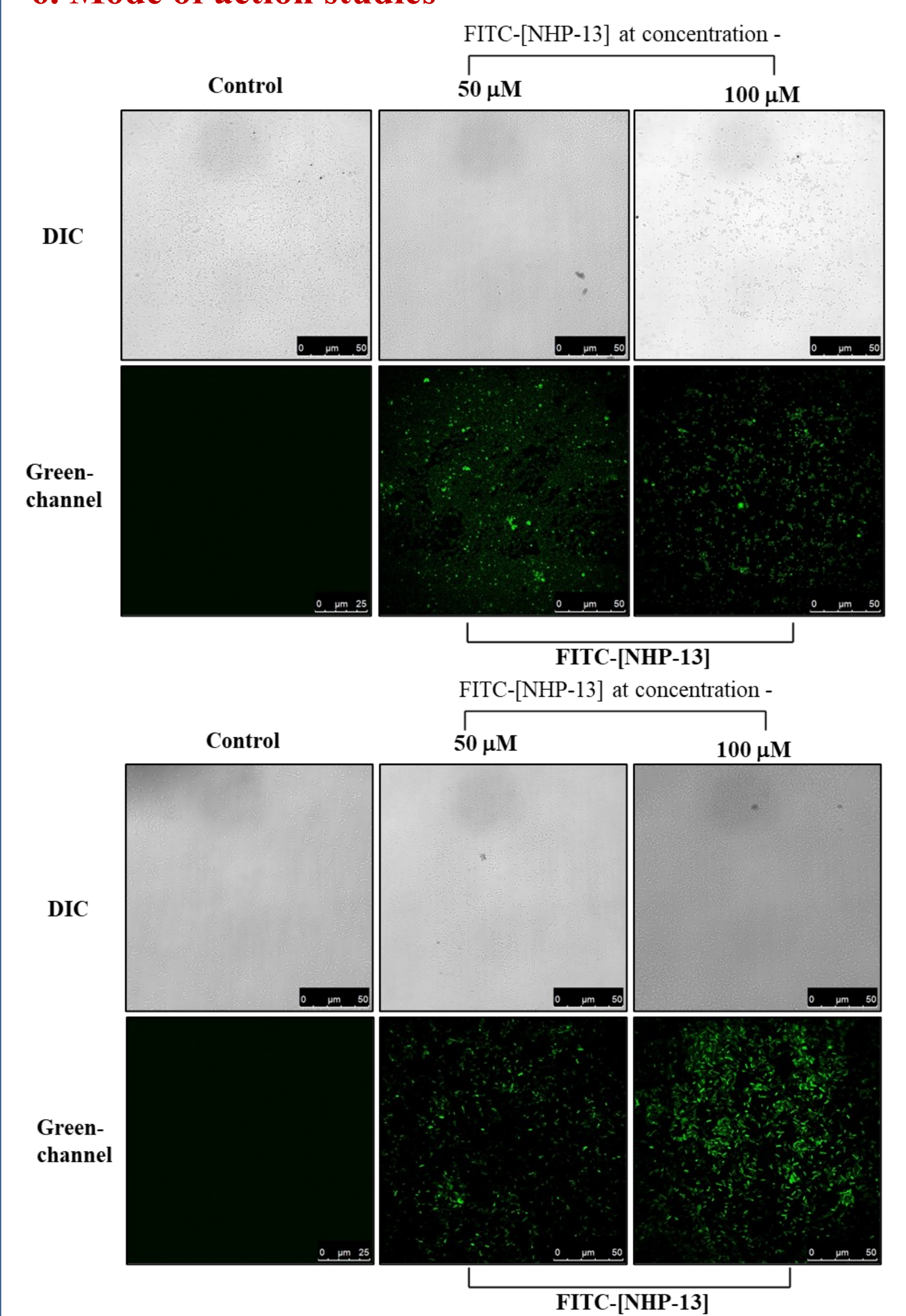


Fig 6.1: Peptide internalization studies by Confocal Microscopy.

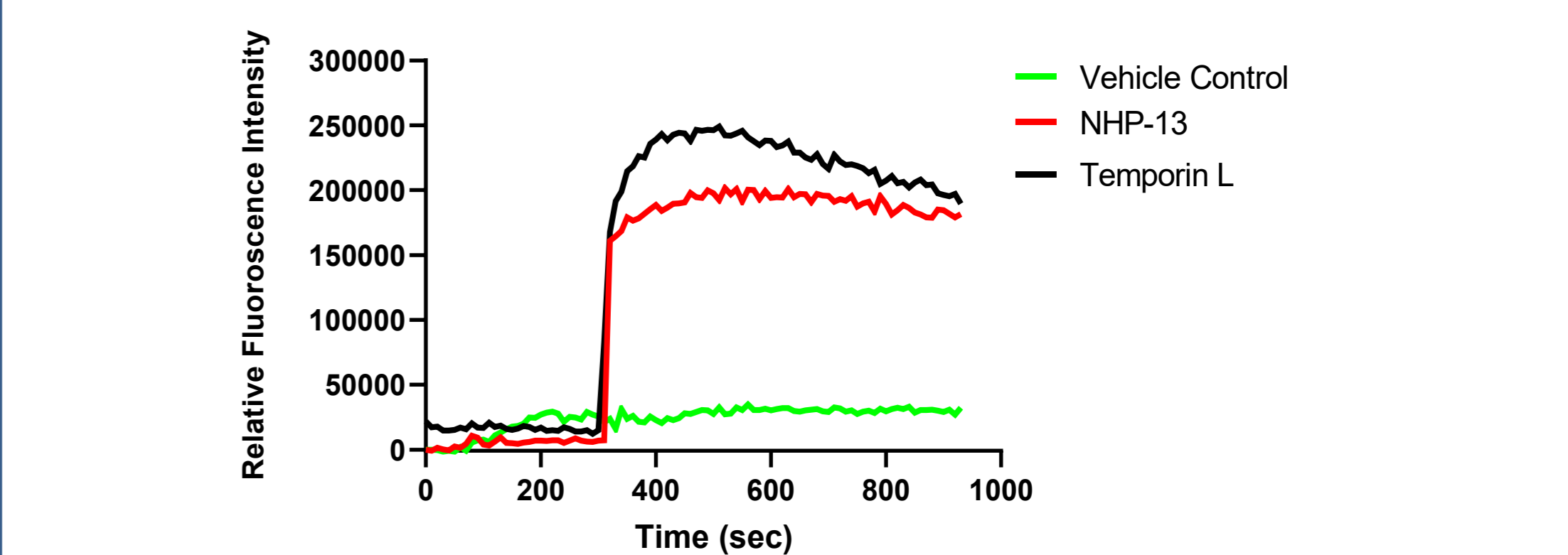


Fig 6.2: Depolarization assay of NHP-13 peptide in live MRSA cells (at 2 × MIC).

Key findings

- The peptide templates, PRRPPR and QLDLY were employed to design for dual inhibition of protein and DNA synthesis, respectively.
- Peptide NHP-13 and SPN-4 was found to be active in the MIC range of 7.8-125 μM concentrations against tested strains with no hemolytic activity observed up to 500 μM concentrations.
- Further studies was performed to evaluate peptide internalization in bacteria by labelling with FITC.
- The most active peptide (NHP-13) was hybrid of above two templates with linker of triple arginine, highlighting arginine's role in cell penetration and enhancing antibacterial activity.
- In vitro* LPS neutralization activity of NHP-13 shown potential effects on reduction in inflammatory cytokines in human monocytes.
- Further studies are going on for evaluation of protein synthesis by interaction with 70s ribosomes and DNA replisomes.

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