

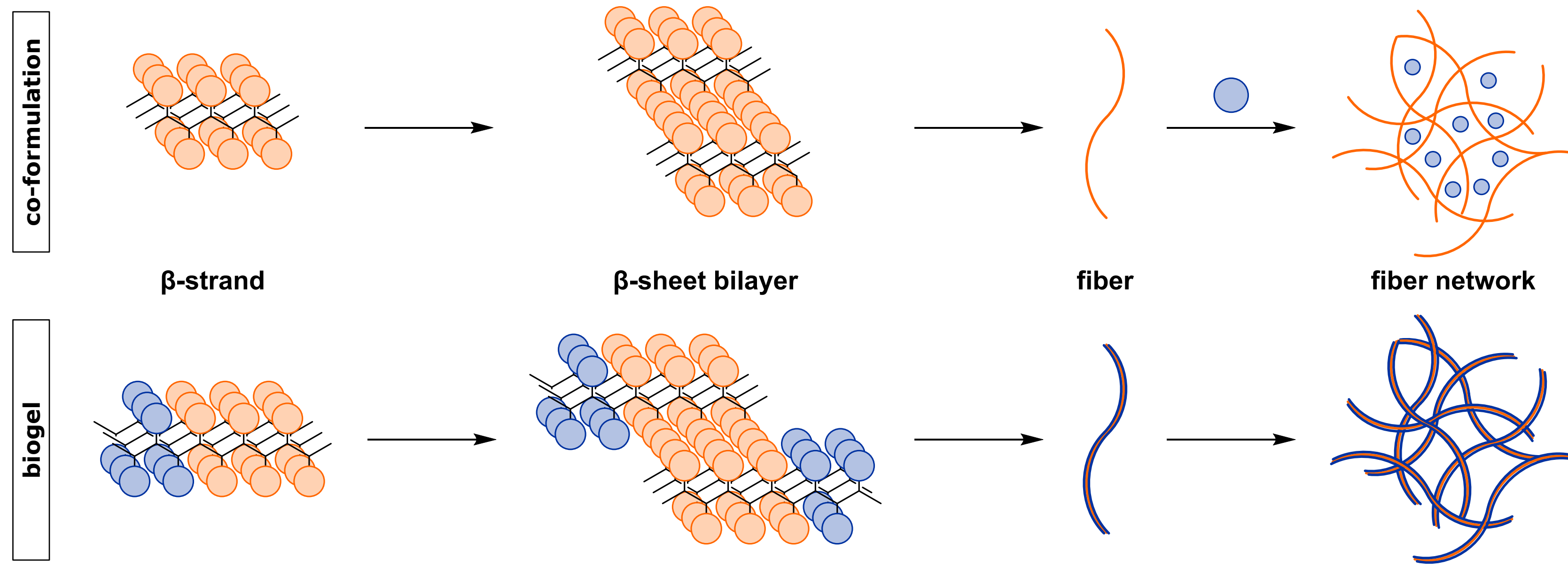
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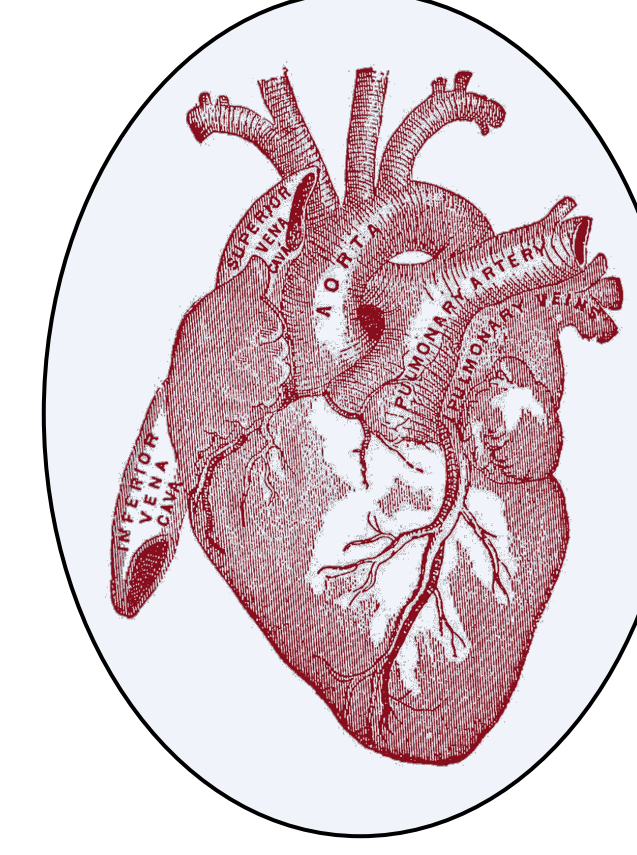
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CO-FORMULATION VS. BIOGEL FORMULATION [1]



Templated on our in-house developed amphipathic hexapeptide hydrogel (H-Phe-Gln-Phe-Gln-Phe-Lys-NH₂), we designed a **self-assembling hydrogel-drug conjugate** platform, termed **biogel**, with a proof-of-concept for the sustained release of opioid peptides. This **prodrug** approach offers an elegant solution to the often difficult-to-control and fast release properties characteristic for traditional co-formulation-based hydrogel platforms. The work presented here focuses on the implementation of the developed biogel concept in **targeting the urotensin-II receptor (UT-IIR)**.

