

Design and Structural Analysis of Fluorinated Polyproline-Type Foldamers and their Ability to Interact with Membrane Models

Chloé Cayrou,^{1,2} Simon Gonzalez,^{1,2} Astrid Walrant,³ Delphine Ravault,³ Karine Guitot,^{1,2} Sylvie Noinville,³ Françoise Illien,³ Sandrine Sagan,³ Thierry Brigaud,^{1,2} Olivier Lequin,³ Sandrine Ongerier,² Grégory Chaume^{1,2}

¹ BioCIS, CNRS, CY Cergy-Paris Université, 5 mail Gay-Lussac, 95000 Cergy-Pontoise, France; ² BioCIS, CNRS, Paris Saclay Université, Bat. Henri Moissan, 17 av. des Sciences, 91400 Orsay, France; ³ Laboratoire des Biomolécules, Sorbonne Université, École normale supérieure, PSL University, CNRS, 4 place Jussieu, 75005 Paris, France

chloe.cayrou@cyu.fr

INTRODUCTION

Foldamers are oligomers with a strong tendency to fold into a well-defined secondary structure.¹ Among peptidic foldamers, oligomers of prolines are known to adopt a **polyproline type II helix (PPII)** in water where amide bonds are **all-trans** and a more compact helix (PPI) in organic solvent with all-*cis* amide bonds (fig. 1).

In this work, we report the incorporation of the trifluoromethyl pseudoproline (**CF₃ΨPro**) into a polyproline backbone and the **structural analysis** of the resulting oligomers. Then, in order to increase the interaction with membranes, **amphipathic oligomers** are synthesized with the addition of a guanidyl group on prolines.

all-trans PolyProline II helix
Extended left-handed helix
($\varphi = -75^\circ$, $\psi = 146^\circ$, $\omega = 180^\circ$)
Favored in water

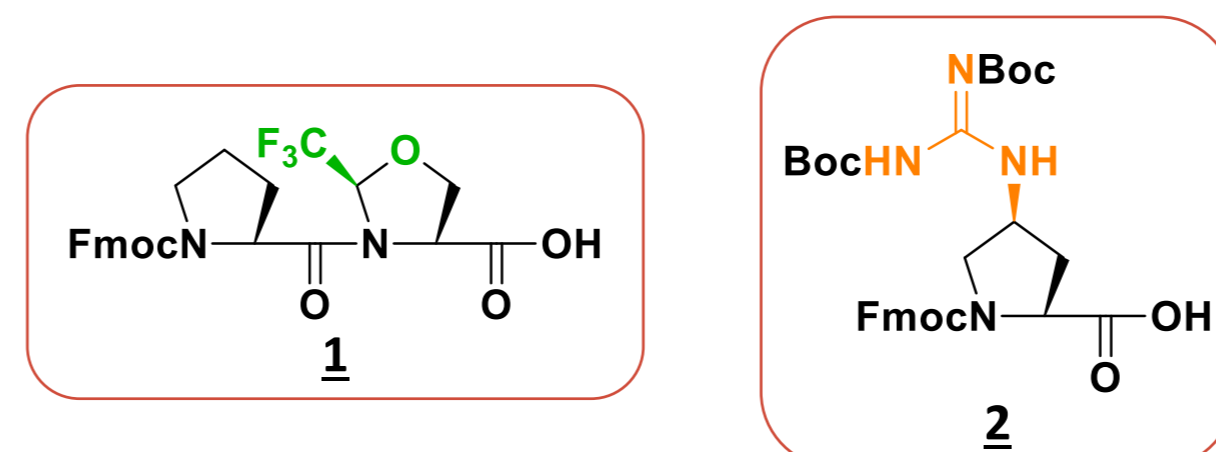
High energy of interconversion

all-cis PolyProline I helix
Compact right-handed helix
($\varphi = -75^\circ$, $\psi = 160^\circ$, $\omega = 0^\circ$)
Favored in organic solvent (*n*-propanol)



