

Peptide Nanonets as Antimicrobial and Anti-inflammatory Agents

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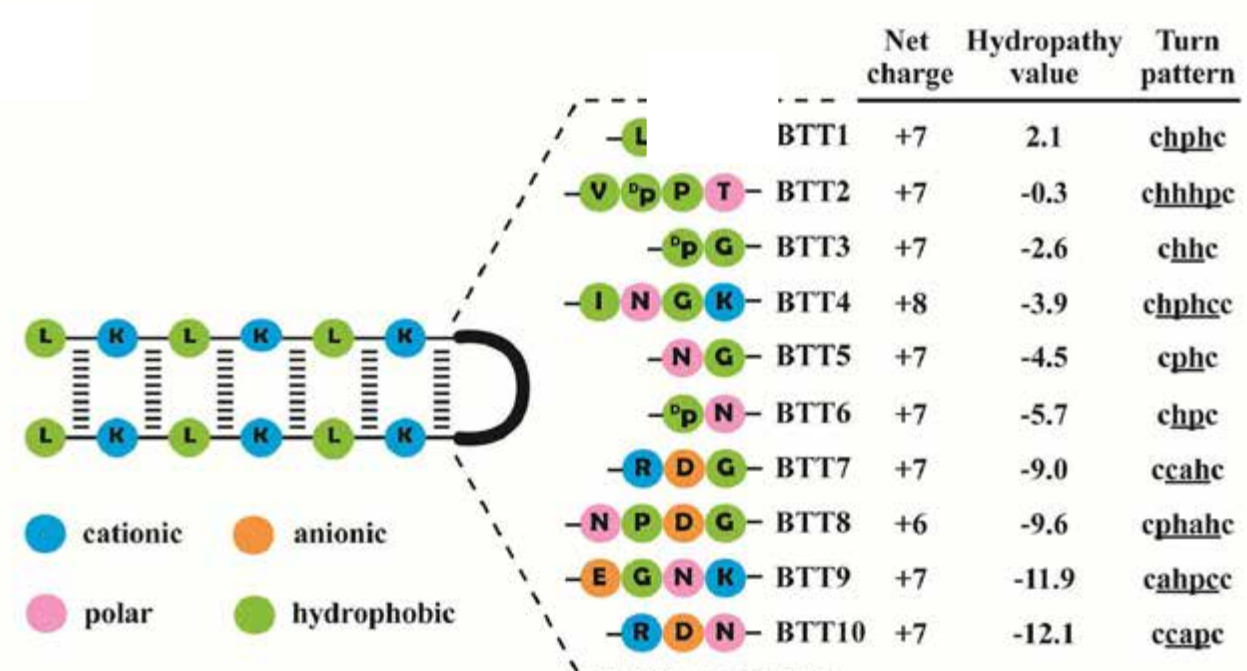
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De novo design of beta-hairpin peptides