UROTENSIN II RECEPTOR [2,3]



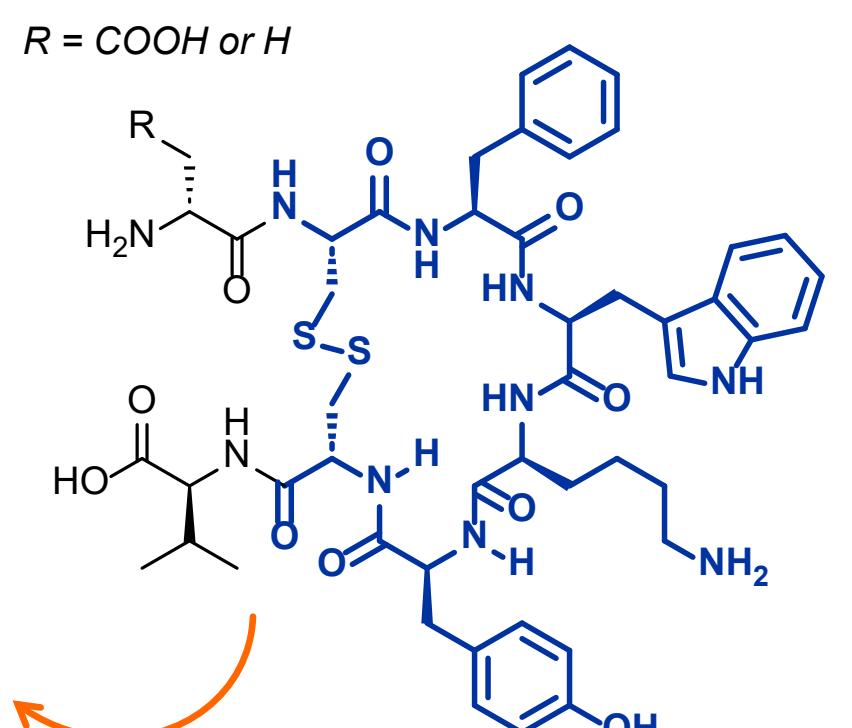
- > G protein-coupled receptor
- > somatostatin receptor homologue
- > expressed in heart & pancreas
- > intracellular Ca²⁺ mobilization
- > strong vasoconstricting effect
- > involved in renal failure & diabetes