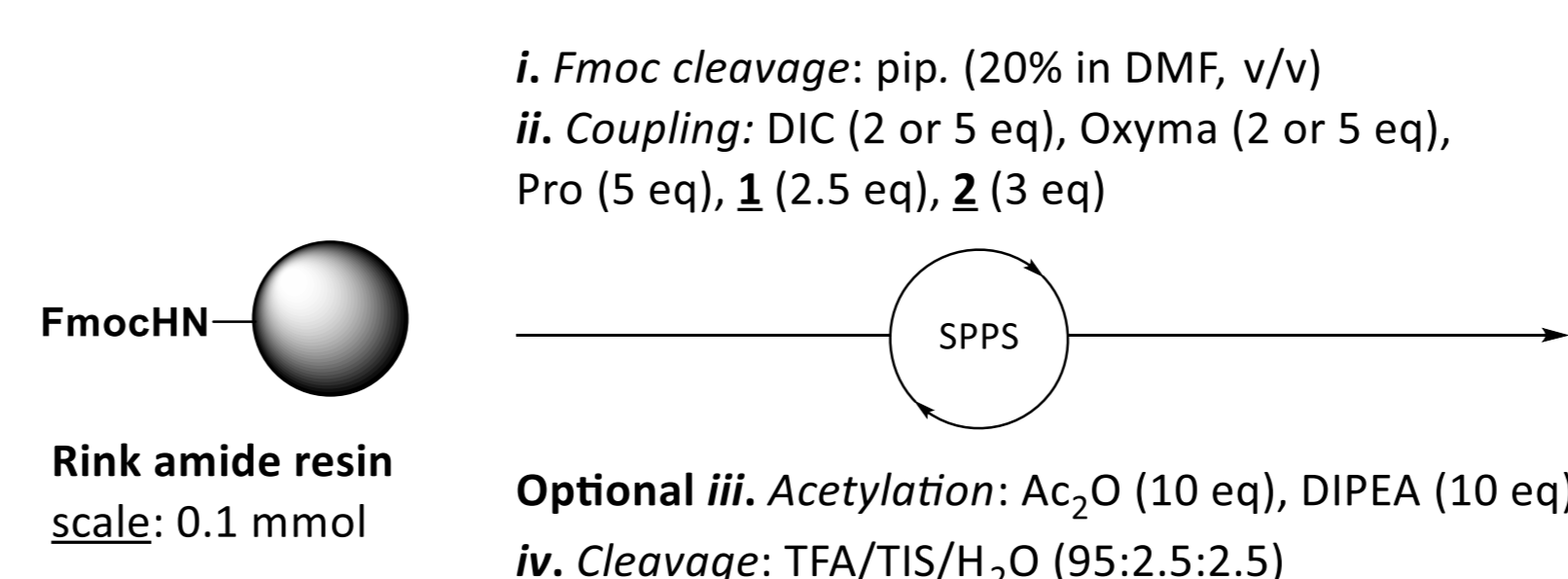
fig. 1: Polyproline secondary structure²