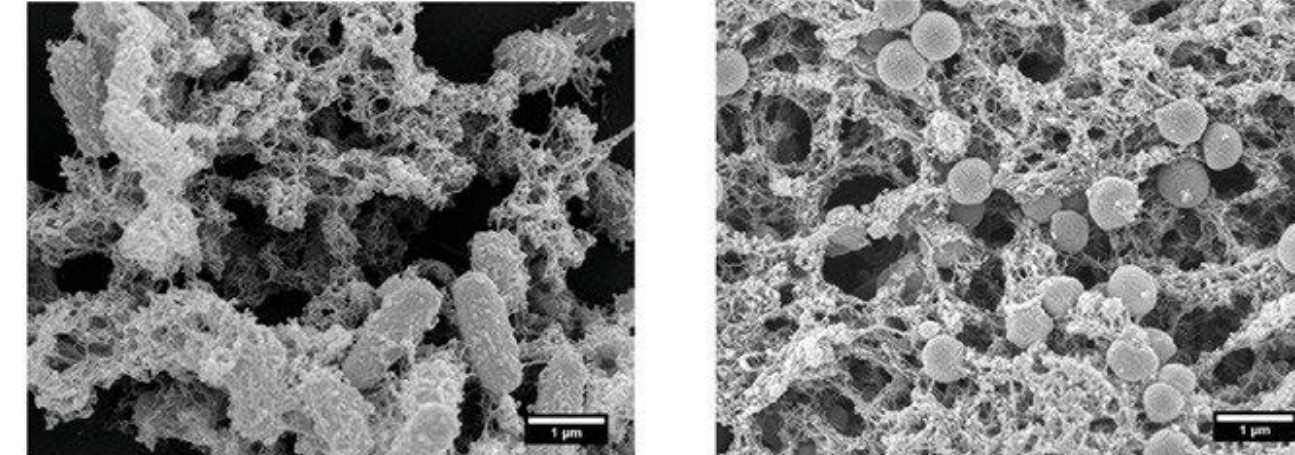


We have designed a series of de novo β -hairpin peptides with identical side strands but varied turns. We demonstrated that mutations of only 2 to 4 amino acids at the turn region could impart a wide range of antimicrobial profiles among synthetic β -hairpin AMPs.

Through turn sequence modification by Ala scanning, we identified specific analogues capable of forming bacteria-responsive nanonets.

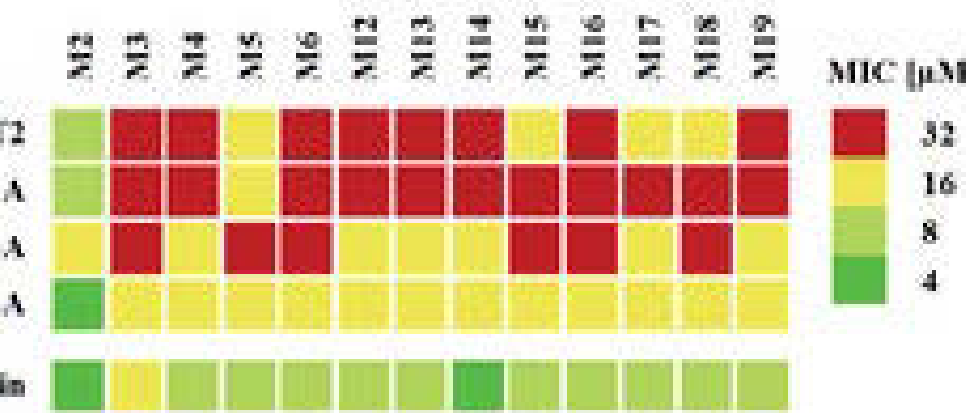
Nanonet forming analogues have anti-microbial activity