hUT-II₍₄₋₁₁₎ & URP

Native agonists
Ultrapotent vasoconstrictors

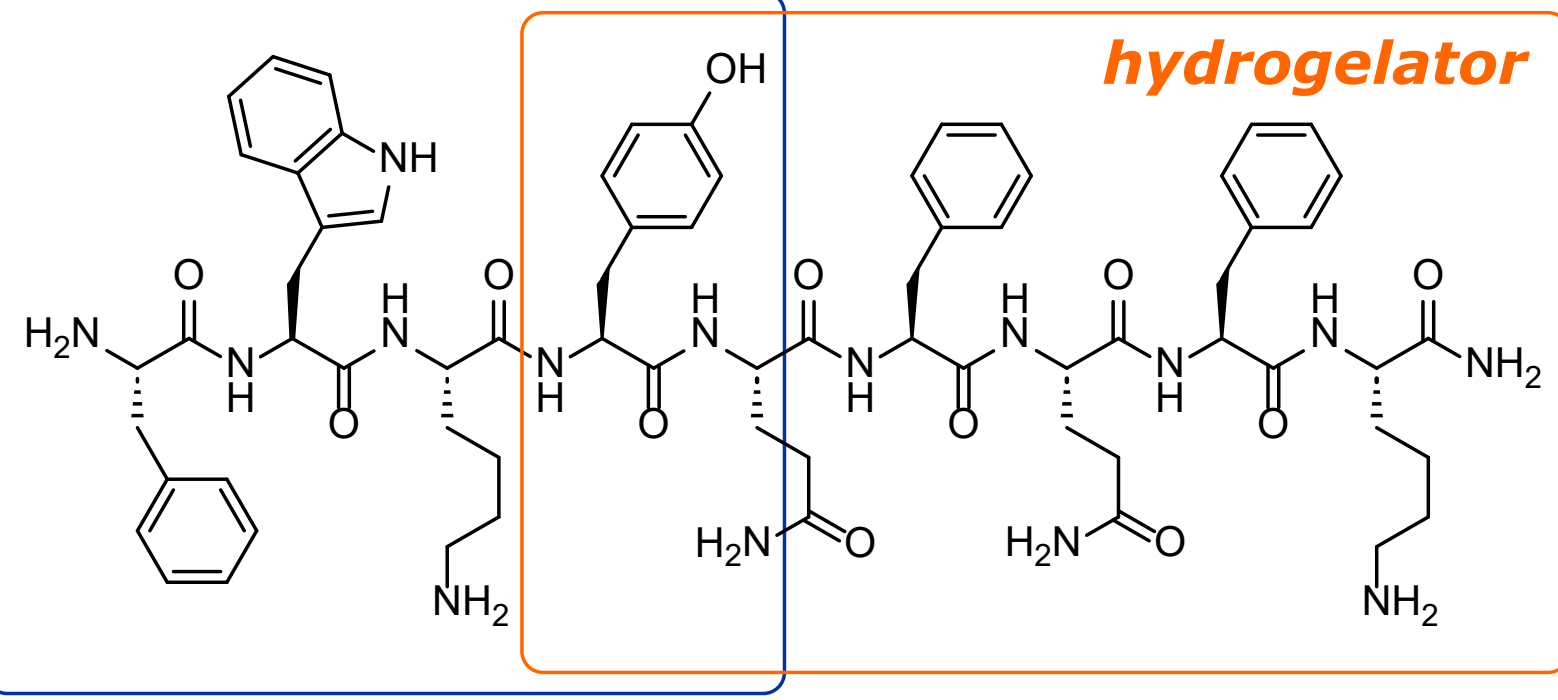
Key ligand features

- ✓ -FWKY- (or closely related) core motif
- ✓ -SS- cyclic hexapeptide
- ✓ Type II' β-turn
- ✓ N- and C-terminal capping is tolerated



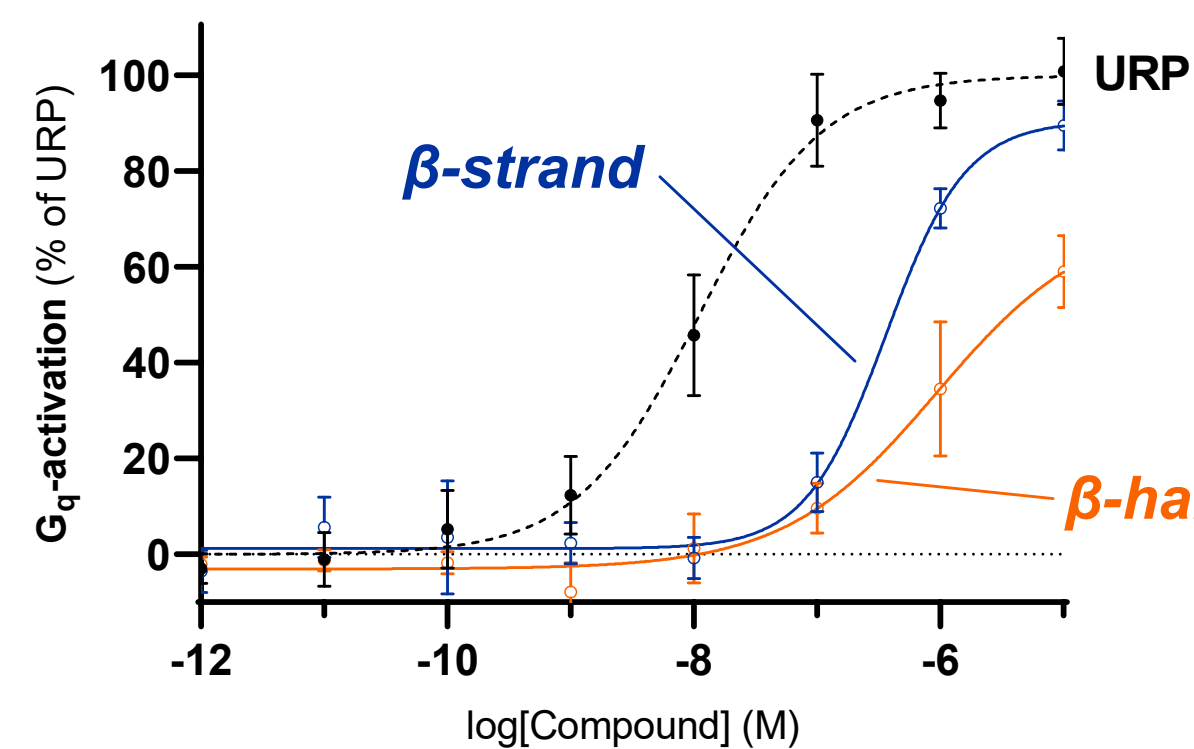
LEAD BIOGEL DESIGN [4]