SYNTHESIS

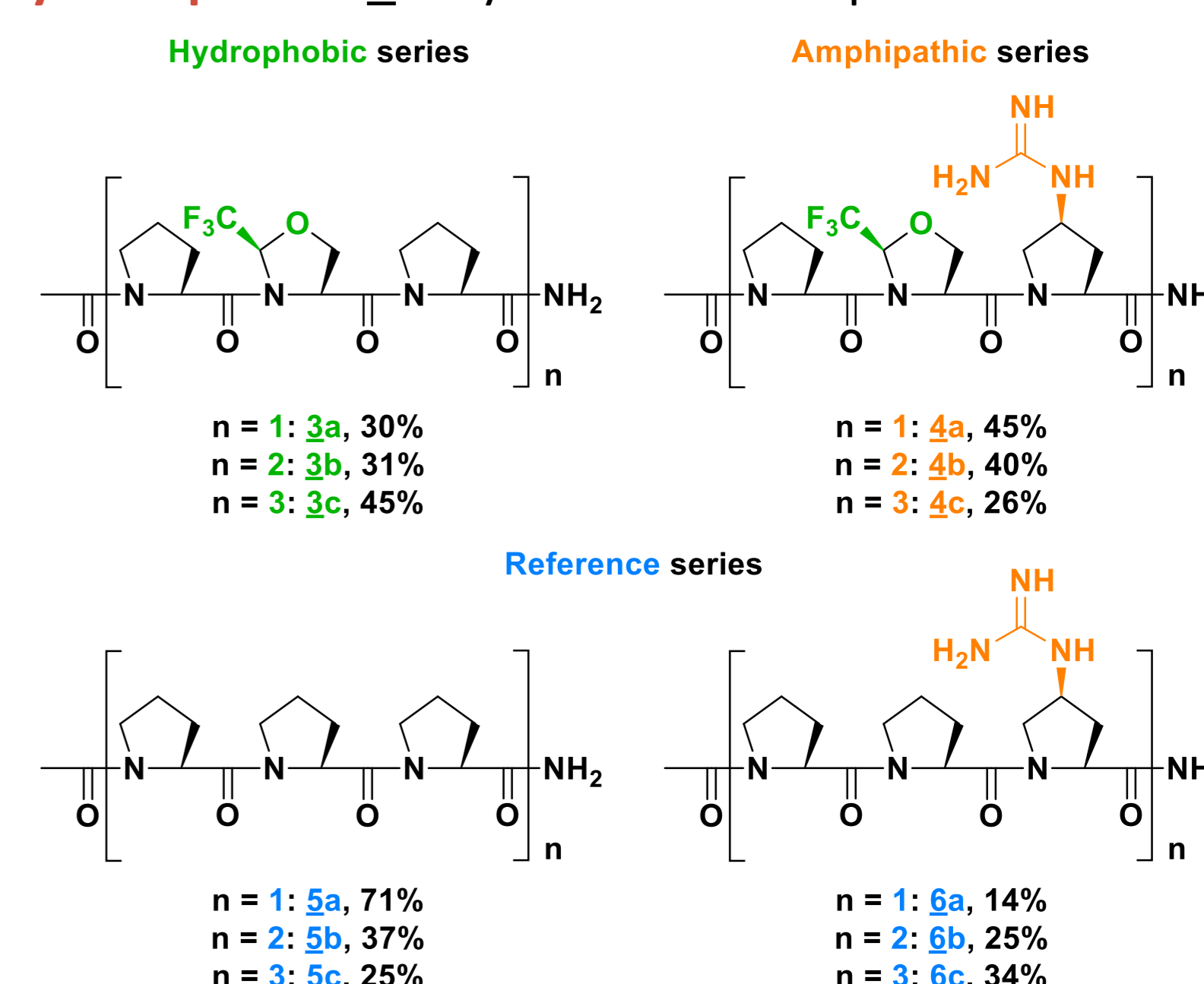


As the CF₃ group decreases the nucleophilicity of the amine, the coupling of the **CF₃ΨPro** is performed in solution to access the ready-to-use **building block 1** for SPPS.³

The **guanidylated proline 2** is synthesized as reported.⁴



Polyproline oligomers are obtained in good yield
(MW activation is used for oligomers **3** and **5**)
(Oligomers **4** and **6** are TFA salt)



NMR ANALYSIS

- Determination of the rotamer populations (¹⁹F NMR).
- Full characterization of fluorinated oligomers (¹H, ¹³C and 2D NMR).
- Assignment of amide bond conformation (NOESY and ROESY NMR, fig. 2 and 3).

	<i>n</i> = 1	<i>n</i> = 2	<i>n</i> = 3
3	All-trans (>95%)	All-trans (>95%)	All-trans (>95%)
4	All-trans (>95%)	All-trans (>95%)	All-trans (>95%)