Peptide	MIC [μ M]	
	<i>E. coli</i>	<i>S. aureus</i>
BTT1	4	8
BTT1-3A	8	16



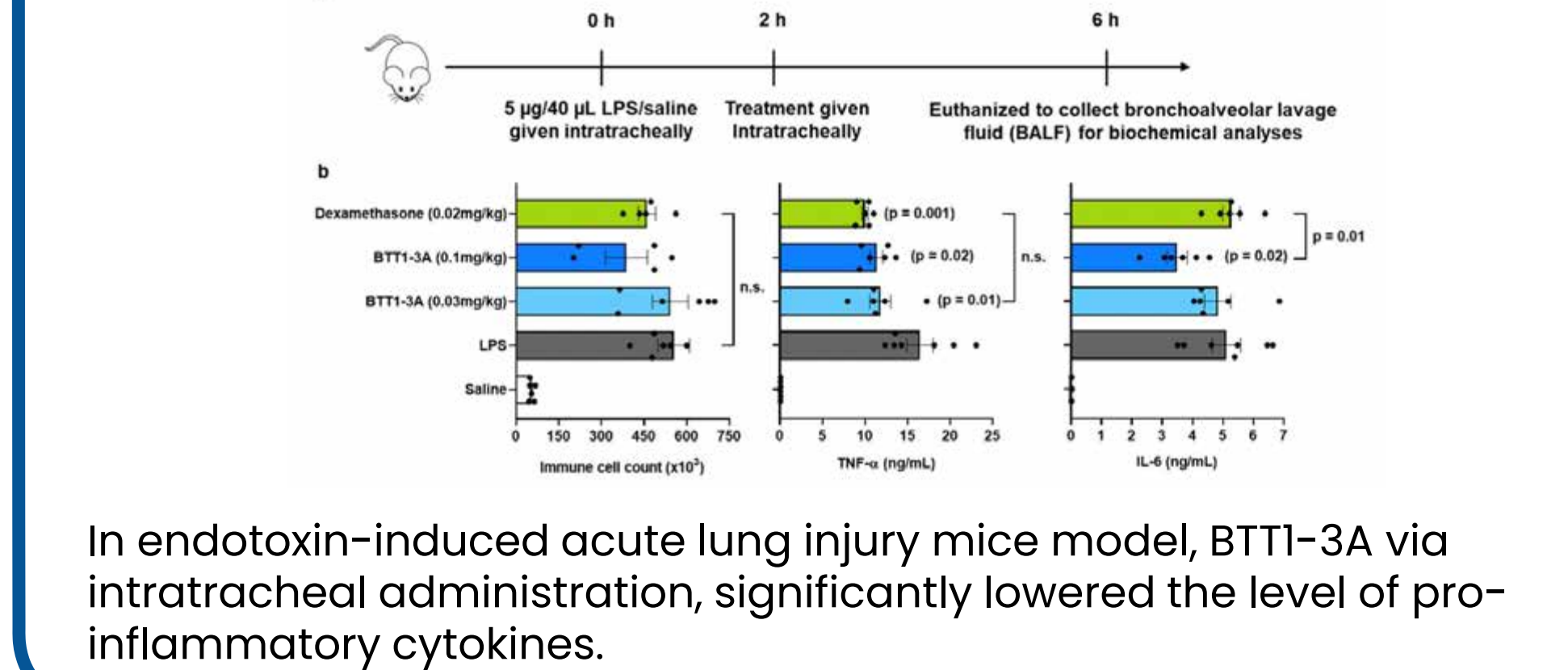
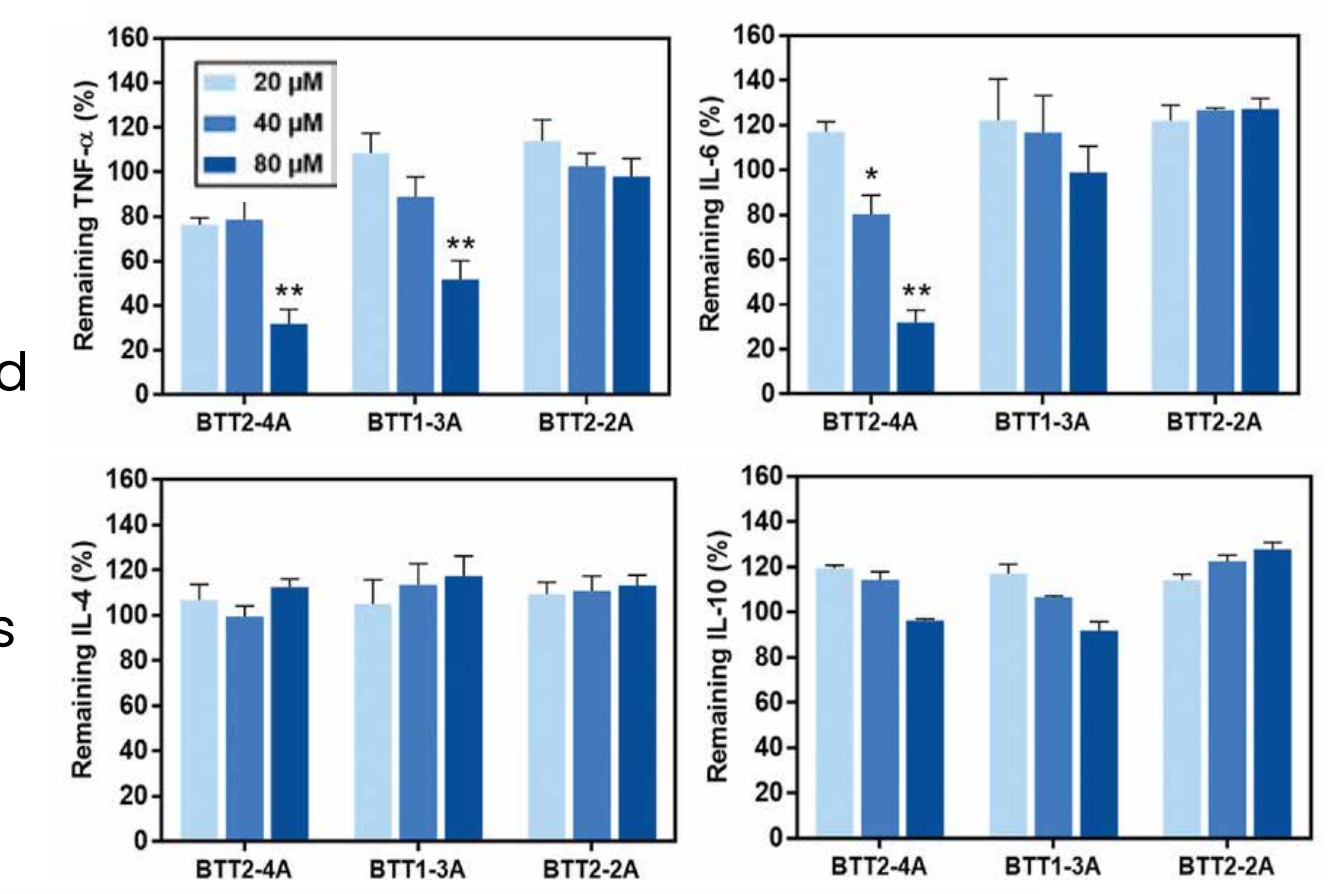
BTT1-3A was found to be the only analogue capable of forming bacteria-responsive nanonets. We successfully converted the kill-only BTT1 to the trap-and-kill BTT1-3A.