UT-II pharmacophore



β-strand lead peptide
gel formation @ 2% w/v in PBS

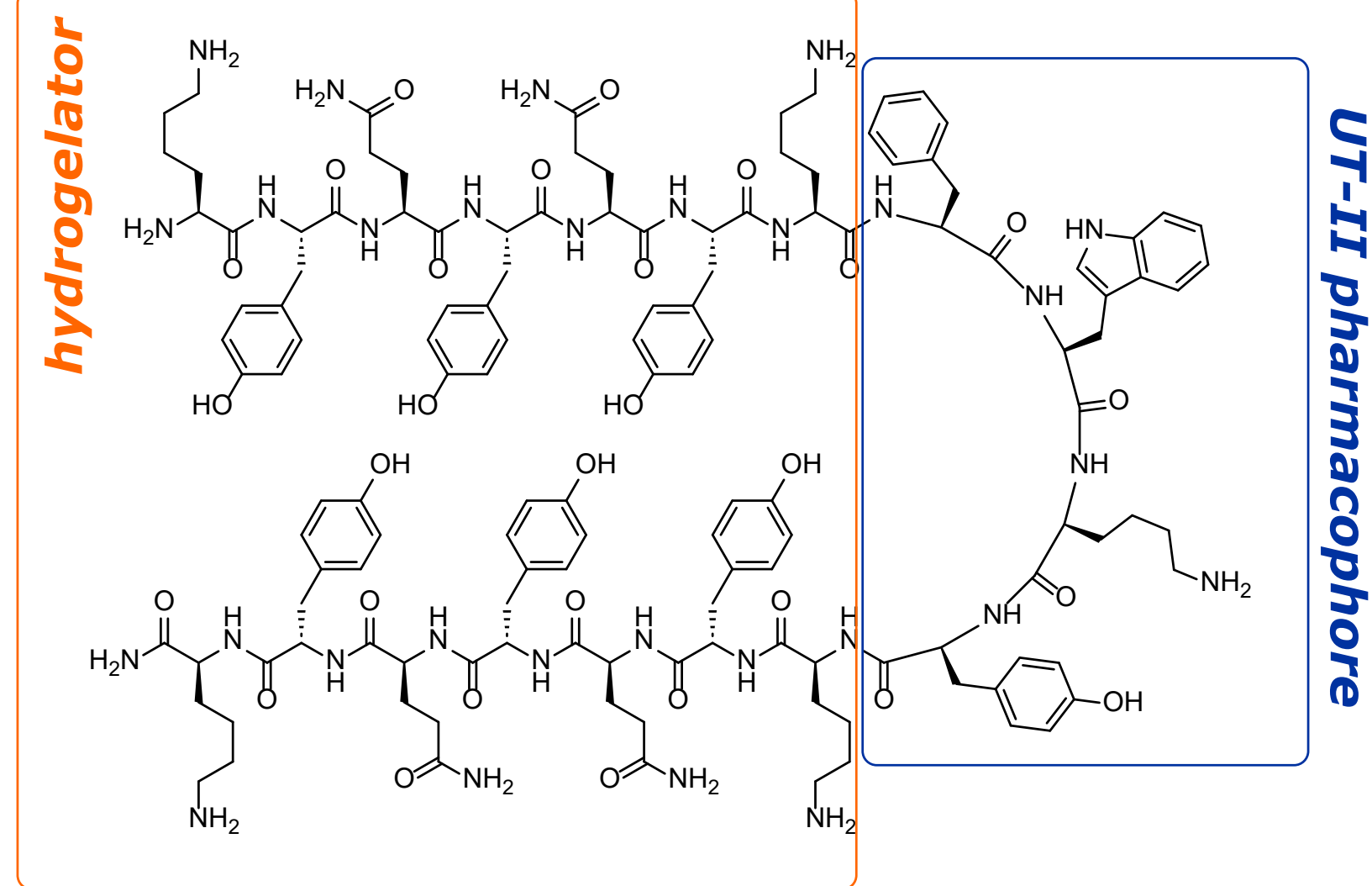
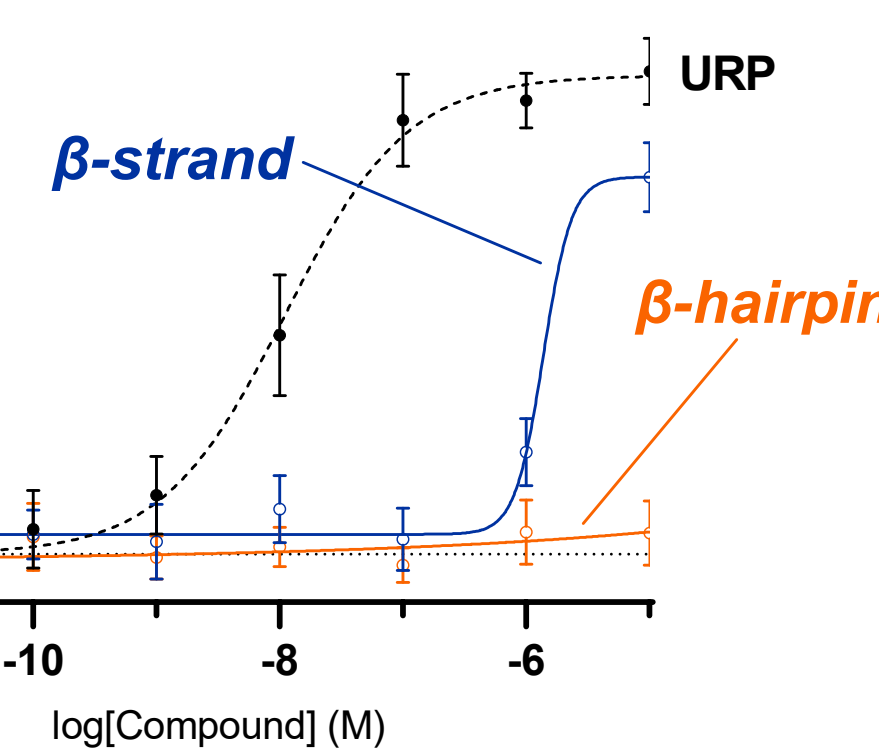
β-STRAND