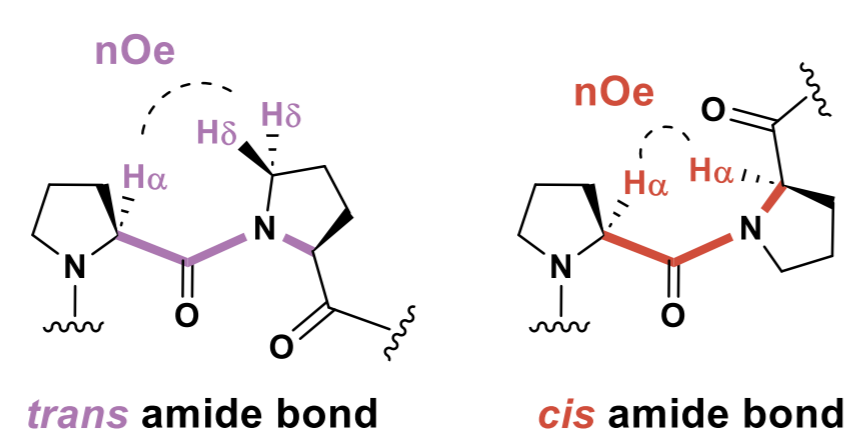


fig. 2: Amide bond ratio of synthesized fluorinated oligomers determined in D₂O at 20°C

fig. 3: Assignment of amide bonds with *n*Oe effect

all-trans amide bond conformation is