By replacing single amino acid at the run segment of BTT2, we obtained BTT2-4A with improved antimicrobial potency against clinically relevant antibiotic-resistant strains, while preserving the ability to form bacteria-trapping nanonets.



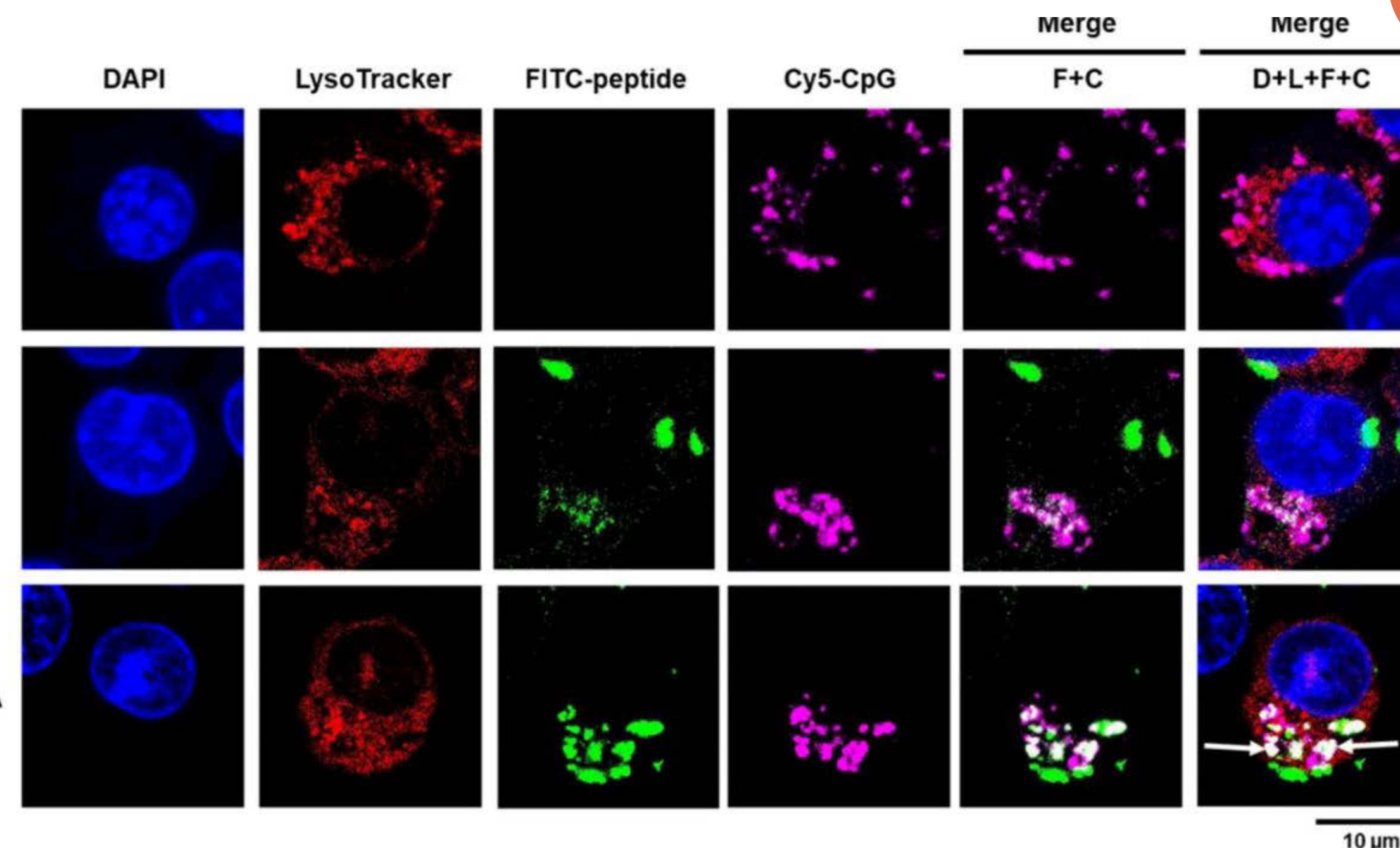
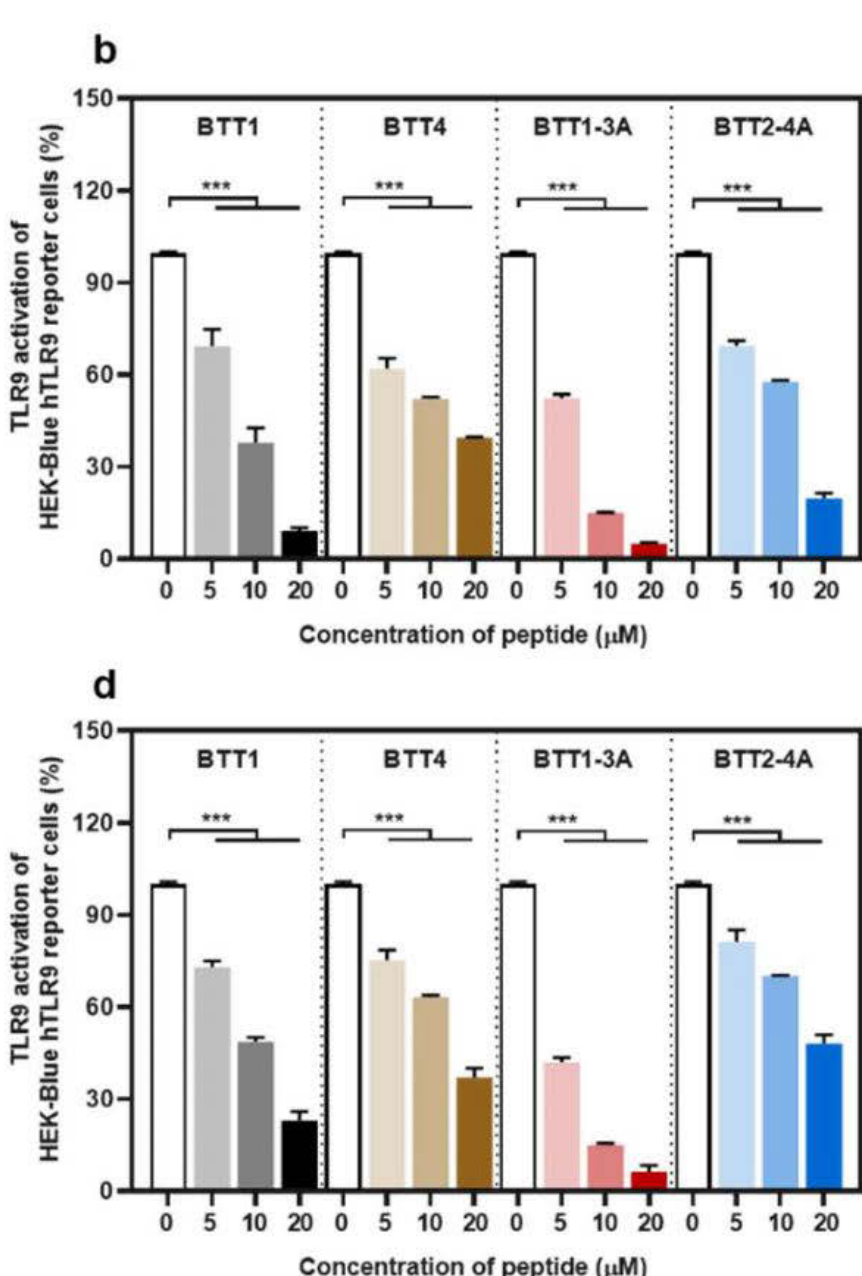
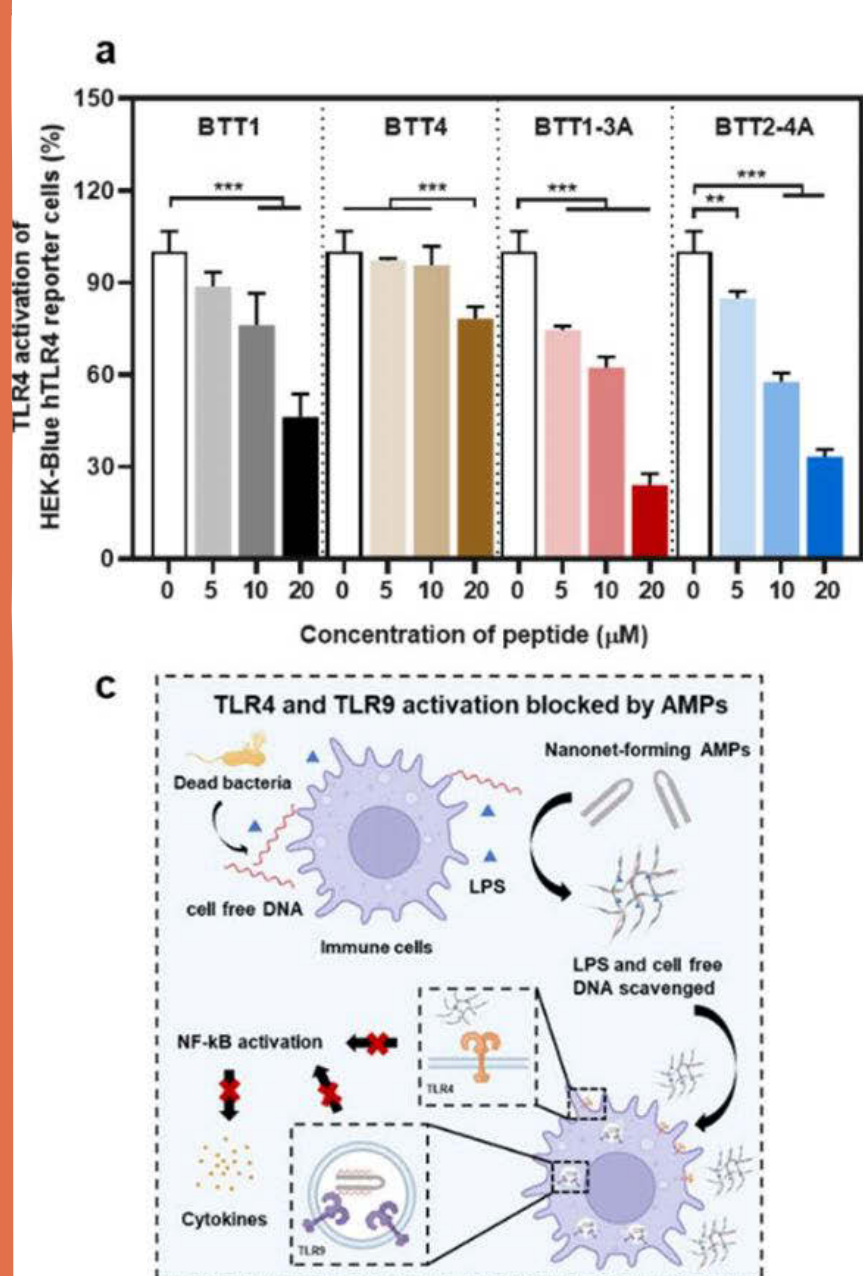
Nanonet forming analogues are anti-inflammatory