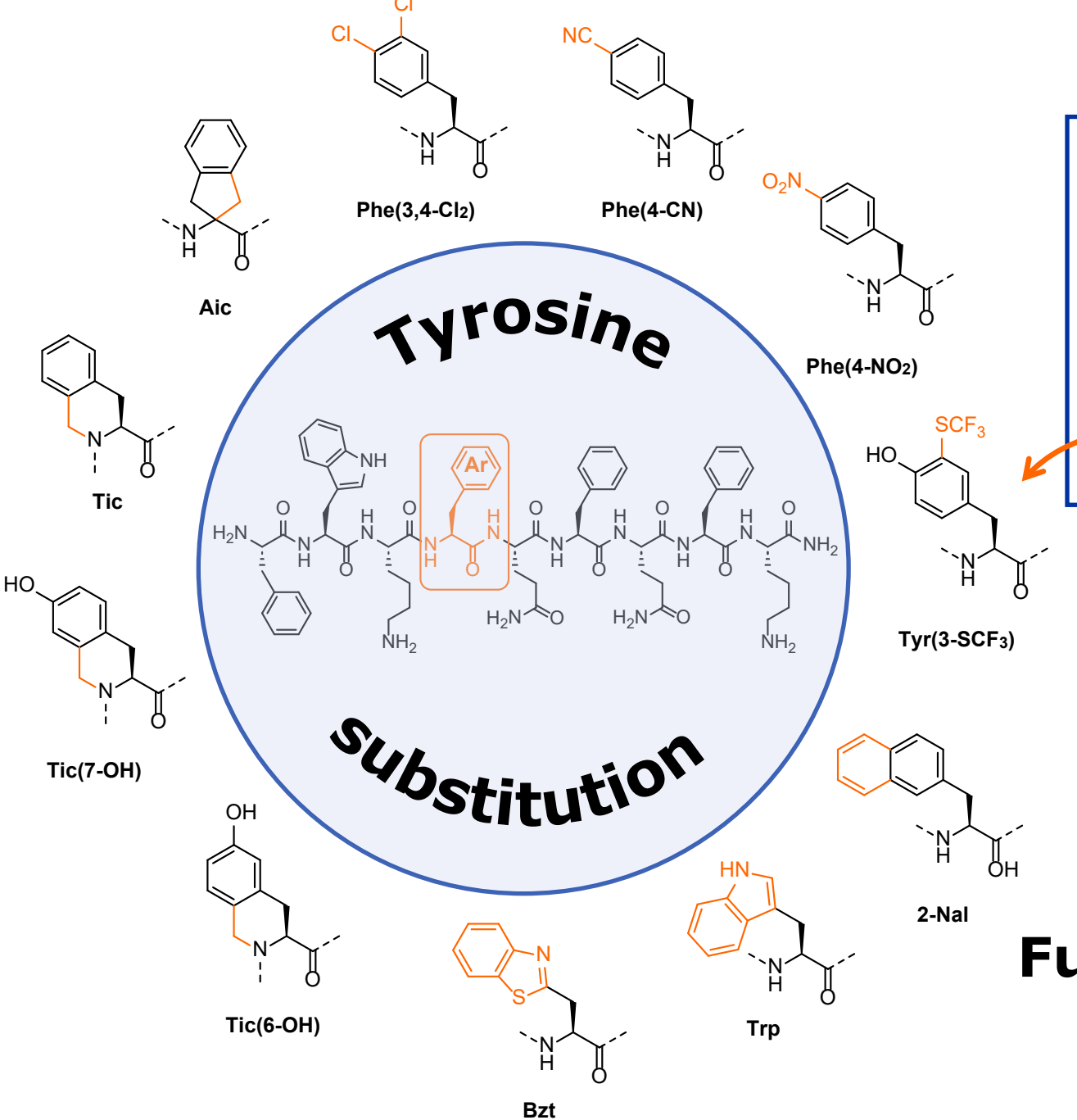
GPCR activation

(BRET biosensor assay on HEK293 cells)