strongly favored as expected for PPII helix

PPII CD SIGNATURE CONCEPT QUESTIONED FOR OLIGOMERS **3**

Impact of the solvent on the CD signature

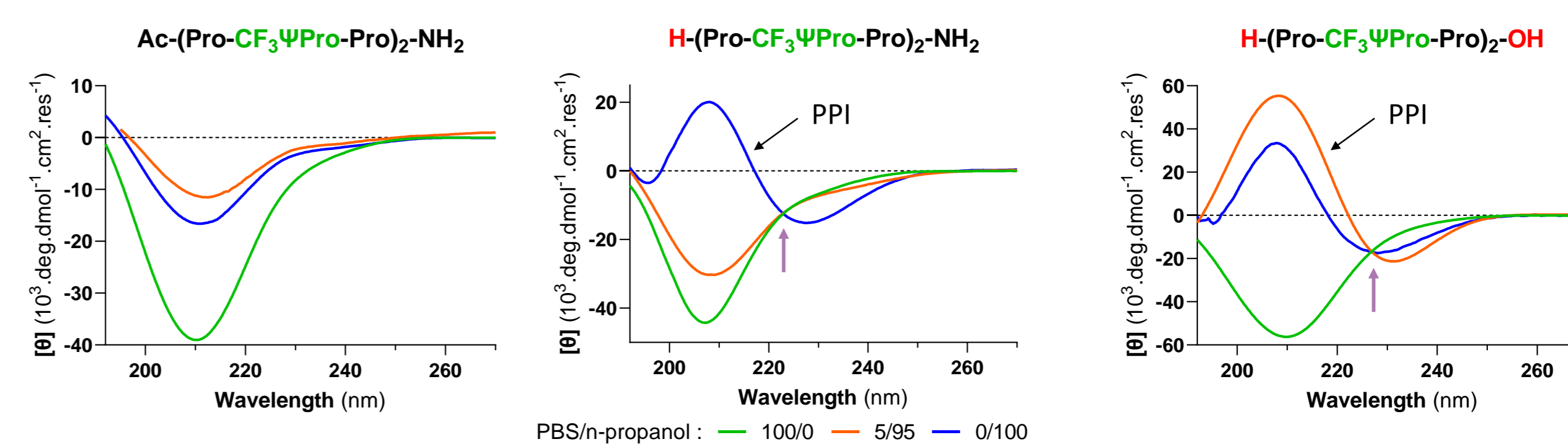
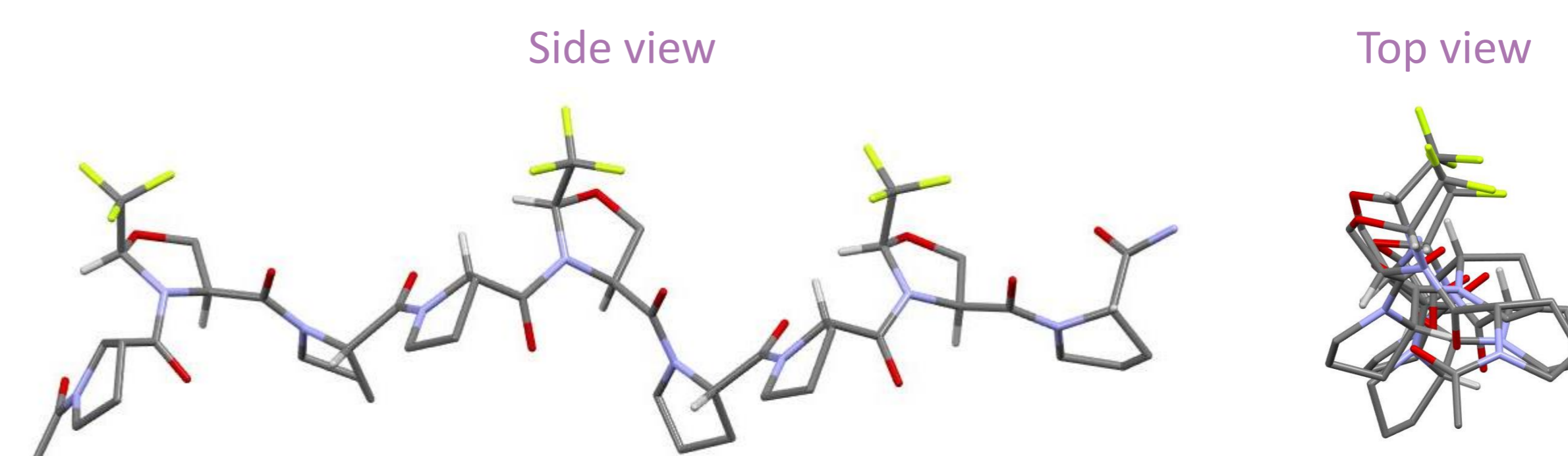