BTT2-4A and BTT1-3A nanonets also displayed selective trapping activity toward pro-inflammatory cytokines while not affecting anti-inflammatory cytokines



In endotoxin-induced acute lung injury mice model, BTT1-3A via intratracheal administration, significantly lowered the level of pro-inflammatory cytokines.

Nanonet-forming peptides inhibit TLR4 and TLR9 activations in cells by inflammatory mediators



Co-localization of BTT peptides with intracellular CpG DNA in RAW264.7 cells. Most of FITC-BTT1-3A but less of FITC-BTT4 intracellularly co-localized with CpG DNA and endo-lysosomes, demonstrating that BTT1-3A had stronger capacity of entering cells and competitively binding with intracellular CpG DNA than BTT4.

(a) BTT peptides inhibited TLR4 activation in HEK-Blue™ hTLR4 cells by entrapping extracellular LPS. (b, d) BTT peptides inhibited TLR9 activation in HEK-Blue™ hTLR9 cells by scavenging extracellular (b) and intracellular (d) CpG DNA. (c) Scheme of nanonet-forming AMPs blocking TLR4 and TLR9 activations and the downstream NF- κ B pathway.

Objective: To explore anti-inflammatory activity via inhibition of TLR4 and TLR9 pathways

Methodology:

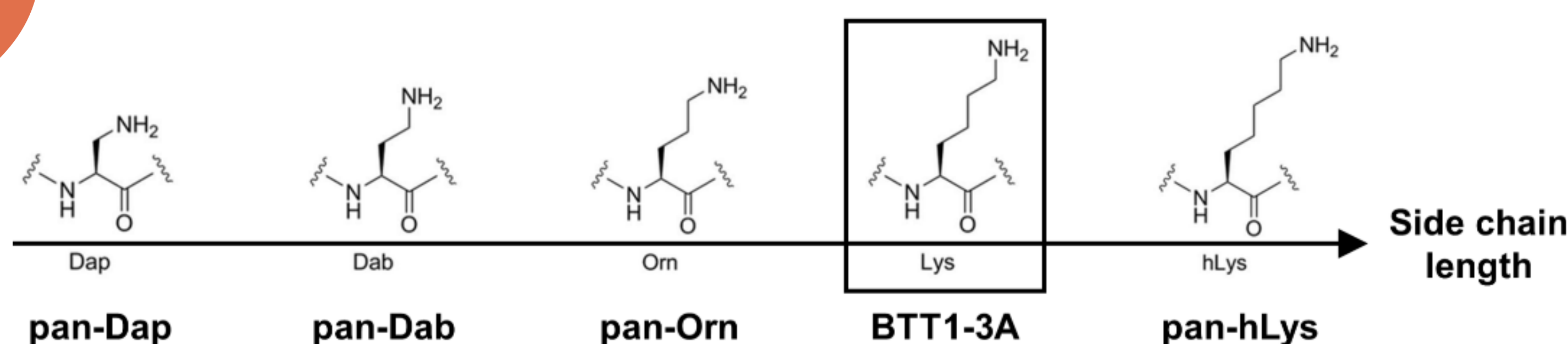
- CpG DNA binding assay
CpG DNA was incubated with PicoGreen and peptide solutions. Fluorescence intensity was measured.
- Cellular co-localization of peptides with CpG DNA
Overnight culture of RAW 264.7 cells were incubated with 1 μ g/mL of Cy5-CpG DNA for 1 hour. FITC-peptides were then added in the medium and incubated for 4 hours. The cells were stained by LysoTracker Red DND-99, followed by fixation with 4% formaldehyde in PBS and staining with DAPI.

Objective: To improve the proteolytic stability of β -hairpin antimicrobial peptide

Methodology:

- Minimum inhibitory concentration (MIC) determination
MIC of peptides was determined by broth microdilution method. The MIC presented the lowest peptide concentration at which at least 90% of OD600 was reduced compared to control without peptide treatment.
- Field-emission scanning electron microscope (SEM) was employed for the observation of bacteria-trapping nanostructures.
- NPN (N-phenyl-naphthalen-1-amine) uptake assay for outer membrane permeability; SYTOX green uptake assay for indication of inner membrane permeability.