β-HAIRPIN



TYR SAR STUDY [5]



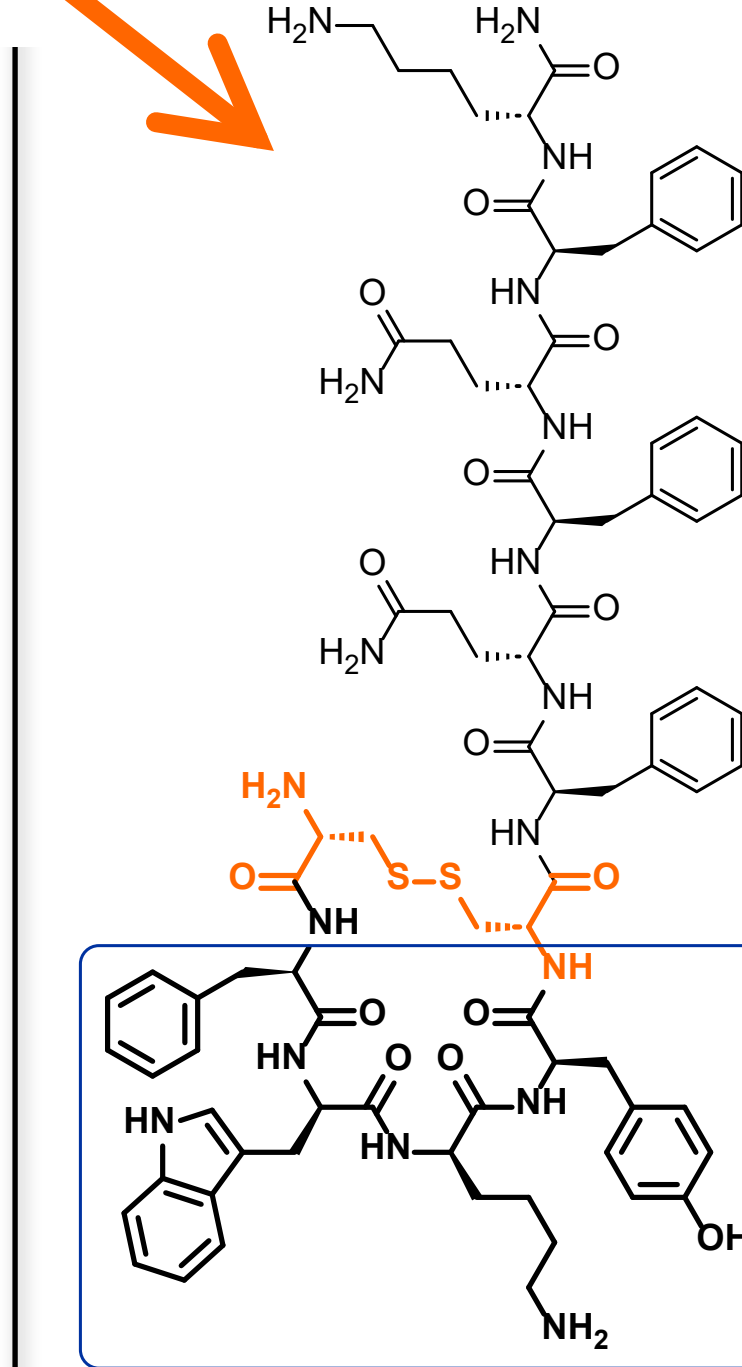
- General considerations**
- ☐ Phe most tolerant residue
 - ☐ Trp tolerant to stereochemical inversion
 - ☐ Lys 3/4 carbon (primary) aliphatic amine!
 - ☑ Tyr aromatic moiety!

Strong gel formation @ 2% w/v in PBS
except for constrained AAs (Tic(x-OH), Aic)

however

Full loss of G_q & β_{arr1} activation observed...
> under investigation

CYCLIZATION [6]