fig. 5: Impact of the *n*-propanol depending on the termini (100 μM at 4°C); — : isobestic point

By increasing *n*-propanol content, **PPI signature is observed** with charged termini (*N*-term: H, *C*-term: OH). Neutral termini (*N*-term: Ac, *C*-term: NH₂) are known to stabilize PPII helix.⁵ Moreover, the presence of **isobestic points** reveal that **only two conformations** are involved in this equilibrium: **PPI ⇌ PPII**.⁶

X-ray structure of **3c**



X-ray structure of **3c** reveals a **left handed helix** with **slightly distorted C₃ symmetry**.

fig. 6: X-ray structure of **3c** (obtained in MeOH with an accuracy of 0.04 Å for bond C-C, against the 0.01 Å expected)

Conclusion

We have assembled a set of CD and X-ray diffraction data consistent with hydrophobic fluorinated oligomers **3** adopting a helical structure close to a PPII helix

CIRCULAR DICHROISM (CD) ANALYSIS

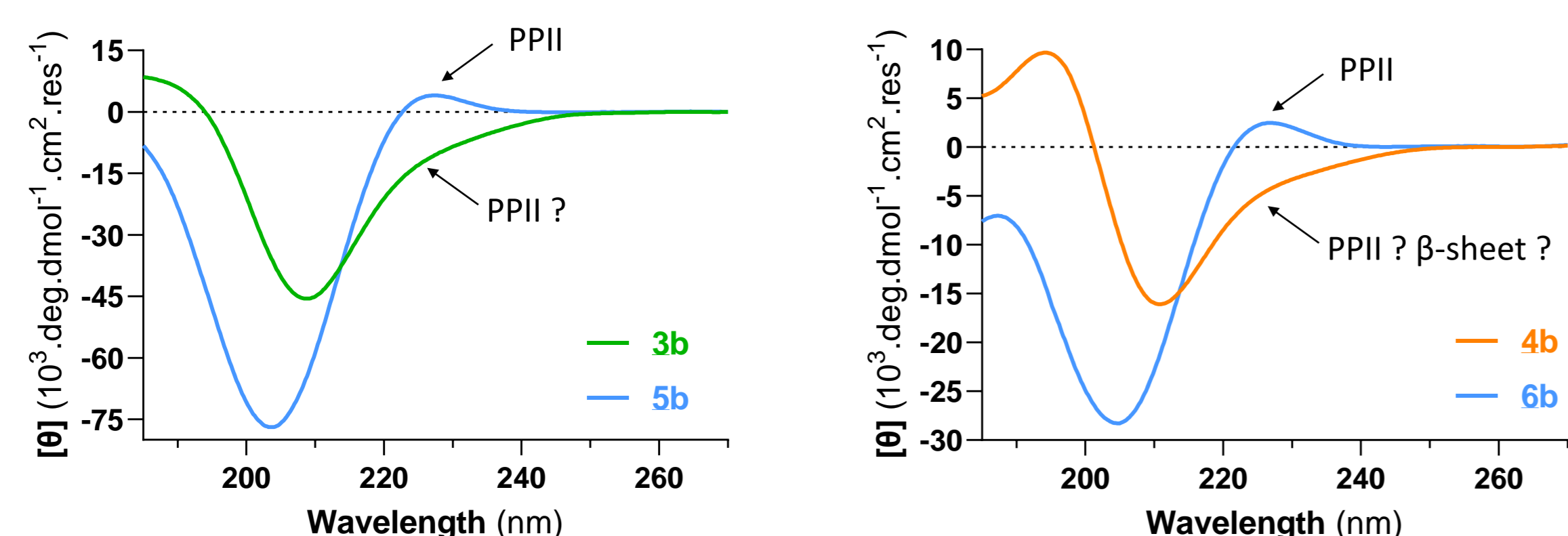


fig. 4: CD spectra of **hydrophobic** (left) and **amphipathic** (right) hexamers with their respective non fluorinated reference (100 μM in PBS 20 mM (pH = 7.0) at 4°C)

CD spectra of fluorinated oligomers **do not reveal typical PPII signature** (absence of positive band at 226 nm)

INTERACTION WITH MEMBRANE MODELS MONITORED BY ¹⁹F NMR (LEFT) AND DSC (RIGHT)