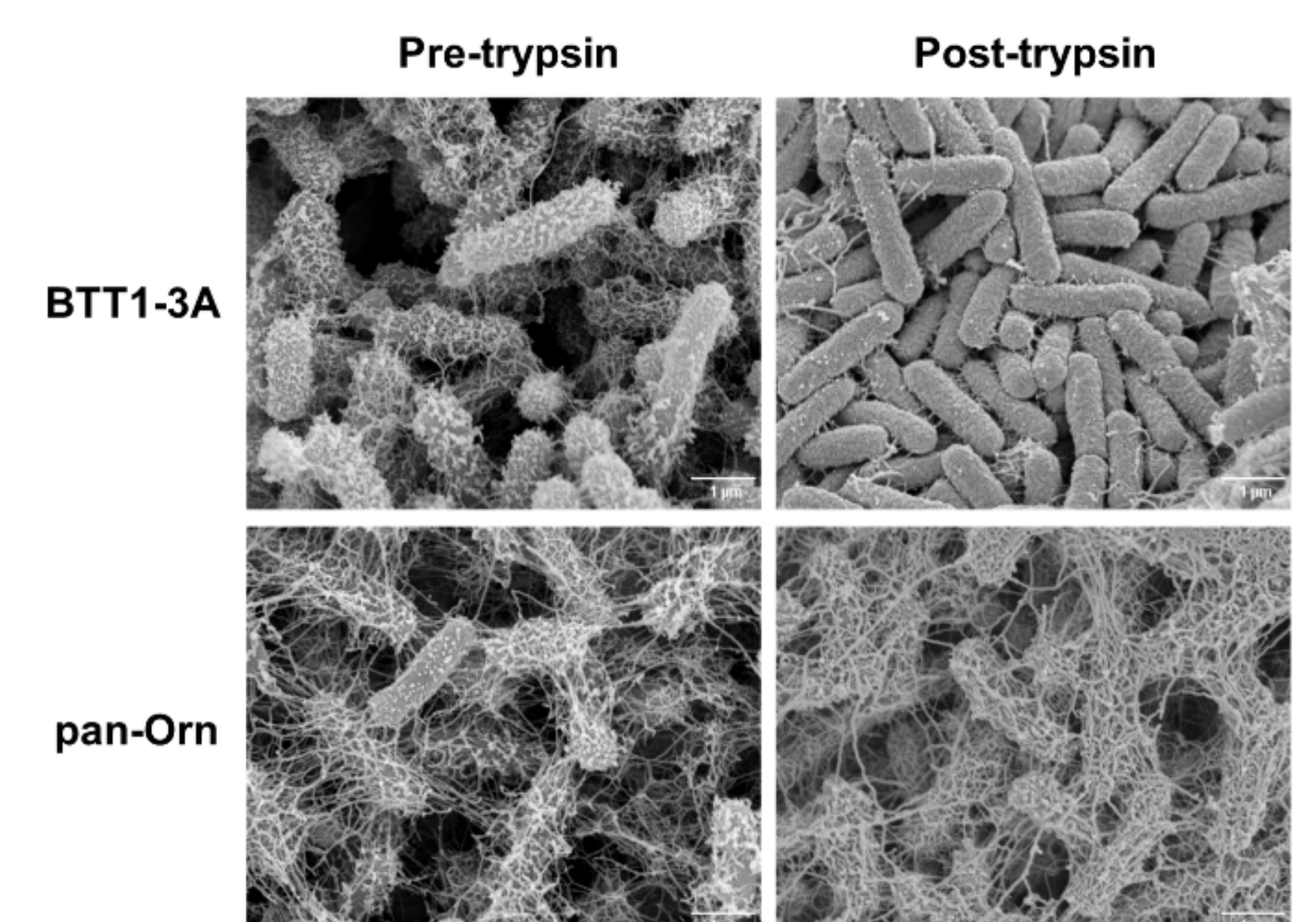
Pan-substitution with Ornithine confer robust enzymatic stability to BTT1-3A while retaining activity



Design scheme for pan-substitution of Lysine residue in BTT1-3A using unnatural amino acids with unaltered stereochemistry but different side chain chemistry.

Peptide	MIC against <i>E. coli</i> (μ M)	
	Pre-trypsin	Post-trypsin
pan-hLys	128	> 128
BTT1-3A	16	> 128
pan-Orn	8	8
pan-Dab	64	> 128
pan-Dap	>128	> 128

Without trypsin treatment, only pan-Orn displayed better antimicrobial potency relative to BTT1-3A; following trypsin treatment, pan-Orn retains antimicrobial activity with unchanged MIC, suggesting excellent trypsin stability.



In contrast to BTT1-3A whose nanonet-forming capacity drastically degraded post-trypsin, pan-Orn formed expansive bacteria-trapping nanonets both pre- and post-trypsin, further strengthening the claim on superior proteolytic stability of pan-Orn.