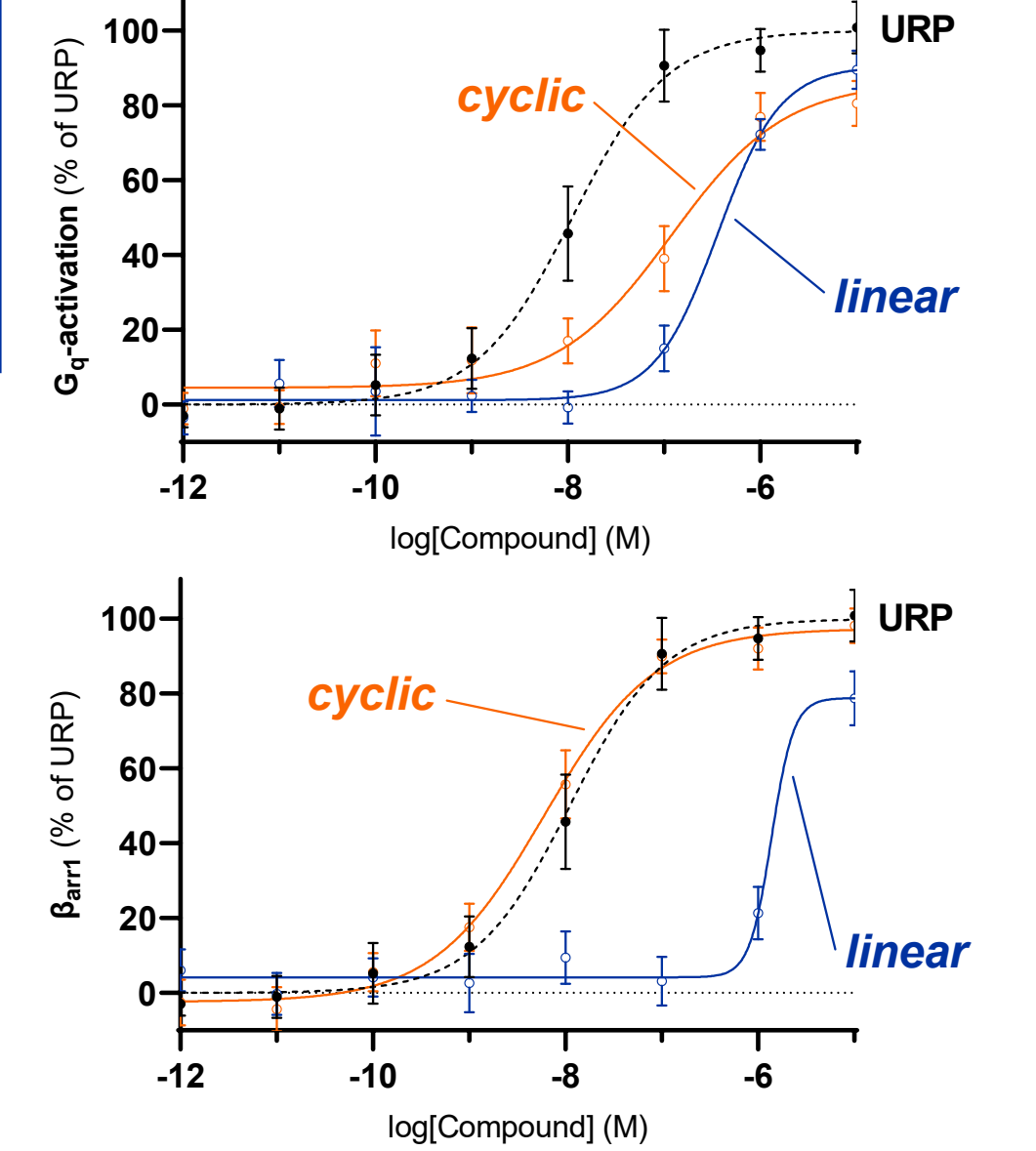


"Disulfide bridge is crucial for receptor recognition and activation"

Slightly reduced gel strength
but
Improved activity!