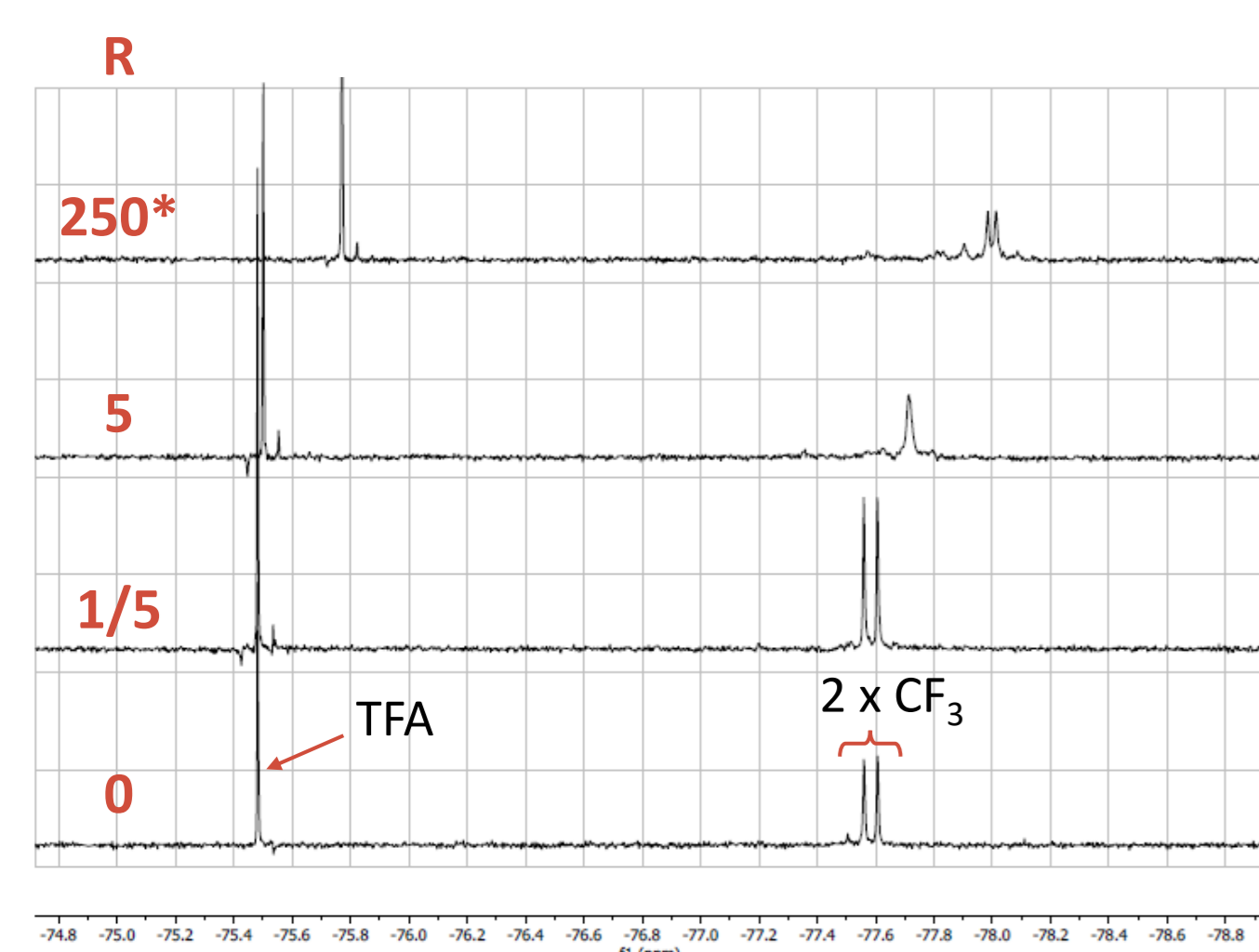


fig. 7: Interaction with SDS micelles monitored by ¹⁹F NMR of **4b** with different concentration of SDS (foldamer as TFA salt at 200 μM in H₂O/D₂O (90/10), *: above CMC, R = [SDS]/[foldamer])

As ¹⁹F is very sensitive to its environment, we assume that the chemical shift displacement is due to **interaction with micelles**. This is confirmed by DOSY ¹H-¹⁹F assay.

