

https://doi.org/10.17952/37EPS.2024.P2078

# **Characterization of Aurora-A Activation and Allosteric Regulation by TACC-3 Peptidomimetics**

# **Diana Gimenez**,<sup>a,b</sup> Martin Walko,<sup>b,d</sup> Jennifer A. Miles,<sup>c,d</sup> Richard Bayliss,<sup>c,d</sup> Megan H. Wright<sup>b,d</sup> and Andrew J. Wilson<sup>a,b,d,\*</sup>

<sup>[a]</sup> School of Chemistry. University of Birmingham, Birmingham B15 2TT, UK. <sup>[b]</sup> School of Chemistry and <sup>[c]</sup> Faculty of Biological Sciences, University of Leeds, Leeds, LS2 9JT, UK. <sup>[d]</sup> Astbury Centre for Structural Molecular Biology, University of Leeds, Leeds LS2 9JT, UK.

**TACC3 / Aur A Protein-Protein Interaction** 

Aurora-A is a Ser/Thr protein kinase that regulates key mitotic events by recruiting and phosphorylating a range of different intrinsically disordered proteins. Among them, the Aurora-A interaction with TACC-3 (Transforming Acidic Coiled-Coil Containing Protein 3) is instrumental for spindle assembly and chromosome segregation,<sup>2,3</sup> and is frequently upregulated in many different types of cancer.<sup>4</sup>





Figure 1. Key features of the Aurora-A/TACC-3 complex. (a) Crystal structure of the Aurora-A catalytic domain (green) in complex with TACC-3<sub>522-563</sub> (orange; PDB: 50DT). (b) Sequence of TACC-3 docking region to Aurora-A, TACC-3 <sub>522-563</sub>, and individual FA dissociation constants (K<sub>d</sub>) for each domain. (c-d) MD calculated energy minimum structure of WT TACC-3<sub>522-536</sub> in the presence of Aurora-A, showing the key Phe525<sup>TACC-3</sup> and Pro528<sup>TACC-3</sup> exo-pucker conformation.

## **Optimization of Phe525 and Pro528 interaction with Aur-A**

By using unnatural phenylalanine analogs and exploiting stereo-electronic effects (i.e. gauche effect) twelve-fold improved Kd values were measured for new constrained peptides when compared to the linear sequences, with all variants showing low micromolar/high nanomolar affinities.











E-mail: d.gimenez-ibanez@bham.ac.uk

Bph constraints restrict the conformational landscape and orientates the key <sup>525</sup>Phe predisposing the peptide towards Aur A binding

Enhanced binding of peptidomimetics to Aur-A is entropically driven

To explore how restricting the accessible conformations influences the thermodynamics of binding, we carried out isothermal titration calorimetry (ITC) and variable-temperature FA experiments. Constrained variants exhibited a more favorable entropy of binding in comparison to the linear variants.





Figure 2. Key features of TACC-3 constrained peptidomimetics. (a) Iodinated (4-I)-Phe525<sup>TACC-3</sup> in its binding pocket. (b) Exo-pucker conformation of fluorinated trans-(4-F)Pro528<sup>TACC-3</sup> in its binding pocket.

**TACC-3**<sub>518-532</sub> peptidomimetics stimulate Aur-A

Linear WT variants are relatively poor activators of the kinase, whereas the constrained peptidomimetics stimulate Aurora-A and promote substrate phosphorylation in a dose-dependent manner by up to 160-190%.





Figure 3. Thermodynamics of binding in the presence of Aurora-A. (a) ITC thermodynamic signatures of linear TACC-3<sub>518-532</sub> and (b) constrained TACC-3<sub>518-532-S/E-Bph-I/fP</sub> binding to Aurora-A<sub>122-403-C290A/C393A</sub>; (c) Diagram schematically illustrating the hypothetical free energy profile of a one-step/one-barrier peptide-protein binding event for a linear and a constrained peptide.

TACC-3<sub>518-532-L/R-Bph-I/fP</sub> as an allosteric inhibitor of the N-MYC/Aur-A interaction

Constrained peptidomimetics exhibited limited evidence of TPX2 displacement, indicating promising specificity for the TACC-3 binding site (IC50 >> 200 µM). Surprisingly, the constrained variants were observed to out-compete N-MYC.



Figure 3. Kinase activation by peptidomimetics. (a) Qualitative kinase assay monitoring the autophosphorylation of unphosphorylated Aurora-A in the presence of TACC-3<sub>522-536</sub> and constrained peptides (all peptides 10 μM). (b) % Kinase activation of Aurora-A (10 nM) in the presence of linear and constrained TACC-3 peptides as measured in ADP-Glo assays.

Figure 7. Constrained TACC-3<sub>518-532\_Bph\_LR\_IF/fP</sub> as an allosteric inhibitor of N-MYC/Aurora-A PPI: (a) Aligned x-ray crystal structures of TACC-3/Aurora-A complex (PDB: 50DT), TPX2 (PDB: 10L5), and N-MYC (PDB: 5G1X); (b) FA competition assay of control N-MYC<sub>61-89</sub> (black line) and constrained TACC-3 518-532\_Bph\_LR\_IF/fP (forest green) against FAM-Ahx-N-MYC<sub>61-89</sub> (200 nM), and reverse assay in the presence of Aurora-A; (c) Competition FA N-MYC IC50's values for N-MYC<sub>61-89</sub> at increased TACC-3<sub>518-532 Bph LR IF/fP</sub> concentrations; (d) X-ray crystal structures of TACC-3/Aurora-A and N-MYC/Aurora-A complexes.

Conclusions We have developed a series of constrained peptides that inhibit TACC3/Aurora A PPI with up to 12-fold enhanced IC<sub>50</sub>/K<sub>d</sub> values. NMR and MD analysis of the peptides show that improved affinities are achieved due to the combined effect of restricting the conformational landscape of the peptide (Bph constraints), fine-tunning the orientational preference of the key Phe525 (4) and 4Br-Phe substitutions) and conformational pucker-ring selection at the Pro528 level (4F-Pro). Our minimal peptidomimetics are sufficient to induce the conformational changes needed to activate the kinase by binding only the "F" pocket on the N-lobe, and to allosterically displace N-MYC from its binding site. Knocking-out the TACC-3/Aurora-A interaction without affecting kinase activity represents a promising alternative to active-site kinase inhibition. Similarly, inhibition of the Aurora-A/N-MYC interaction without downregulating other Aurora-A essential functions also represents a major target for anticancer drug-development.

### References

1. Barr, A. R.; Gergely, F. J Cell Sci. 2007, 120 (Pt 17), 2987–2996. 2. Burgess, S. G.; Peset, I.; Joseph, N.; Cavazza, T.; Vernos, I.; Pfuhl, M.; Gergely, F.; Bayliss, R. PLOS Genetics. 2015, 36. 3. Burgess, S. G.; Mukherjee, M.; Sabir, S.; Joseph, N.; Gutiérrez-Caballero, C.; Richards, M. W.; Huguenin-Dezot, N.; Chin, J. W.; Kennedy, E. J.; Pfuhl, M.; Royle, S. J.; Gergely, F.; Bayliss, R. The EMBO Journal. 2018, 37 (8). 4. Ha, G.-H.; Kim, J.-L.; Breuer, E.-K. Y. Cancer Letters. 2013, 336 (1), 24–335.

#### Acknowledgements

This work is financed by the Biotechnology and **Biological Sciences Research Council (BBSRC)** BB/V003577/1



**Biotechnology and Biological Sciences Research Council**