GPCR activation

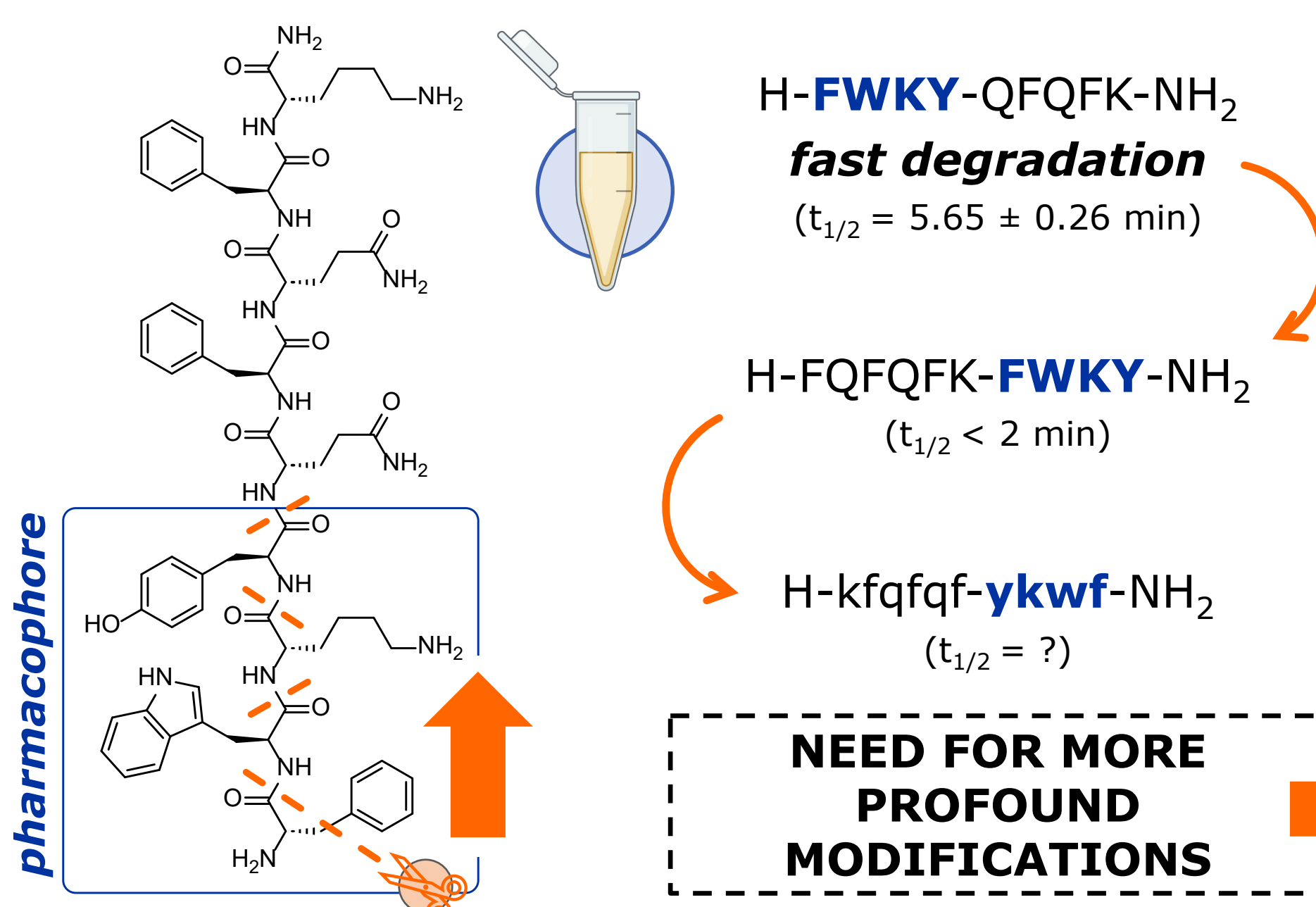
(BRET biosensor assay on HEK293 cells)



cyclization conditions

- on resin
- I₂ (10 equiv.)
- 2% v/v anisole in NMP
- MW
- 40 °C
- 15 min

PROTEOLYTIC RESISTANCE



H-FWKY-QFQFK-NH₂
fast degradation
(t_{1/2} = 5.65 ± 0.26 min)

H-FQFQFK-FWKY-NH₂
(t_{1/2} < 2 min)

H-kfqfqf-ykwf-NH₂
(t_{1/2} = ?)

NEED FOR MORE PROFOUND MODIFICATIONS

N-capping

D-Asn-

t_{1/2} = 93.59 ± 2.67 min
C-ter cleavage
H-nFW-OH metabolite

Ac-

t_{1/2} = 70.64 ± 2.59 min
C-ter cleavage
Ac-FWK-OH metabolite

Gdn-

t_{1/2} = 57.22 ± 3.80 min
C-ter cleavage
Gdn-FWK-OH metabolite

Trp modification

D-Trp

t_{1/2} = 26.87 ± 4.62 min

D-Trp D-hydrogelator

t_{1/2} > 24 h

D-Trp (N-Me)Gln

t_{1/2} = 30.74 ± 2.06 min

(N-Me)Trp

t_{1/2} = 45.16 ± 0.90 min

Tcc

t_{1/2} = 47.56 ± 2.28 min

C-ter cleavage

Tcc preserves pharmacophore
substantial impact on gelation

Cyclization

R = H (Cys)

t_{1/2} = 122.16 ± 2.63 min

R = Me (Pen)

t_{1/2} = 53.36 ± 1.51 min

pharmacophore preserved up to 24 h

CONCLUSIONS

- > Search for highly active, long-acting UT-II-targeting conjugate hydrogel ('biogel')
- > Cyclization of the pharmacophore enhances activity and proteolytic stability
- > Unexpectedly, Tyr substitution yielded inactive linear peptide derivatives
- > Proteolytic stabilization is needed to prolong peptide activity and preserve the pharmacophore

- > Increase peptide activity, to compete with the parent peptide
- > Test *in vitro* activity of the stabilized biogel analogues and their metabolites
- > Further study & optimize the β-hairpin design
- > Expand the biogel platform to other targets

PERSPECTIVES

ACKNOWLEDGEMENTS

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