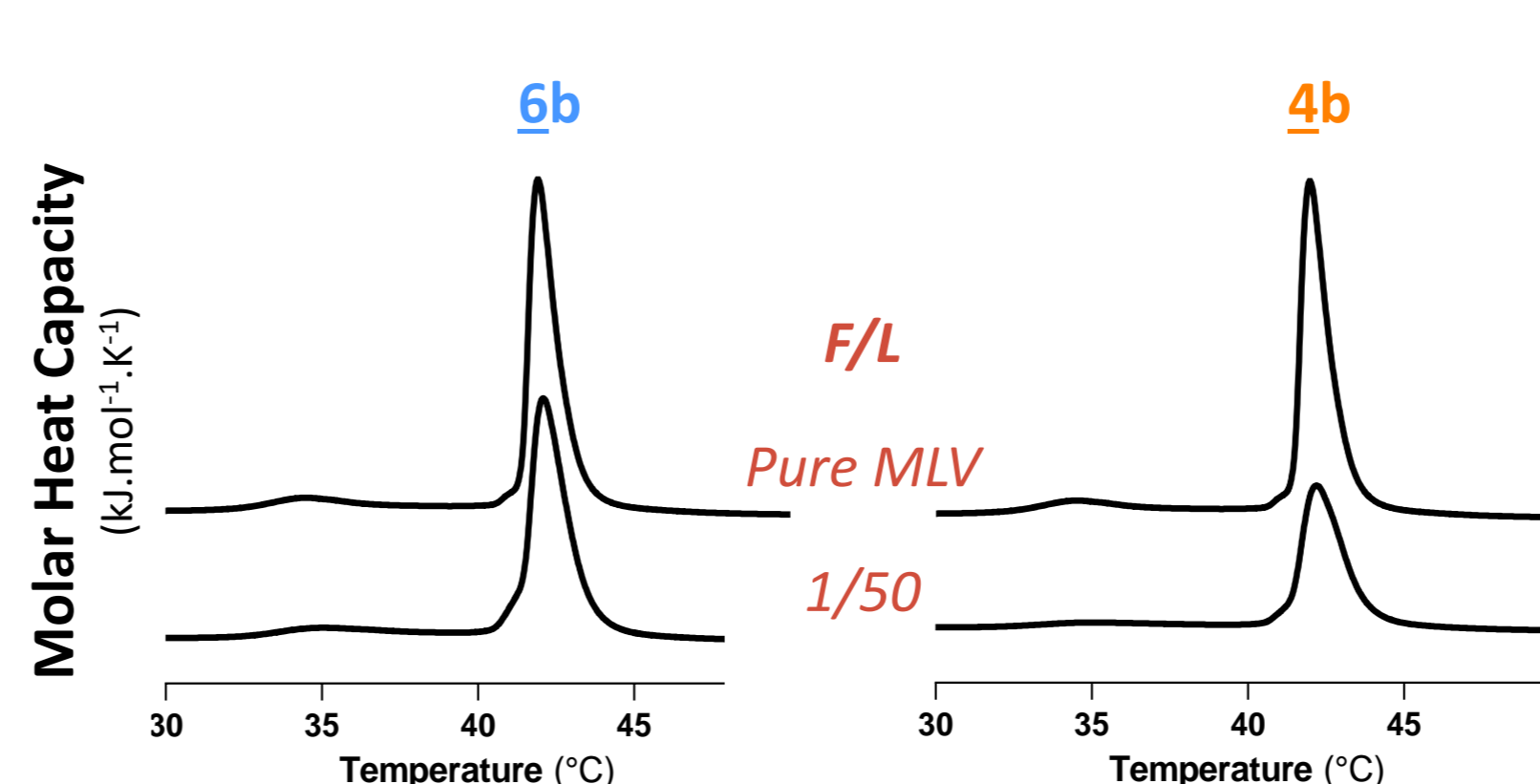


fig. 8: Interaction of hexamers **4b** and **6b** with Multi-Lamellar Vesicles, MLV, of DPPG (anionic lipid) monitored by DSC (1 mg/mL MLV, F/L: molar ratio of foldamers on lipids)

DSC experiments reveal that **foldamers 6b and 4b disrupt MLV thermal transitions**, with the decrease of the area under the curve, providing evidence of an **interaction**. A **more significant effect of the fluorinated foldamer** can be highlighted.

Foldamer **4b** interacts with membrane models, SDS micelles and DPPG MLV

CONCLUSION

- X-ray crystallography, CD and NMR spectroscopies are consistent with **3** adopting a **helical secondary structure close to PPII**. The presence of the **CF₃ group** in δ position of the oxazolidine ring **may cause steric clashes** that can partially (or locally) disrupt PPII helix structure.
- Concerning the amphipathic series **4**, more **experiments** have to be done to **decipher the secondary structure**, between **PPII helix** or **β-sheet**.
- The introduction of cationic groups on our fluorinated oligomers leads to **foldamers able to interact with membrane models**. Moreover, ¹⁹F NMR revealed to be a **useful tool** to investigate these **interactions**.

REFERENCES

- Gellman, S. H. *Acc. Chem. Res.* **1998**, *31*, 173.
- Moradi, M. *et al. J. Chem. Phys.* **2010**, *133*, 125104.
- Chaume, G. *et al. J. Org. Chem.* **2013**, *78*, 10144.
- Hruby, V. J., *et al. J. Org. Chem.* **2001**, *66*, 1038.
- Wennemers, H., *et al. J. Am. Soc.* **2009**, *131*, 15471.
- Hornig, J.-C., *et al. Protein Science.* **2009**, *18*, 1967.

Acknowledgements



The CY Initiative of Excellence (grant « Investissement d'Avenir » INEX 2021 Biomol) is thanked for the financial support of Chloé Cayrou. Part of this work is also supported by the ANR FluFOLD N°ANR-22-CE44-0020-01.

